THE CONTRIBUTION OF FINE NEEDLE ASPIRATION BIOPSY IN
THE DIAGNOSIS OF MYCOBACTERIAL LYMPHADENOPATHY
WITH PARTICULAR REFERENCE TO CHILDREN

Dissertation presented by

COLLEEN ANNE WRIGHT

For a PhD degree in Anatomical Pathology

at

Stellenbosch University
August 2009

Promoter: Prof R.M.Warren

Co-promoter: Prof B.J.Marais
DECLARATION

I, the undersigned, hereby declare that the work contained in this dissertation is my own original work and that I have not previously in its entirety or in part submitted it at any other university for a degree.

Signature:

Date: 5 August 2009

In all first author papers the candidate obtained ethics consent, assisted with the sample collection, collected the data, drew up the analysis plan and wrote the first draft of the manuscript.

In addition the candidate was involved in all the microscopic evaluations and learnt the laboratory techniques used in the molecular analyses.

In all the other papers the candidate helped conceptualise the study, assisted with sample collection and evaluation of microscopic slides and made a substantial contribution to the manuscript.
SUMMARY

Expediting a diagnosis of tuberculosis in children, particularly those who are immunocompromised due to HIV/AIDS, is essential, as they are vulnerable to develop severe forms of disease due to their immature or compromised immune systems. A significant percentage of children (8 to 10%) with TB have TB lymphadenitis, in isolation, or in combination with other disease manifestations.

Fine needle aspiration biopsy (FNAB) is a simple and minimally invasive procedure well tolerated by children. It may be performed as an outpatient procedure by clinicians as well as nurses, and excellent results can be achieved with training in the correct procedure.

The aim of this dissertation was to demonstrate that FNAB may contribute significantly to the diagnosis of mycobacterial lymphadenitis, with particular reference to children TB suspects. We first established that TB lymphadenitis is a common clinical problem in children in TB endemic areas and that FNAB is an efficient simple and effective diagnostic modality in children with peripheral lymphadenopathy.

We then proceeded to document the diagnostic yield and time to diagnosis of FNAB compared to conventional laboratory specimens collected in children.

We investigated the value of additional diagnostic modalities such as autofluorescence in improving the ability of cytology to make a definitive diagnosis of mycobacterial infection based on cytomorphology and identification of the organism.

In countries where organisms such as Mycobacterium bovis BCG and non-tuberculous mycobacteria are prevalent, culture with subsequent speciation is essential. The amount of material harvested during FNAB is minuscule, and requires immediate bedside inoculation for optimal yields.

We developed an inexpensive and effective transport medium to facilitate mycobacterial culture from FNAB, even if this is collected at an outside facility. It is ideally
suited for use in clinics and rural hospitals as it is stable at room temperature, maintains viability of the organism for seven days, and the closed lid format reduces contamination.

Mycobacterial culture even using liquid-based media, takes up to 6 weeks, and this delay is unacceptable particularly in children. We developed a Nucleic Acid Amplification Technique (NAAT) using High Resolution Melt Analysis and applied this novel technique to FNAB specimens submitted in transport medium. Although sensitivity remained suboptimal, the technique is highly specific, simple and rapid. Its use could be incorporated into routine microbiology laboratories, to assist with rapid diagnosis while cultures are pending.

We collected a solid body of evidence, which will promote the use of FNAB in suspected mycobacterial lymphadenopathy, particularly in children in resource-limited countries. The utilisation of the diagnostic methods identified will expedite speciation and allow early and appropriate initiation of therapy. This is in keeping with Millennium Development Goal 6: to combat TB by early detection of new cases and effective treatment.
OPSOMMING

Kinders met tuberkulose (TB), en veral diegene met gekompromiteerde immuniteit as gevolg van MIV/VIGS, het ‘n verhoogde neiging om ernstige siektebeelde te ontwikkel vanweë hul onvolwasse of gekompromiteerde immuunsisteme. ‘n Spoedige diagnose van TB in kinders is dus noodsaaklik. ‘n Betekenisvolle persentasie van kinders (8 tot 10%) met TB het TB limfadenitis met of sonder meegaande ander siekteverskynsels.

Fynnaaldaspirasiebiopsie (FNAB) is ‘n eenvoudige en minimale indringende prosedure wat geredelik deur kinders aanvaar word. Geneeshere en verpleegkundiges wie toepaslike opleiding in die uitvoering van FNAB ontvang het, kan die prosedure op buitepasiënte uitvoer en uitstekende resultate behaal.

Die doel van hierdie studie was om aan te toon dat FNAB betekenisvol kan bydra tot die diagnose van mikobakteriële limfadenitis in veral kinders met vermoedelike TB. Daar was eerstens bevestig dat TB limfadenitis ‘n algemene kliniese probleem is in kinders in TB endemiese areas en dat FNAB ‘n doeltreffende, eenvoudige en effektiewe diagnostiese modaliteit is in kinders met perifere limfadenopatie.

Vervolgens was FNAB se diagnostiese opbrengs en die tydsverloop tot diagnose vergelyk met dié van konvensionele laboratoriummonsters wat in kinders verkry word.

Die bydrae van verdere diagnostiese modaliteite soos outofluoressensie tot ‘n verbetering in sitologie se rol in die diagnose van mikobakteriële infeksie, soos gebaseer op sitomorfologie en identifisering van organismes, is ondersoek.

In lande waar organismes soos Mycobacterium bovis BCG en nie-tuberkuleuse mikobakterië heersend is, is kultuur en spesiebepaling noodsaaklik. Die hoeveelheid materiaal wat met FNAB verkry word is baie min en vereis onmiddellike okulasie vir die beste resultate.
Tydens hierdie studie is 'n goedkoop en effektiewe vervoermedium ontwikkel om mikobakteriële kultuur van FNAB verkreë monsters te fasileer, selfs al is die monster vanaf 'n buite fasilitéite bekom. Die vervoermedium is baie geskik vir gebruik in klinieke en plattelandse hospitale. Dit is stabiel by kamertemperatuur, handhaaf lewensvatbaarheid van organismes vir sewe dae, en die geslote dekselformaat verminder kontaminasie.

Mikobakteriële kultuur neem tot ses weke, selfs met die gebruik van vloeistofgebaseerde mediums. Sodanige vertraging in die diagnose is veral in kinders onaanvaarbaar. Tydens hierdie studie is 'n Nukleïnsuur Amplifikasietegniek ontwikkel deur die aanwending van Hoë Resolusie Smeltanalise en is hierdie nuwe tegniek toegepas op FNAB verkreë monsters wat in die vermelde vervoermedium versamel was. Alhoewel sensitiwiteit nie optimaal was nie, is die tegniek baie spesifiek, eenvoudig en vinnig. Dit kan in roetine mikrobiologie laboratoriums gebruik word om vinnige diagnose te bewerkstellig terwyl daar gewag word vir die kultuur se resultaat.

Hierdie studie bied omvattende bewys ter ondersteuning van die gebruik van FNAB in veral kinders met vermoelde mikobakteriële limfadenopatie in lande met beperkte hulpbronne. Die toepassing van die diagnostiese metodes wat in hierdie studie identifiseer is sal spesiebepaling bespiedig en vroegtydige en toepaslike behandeling verseker. Dit stem ooreen met Millennium Ontwikkelingsdoelwit 6: om TB te beveg deur vroeë opsporing van nuwe gevalle en effektiewe behandeling.
ACKNOWLEDGEMENTS

This mortal world is fickle and unstable like unto a shifting shadow and the human life is like
unto a mirage and a reflection on the water. (Abdu'l-Baha)

Our lives, and what we achieve, are but a reflection of the lives around us, therefore

I would like to thank

All those who have assisted either directly or indirectly in the completion of these studies and this project. If you are not mentioned by name in the list below forgive me, I will remain indebted to you all. I hope the envisaged improved diagnosis and outcome for children with tuberculosis will be a testament to this combined effort.

Prof Ben Marais and Prof Rob Warren, my supervisors, for their encouragement, insight and guidance throughout the course of this dissertation. For giving so generously of their time and often prioritizing my research needs above their own commitments.

Kim Hoek for the hours spent optimizing the molecular techniques in this study. I will always be grateful to you for your assistance and your kindness.

Prof Gladwyn Leiman who started me on the long quest to establish Fine Needle Aspiration Biopsy as an invaluable diagnostic tool, and whose legacy in gaining acceptance of this modality in South Africa will live on forever.

Prof Paul van Helden, for allowing me access to expertise and infrastructure within his department, and for his encouragement.
Prof Johann Schneider, Prof Juanita Bezuidenhout and Dr Mercia Louw, who took over many of my duties and freed me to complete this work.

Prof Peter Wranz, for his love for cytology, unfailing encouragement and support.

All the registrars in Anatomical Pathology, present and past, who uncomplainingly obtained consent from patients and collected samples for the studies. I cannot sufficiently express my appreciation for your participation.

All the staff in Anatomical Pathology who tolerated my absence during the periods that I spent committed to completing the study.

Prof Martin Kidd for his statistical analysis, assistance and patience.

The National Health Laboratory Service, for providing me with the environment and infrastructure to conduct this research. I sincerely hope the mechanisms and diagnostic techniques that have evolved and which we envisage will be implemented in our laboratory services contribute to patient care.

The Faculty of Health Sciences, for their support, particularly Prof Wynand van der Merwe and Prof Usuf Chikte, who always believed I could do it. Thank you.
FUNDING

This project was made possible by the following generous contributions:

The establishment fund of the Dean of the Faculty of Health Sciences.

The National Health Laboratory Systems Trust fund.

Staff development bursary.

Division of Molecular Biology and Human Genetics, Stellenbosch University.
DEDICATION

This work is dedicated to my family, without whose support and understanding this would not have been completed.

Ariel, my son, who has survived a lifetime of a mother as a student and of whom I am so proud.

Thandiwe, my daughter, beloved child of the light, who brings such joy into my life.

Alethe, Richard and Jonathan, may your lives be blessed and protected and

Steve, my husband, partner and friend.

You will always be the centre of my world.

And to the children of Africa who continue to bear the burden of this disease

May you say of us, that we tried.....

*There can be no keener revelation of a society’s soul than the way in which it treats its children.*

Nelson Mandela
**ABBREVIATIONS**

<table>
<thead>
<tr>
<th>Abbreviation</th>
<th>Full Form</th>
</tr>
</thead>
<tbody>
<tr>
<td>AIDS</td>
<td>Acquired Immune Deficiency Syndrome</td>
</tr>
<tr>
<td>BCG</td>
<td>Bacillus Calmette-Guerin</td>
</tr>
<tr>
<td>CDC</td>
<td>Centers for Disease Control and Prevention</td>
</tr>
<tr>
<td>CI</td>
<td>Confidence interval</td>
</tr>
<tr>
<td>DOTS</td>
<td>Directly observed therapy, short-course</td>
</tr>
<tr>
<td>ELISA</td>
<td>Enzyme-linked immunosorbent assay</td>
</tr>
<tr>
<td>FNAB</td>
<td>Fine Needle Aspiration Biopsy</td>
</tr>
<tr>
<td>HAART</td>
<td>highly active anti-retroviral therapy</td>
</tr>
<tr>
<td>HIV</td>
<td>Human Immunodeficiency virus</td>
</tr>
<tr>
<td>HRM</td>
<td>High Resolution Melting</td>
</tr>
<tr>
<td>IRIS</td>
<td>Immune reconstitution syndrome</td>
</tr>
<tr>
<td>LED</td>
<td>Light emitting diode</td>
</tr>
<tr>
<td>LIP</td>
<td>Lymphocytic interstitial pneumonitis</td>
</tr>
<tr>
<td>MDR</td>
<td>Multi drug resistant</td>
</tr>
<tr>
<td>MGIT</td>
<td>Mycobacterial growth indicator tube</td>
</tr>
<tr>
<td>MODS</td>
<td>Microscopic observation drug susceptibility assay</td>
</tr>
<tr>
<td><strong>M.tuberculosis</strong></td>
<td>Mycobacterium tuberculosis</td>
</tr>
<tr>
<td><strong>M.bovis-BCG</strong></td>
<td>Mycobacterium bovis BCG</td>
</tr>
<tr>
<td>MVP</td>
<td>Mercury vapor lamp</td>
</tr>
<tr>
<td>NAAT</td>
<td>Nucleic acid amplification tests</td>
</tr>
<tr>
<td>NHLS</td>
<td>National Health Laboratory Services</td>
</tr>
<tr>
<td>NTM</td>
<td>Non - tuberculous mycobacteria</td>
</tr>
<tr>
<td>OR</td>
<td>Odds ratio</td>
</tr>
<tr>
<td>PBS</td>
<td>Phosphate buffered saline</td>
</tr>
<tr>
<td>PCR</td>
<td>Polymerase chain reaction</td>
</tr>
<tr>
<td>PPL</td>
<td>Persistent peripheral lymphadenopathy</td>
</tr>
<tr>
<td>SA</td>
<td>South Africa</td>
</tr>
<tr>
<td>TB (PTB)</td>
<td>Tuberculosis (Pulmonary tuberculosis)</td>
</tr>
<tr>
<td>TBH</td>
<td>Tygerberg Hospital</td>
</tr>
<tr>
<td>TST</td>
<td>Tuberculin skin test</td>
</tr>
<tr>
<td>WHO</td>
<td>World Health Organization</td>
</tr>
<tr>
<td>ZN</td>
<td>Ziehl-Neelson</td>
</tr>
<tr>
<td>XDR</td>
<td>Extensively drug resistant</td>
</tr>
</tbody>
</table>
TABLE OF CONTENTS

Chapter 1.  Introduction 1

Chapter 2.  Comprehensive literature review 7
   Fine needle aspiration Biopsy - an undervalued diagnostic modality in pediatric mycobacterium disease. 8

Chapter 3.  TB lymphadenitis a common clinical problem 33
   Tuberculous Lymphadenitis as a Cause of Persistent Cervical Lymphadenopathy in Children from a Tuberculosis-Endemic Area. 34

Chapter 4.  Diagnostic utility and feasibility of fine needle aspiration biopsy 39
   Fine needle aspiration biopsy: diagnostic utility in resource-limited settings. 40
   Fine Needle Aspiration Biopsy – a first line diagnostic procedure in pediatric tuberculosis suspects with peripheral lymphadenopathy. 46

Chapter 5.  Optimizing cytopathological diagnosis 66
   Auto fluorescence of Mycobacteria on Lymph Node Aspirates – A glimmer in the dark. 67
   Diagnosing Mycobacterial lymphadenitis in children using Fine Needle Aspiration Biopsy: Cytomorphology, ZN staining and Autofluorescence – making more of less. 71

Chapter 6.  Optimizing culture-based and molecular diagnosis 78
   Mycobacterial Transport medium for routine culture of Fine Needle Aspiration Biopsies 79
   Combining Fine Needle Aspiration Biopsy and High Resolution Melt Analysis to reduce diagnostic delay in Mycobacterial Lymphadenitis. 89
CHAPTER 1

INTRODUCTION

Tuberculosis (TB) was declared a global emergency in 1993 by the World Health Organisation (WHO) when it was estimated that a third of the world's population is infected with TB.\(^1\) The developing world bears the brunt of this disease burden; 95% of TB cases and 98% of TB deaths occur in the developing world, with sub-Saharan Africa reporting the highest TB incidence in the world.\(^2\) In 2005 there were an estimated 8.8 million new TB cases globally and 7.2 million of these were in Asia and sub-Saharan Africa. Despite efforts to contain the epidemic new TB cases increased by 6% between 2000 and 2005, predominantly in sub-Saharan Africa and Asia.

The problem is compounded by the parallel human immunodeficiency virus (HIV) pandemic with 33.2 million adults and children living with HIV in 2007, and 68% of these in sub-Saharan Africa.\(^3\) Infection with the HIV virus is estimated to increase the lifetime risk of a patient infected with *M tuberculosis* of developing TB by 10 fold to 50%.\(^4\) South Africa with 5.5 million people leads the world in the number of patients living with HIV, as well as TB/HIV co-infection.\(^5\)

**TB in childhood**

TB is a major cause of childhood morbidity and mortality in developing countries.\(^6,7\) It is estimated that 11% of the 8.3 million new TB cases diagnosed globally in 2000 occurred in children,\(^8\) and in endemic areas children contribute at least 15-20% of the total disease burden.\(^9\) In the Western Cape Province, a high burden region in South Africa, the TB notification rate in children less than 14 years of age was reported at 620 /100,000 in 2007.\(^10\)

Official child TB figures almost certainly represent a gross underestimate due to problems in obtaining an accurate diagnosis in resource-limited settings, poor record-keeping and under reporting.\(^8\) Children rarely have sputum smear-positive TB, although this reduces
the risk of actively spreading the disease, it makes it more challenging to establish a definitive diagnosis. Expectorated sputum specimens are difficult if not impossible to obtain in children under the age of 7-8 years. Collection of induced sputum and gastric aspirate specimens are possible, but difficult to collect and reported bacteriological yields are low (30-40%).\textsuperscript{8,11}

A clinical diagnosis of TB in children is problematical and although chest radiography is regarded as the most practical test to provide a reasonably accurate diagnosis of intrathoracic TB in children with suspicious symptoms, it has multiple limitations.\textsuperscript{12} In HIV-infected children the diagnosis of both intrathoracic and extra thoracic TB is difficult due to underlying chronic pathology such as lymphocytic interstitial pneumonitis (LIP) or bronchiectasis and opportunistic infections. Chest radiography can also not provide diagnostic confirmation in children with extra thoracic TB. In a study in the Western Cape, South Africa 10.7\% of children treated for active TB had extra thoracic disease manifestations only; cervical TB lymphadenitis was the most common condition in these children being present in 48.6\%. TB lymphadenitis is a common cause of persistent cervical adenopathy in TB endemic areas. Persistent peripheral lymphadenopathy in HIV-infected children is a criterion for the clinical classification of HIV itself. It may also be associated with multiple other pathological entities, from infections to neoplasia.

\textit{Mycobacterium tuberculosis} is not the only mycobacterial organism which involves peripheral lymph nodes. In developed countries non-tuberculous mycobacteria (NT M) is the commonest mycobacterial organism isolated and BCG vaccination may be associated with local complications such as abscess formation at the vaccination site and regional lymphadenitis. It is also the most common manifestation of immune reconstitution syndrome (IRIS) if vaccinated HIV-infected infants are commenced on highly active anti-retroviral therapy (HAART). It is therefore important to develop methods that will provide a rapid and definitive mycobacterial diagnosis in children with persistent peripheral lymphadenopathy.
Fine needle aspiration biopsy (FNAB) is a specimen collection technique which is widely used in the diagnosis of palpable masses including peripheral lymphadenopathy. Its value in the diagnosis of mycobacterial lymphadenitis in adults is well documented. FNAB is a simple effective and safe modality for obtaining a representative sample of material from a lymph node and the diagnosis of mycobacterial adenitis can be confirmed utilising a number of different investigations, including cytomorphology, specific stains to identify the organism, culture and polymerase chain reaction (PCR).

FNAB is not widely used for the diagnosis of mycobacterial disease in children. As this is a rapid and minimally invasive technique which can be performed on an outpatient basis, in a primary health care setting it is ideal for use in resource-limited countries with the highest incidence of mycobacterial disease as well as HIV infection.

**Study objectives**

The main study objective was to evaluate and develop laboratory-based diagnostic techniques to establish a rapid, accurate and cost effective diagnosis of mycobacterial lymphadenitis in children, using FNAB as the sampling method. We applied a stepwise approach using multiple studies with specific aims to achieve this objective.

**To establish that TB lymphadenitis is a common clinical problem in children from TB endemic areas**

This aim is addressed in chapter 3:

A prospective community-based study documented the contribution of TB lymphadenitis to persistent lymphadenopathy among children who present to primary health care clinics with persistent cervical adenopathy in a TB endemic area.

**To evaluate the diagnostic utility and feasibility of FNAB in a resource limited setting**
These aims are addressed in chapter 4:

A laboratory-based retrospective study documented FNAB adequacy rates among various health care professionals who performed the procedure and described the spectrum of pathology seen.

A laboratory-based retrospective study compared FNAB to more established specimen collection.

A laboratory-based retrospective study evaluated the utility of FNAB in HIV-infected children (supplementary manuscript).

**To determine optimal methods for cytopathological diagnosis of mycobacterial disease**

These aims are addressed in chapter 5 and 7:

A prospective laboratory-based study conducted in adults and children to determine the value of incorporating autofluorescence into cytopathological evaluation of lymph node aspirates in patients with suspected mycobacterial disease.

A prospective laboratory-based study in children assessing the ability of FNAB to diagnose mycobacterial lymphadenitis in children, using cytomorphology, autofluorescence on Papanicolaou stained smears, Ziehl-Nielsen (ZN) staining and/or culture.

A retrospective laboratory-based study to assess cytomorphological patterns of *M. bovis BCG* and *M. tuberculosis* on FNAB (supplementary manuscript).

A retrospective laboratory-based study to evaluate fluorescence microscopy in the diagnosis of mycobacterial infection using FNAB samples and a rewind able (power independent) LED light source (supplementary abstract).

**To explore novel methods for optimal culture-based and molecular diagnosis**

These aims are addressed in chapters 6 and 7:
A prospective pilot study, collecting residual material from fine needle aspiration biopsy in 200 µL of sterile water, to investigate the feasibility of utilising PCR amplification techniques with fractionation in agarose gel and visualisation using ethidium bromide directly from FNAB samples (supplementary study).

A prospective hospital-based study to assess the value of using a simple mycobacterial transport medium for bedside inoculation of FNAB specimens.

A prospective hospital-based study utilizing FNAB combined with High Resolution Melt Analysis to reduce diagnostic delay in patients mycobacterial adenitis.

A simplified technique using FNAB and collection of residual material on FTA paper and real-time PCR/HRM for the diagnosis of mycobacterial infection (supplementary manuscript in progress).

References


CHAPTER 2

COMPREHENSIVE LITERATURE REVIEW

Fine needle aspiration Biopsy (FNAB) - an undervalued diagnostic modality in pediatric mycobacterial disease
Colleen A Wright, Rob Warren, Ben J Marais
International Journal of Tuberculosis and Lung Disease – in press
Fine needle aspiration biopsy (FNAB) - an undervalued diagnostic modality in pediatric mycobacterial disease

1Colleen A Wright, 2 Rob Warren, 3Ben J Marais

1Division of Anatomical Pathology, Department of Pathology, Stellenbosch University and NHLS

2 NRF Centre of Excellence in Biomedical Tuberculosis Research / MRC Centre for Molecular and Cellular Biology, Division of Molecular Biology and Human Genetics, Stellenbosch University, South Africa

3Department of Pediatrics and Child Health and Desmond Tutu TB Centre, Stellenbosch University, South Africa

Unstructured summary

Mycobacterial disease, particularly tuberculosis, is an escalating problem in developing countries, fuelled by the parallel HIV pandemic. In tuberculosis (TB) endemic countries children carry a very high burden of disease, which may be unrecognised due to the difficulty in making a diagnosis based on clinical, radiological or laboratory methods. One of the main hurdles is the difficulty of obtaining adequate specimens for bacteriological confirmation of disease in children.

TB lymphadenitis is the most common extra-pulmonary manifestation of TB and up to 22% of children with persistent cervical lymphadenopathy and no local cause may have TB adenitis Therefore fine needle aspiration biopsy (FNAB), which is a simple and safe outpatient procedure that can be performed by nurses in resource-limited settings and provides material for direct microscopy as well as culture and susceptibility testing, provides an excellent opportunity to obtain bacteriologic confirmation. However, it remains a greatly underutilized specimen collection modality.

This review provides a comprehensive overview of the diagnostic difficulties faced in the diagnosis of paediatric TB in resource-limited settings and suggests ways to utilise FNAB
as a practical modality for the rapid and effective diagnosis of mycobacterial disease in the significant subset of patients who present with peripheral lymphadenopathy. It also provides detail on how best to perform the technique and suggests ways of making it more widely available in resource-limited settings that carry the brunt of the paediatric TB disease burden.

The scale of the problem

*Mycobacterium tuberculosis*

In 2005, there were an estimated 8.8 million new tuberculosis (TB) cases globally, 7.4 million in Asia and sub-Saharan Africa. More than 4000 people died every day from TB related illnesses in that year, with the developing world bearing the brunt of this massive disease burden; 95% of TB cases and 98% of TB deaths occurred in the developing world.\(^1\) What is even more alarming is the projected scale of the epidemic to come; the World Health Organization (WHO) has estimated that between 2000 and 2020 1 billion people will be newly infected with *M. tuberculosis*, resulting in 200 million TB cases and 35 million deaths.\(^2\)

TB is a major cause of childhood morbidity and mortality in developing countries.\(^3\) Accurate figures on the extent of paediatric TB are not available due to inadequate health information systems in endemic countries and the limited attention paid to children who contribute little to TB transmission within affected communities. WHO estimates of disease incidence are based on sputum smear-positive cases, but more than 80% of children with TB are sputum smear-negative. Indications are that children contribute at least 15-20% of the total disease burden in endemic areas and that they suffer severe TB related morbidity and mortality.\(^5\) Recognition of this high disease burden and the fact that highly effective treatment is available has led to increased emphasis on the diagnosis and management of paediatric TB. WHO research priority guidelines for paediatric TB identified the evaluation of new techniques to improve the diagnosis and management of paediatric TB as an urgent research priority.\(^6\)
According to 2005 WHO figures, South Africa, despite its relatively good infrastructure and health care services, reported a TB incidence of 600/100,000 population/year with a mortality rate of 71/100,000 population/year.\textsuperscript{1} The Western Cape Province, a very high burden region in 2007 reported the adult TB incidence as 1005.7 per 100,000 compared to the National incidence rate of 739.6/100,000 population.\textsuperscript{7} In 2007 the Western Cape Department of Health reported a TB notification rate of 620 per 100 000 in children (0-14 years of age).\textsuperscript{8} A prospective observational study conducted in Cape Town demonstrated that children less than 13 years of age contributed 13.7% of the total TB caseload with a calculated TB incidence of 407/100,000/year; 52.2% of that in adults (≥ 13 years of age).

\textit{Mycobacterium bovis BCG}

BCG vaccination is recommended by the WHO for infants in TB-endemic areas as it affords protection against disseminated disease in children.\textsuperscript{9} However, BCG vaccination may be associated with local complications, such as abscess formation at the vaccination site and/or regional lymphadenitis. Prior to the emergence of HIV as a cause of immunosuppression, disseminated disease was rare (0.19–2 cases/1 million vaccinated infants) and was associated with congenital immunodeficiency syndromes.\textsuperscript{10}

In South Africa, a change in vaccine policy in July 2000 from percutaneous Tokyo strain BCG to intradermal Danish strain BCG vaccine has seen a number of adverse events reported in both HIV-infected and immune competent children.\textsuperscript{9,11-12} Distant or disseminated disease has become a serious concern in human immunodeficiency virus (HIV)-infected children. In 2007 it was estimated that 417 per 100 000 HIV-infected infants, are affected by disseminated BCG disease,\textsuperscript{12} a rate that is about 1000 times the rate in HIV-uninfected infants, and with a mortality rate of 75–86%.\textsuperscript{13}
WHO recommended in 2007 that HIV-infected infants should not receive BCG vaccination, but this policy is very difficult to implement and may not be beneficial in countries where both HIV and TB are endemic, as the HIV status of the baby is not known at the time of routine vaccination (at birth). Where possible however every effort should be made to introduce selective BCG vaccination based on known HIV status.

**Non-tuberculous mycobacteria (NTM)**

Non-tuberculous mycobacterial infection (NTM) is due to mycobacterial organisms other than *Mycobacterium leprae* or members of the *Mycobacterium tuberculosis* complex and differ from the former in their habitat, contagiousness and susceptibility to chemotherapy.

In developed countries such as the USA, as the incidence of tuberculosis has fallen, NTM are more frequently isolated than those of the *M. tuberculosis* complex. In immune competent adults NTM infection presents as pulmonary disease in a patient with pre-existing chronic pulmonary disease, whilst in immune competent children the organs involved are the lymph nodes and the skin. These children are generally under the age of six years and are otherwise healthy with no constitutional symptoms.

In countries with a high prevalence of TB and HIV, localized or disseminated disease from non-tuberculous mycobacteria may occur, but is uncommon. However, a case study from Thailand showed that of the 153 HIV-infected children who had initiated antiretroviral therapy, nine developed the immune reconstitution syndrome (IRIS) due to non-tuberculous mycobacterial organisms. This highlights the need for bacteriological confirmation and speciation in suspected cases of mycobacterial infection.

**Contribution of HIV**

One of the millennium development goals of the United Nations is to reduce TB-related mortality to 50% of the 1990 levels. Increasingly it appears that this goal will not be achieved,
particularly in sub-Saharan Africa, due in part to HIV infection, which is a major contributor to the escalating TB epidemic in this region and the rest of the world. It is estimated that in 2007 about 420,000 new HIV infections occurred in children, predominantly in sub-Saharan Africa, through perinatal transmission, in a population vulnerable to early contact with tuberculosis. A recent study in the Western Cape Province, South Africa, recorded a TB incidence of 1596 (disseminated TB 240)/100,000 in HIV-infected and 66 (disseminated 14)/100,000 in HIV-uninfected infants.

Autopsy studies in sub-Saharan Africa have shown that 30 to 40% of deaths in HIV-infected adults and up to 20% in HIV-infected children are due to TB. In Zambia, Cote d’Ivoire, Malawi, and South Africa, TB-related mortality is considerably higher in HIV-infected compared to HIV-uninfected children. Despite this high disease risk experienced by HIV-infected children, the majority of child TB cases are still in HIV-uninfected children, as compared to adults relatively fewer children are HIV-infected and young children are highly susceptible to develop TB irrespective of their HIV status. When children are co-infected with HIV and TB however, they show more rapid progression of the disease and increased morbidity and mortality.

The diagnostic dilemma

Children rarely have sputum smear-positive TB. Although this reduces their risk of actively spreading the disease, it makes it more challenging to establish a definitive diagnosis. In contrast to adults where the sensitivity of sputum culture approximates 80-90%, young children are unable to expectorate, alternative specimens such as gastric aspirates or induced sputa are difficult to collect and culture yields are low (widely reported as 30-40%). Chest radiography is regarded as the most practical test that provides a reasonably accurate diagnosis of intra-thoracic TB in children with suspicious symptoms. However, it has numerous limitations including limited availability, expertise is required to interpret child
radiographs,\textsuperscript{33} and it has no value in the diagnosis of extra-thoracic TB. A recent survey demonstrated that 10.7\% of children treated for active TB had extra-thoracic disease manifestations only.\textsuperscript{34} TB lymphadenitis was the most common form of extra-thoracic tuberculosis in these children (50\%).

Traditionally, three fasting gastric aspirates samples are collected on three consecutive mornings, requiring hospitalization of the child and frequently the caregiver. In developing countries this may create severe hardship as caregivers usually have more than one child in their care. Caregivers may also compromise their employment by remaining in hospital for a prolonged period of time. Hospitalization incurs cost and occupies beds which may be needed for other children. Sputum induction using mobilization and hypertonic saline have shown improved yield compared to gastric aspirates,\textsuperscript{30} but more recent studies demonstrated equivalence with gastric aspirates.\textsuperscript{35} The technique requires specialized training and equipment and may present a nosocomial transmission risk.\textsuperscript{27, 32, 36}

Novel culture methods have been developed in an attempt to circumvent the slow turnaround times, poor sensitivity and cost of conventional automated liquid broth systems. The most feasible alternative to date has been the microscopic observation drug susceptibility assay (MODS) that uses an inverted light microscope to rapidly detect “spindle and cord formation” in selective broth culture that is indicative of mycobacterial growth.\textsuperscript{32} Phage amplification assays that use bacteriophages to detect the presence of live \textit{M. tuberculosis} have been less successful.\textsuperscript{27}

There is abundant literature utilizing commercial and in-house PCR for the diagnosis of mycobacterial infection in sputum and body fluids, such as cerebral spinal fluid, but results in the literature are highly variable and have not been well validated in children.\textsuperscript{27} Recent reviews and meta-analyses of PCR in TB meningitis, pleuritis and sputum smear-negative pulmonary TB demonstrated poor sensitivity.\textsuperscript{37} In patients with sputum smear-positive TB, however, PCR may play an invaluable role in offering rapid species identification and
detection of drug resistance. The application in patients with sputum smear-negative or extra-pulmonary TB requires further evaluation and use in low income countries remains limited due to cost constraints..

T-cell assays that measure interferon-γ released by lymphocytes in peripheral blood after exposure to *M. tuberculosis* specific antigens, have been hailed as reliable TB tests; two commercial tests are available T-SPOT.TB and QuantiFERON –TB GOLD. However, study results are highly variable and the experience in children remains limited. Current consensus is that these tests are unable distinguish latent infection (a third of the world population is latently infected) from active disease and adds little to the traditional tuberculin skin test (TST). Tests are also too expensive and complex for routine use in low-income countries.

**Fine Needle Aspiration Biopsy (FNAB)**

**History**

FNAB has been used since the early 20th century to diagnose infectious and neoplastic disease. Prior to the 1930s there were isolated case reports documenting its history, and the first large scale report on needle aspiration biopsies was published from Memorial Hospital, New York in 1930. In the 1960s this technique was championed by the Scandinavians and was widely practiced, with many publications on its utility, mainly in the Scandinavian literature. Gradually the use of FNAB spread through Europe and interest was rekindled in the United States in the 1980’s. FNAB is now widely utilized as a first line diagnostic procedure in the diagnosis of palpable masses, including peripheral lymphadenopathy. Its value in the diagnosis of mycobacterial lymphadenitis in adults is well documented. FNAB is utilized throughout the developed and developing world, although the indications differ in these two groups. In a review of MEDLINE by Das from 1966 to 2002, 849 journals published 5609 articles on FNAB, the number increasing sharply from the 1980s
onwards. Of the developed or high income countries, 52.8% (28/53) published articles compared to 29.7% (46/155) of the low income or developing economies. South Africa was one of the top 10 countries in the developing world with publications on FNAB. Although the developed countries publish significantly more articles than the developing countries, these tend to concentrate on breast and pancreatic disease, while the developing countries publish significantly more articles on small round cell tumors and infectious diseases. This may reflect the health care priorities and diagnostic needs of these developing countries.

Children with pulmonary TB have extra-thoracic disease manifestations in 10 to 30% of cases.\textsuperscript{45} TB lymphadenitis is the most common form of extra-thoracic TB in endemic areas,\textsuperscript{46} where up to 50% of extra-thoracic disease manifests as peripheral lymphadenopathy. In fact, TB lymphadenitis is the most common (22-48%) cause of persistent cervical lymphadenopathy in TB endemic areas.\textsuperscript{47-50} This is rarely recognized as a potential means for specimen collection. Therefore it is important to develop methods that will provide a rapid and definitive mycobacterial diagnosis in children with persistent cervical lymphadenopathy. In these high burden TB endemic areas, where children may contribute up to 40% of the TB caseload,\textsuperscript{51} and assuming a conservative estimate of 5-10% of children with TB have peripheral lymphadenopathy, FNAB could make a significant contribution to the diagnosis of TB and other mycobacterial infections.

A recent retrospective study comparing FNAB to gastric aspirates and induced sputum in children with pulmonary TB and peripheral lymphadenopathy showed FNAB to have a superior diagnostic yield and a significantly reduced time to diagnosis.\textsuperscript{52} An accurate bacteriological diagnosis was made within 3 days in the vast majority of patients, which has important benefits for patient management. FNAB should be regarded as a first line diagnostic modality in paediatric TB suspects with peripheral lymphadenopathy.

TB and HIV have many features in common which contribute to diagnostic difficulties, particularly in children. In this population both are chronic diseases that
commonly present with pulmonary symptoms/signs and/or lymphadenopathy. Persistent peripheral lymphadenopathy is itself a criterion for the classification of HIV related disease in children. Radiological features may be impossible to interpret with certainty and in children with CD4 T-cell depletion, alternative tests such as the TST and or novel T-cell assays offer little diagnostic assistance due to poor sensitivity.

A study from South Africa demonstrated that the majority of HIV-infected children with persistent lung disease have persistent peripheral lymphadenopathy (PPL) and PPL was present in many children with pulmonary TB. In HIV-infected children, peripheral lymphadenopathy may be associated with many pathological entities, from infections to neoplasia. Thus, enlarged peripheral lymph nodes were identified as important specimen collection sites to consider, in establishing a definitive diagnosis of TB in HIV-infected children. Another important disease presentation results from the immune reconstitution inflammatory syndrome (IRIS), which may exacerbate the symptoms and signs of mycobacterial disease and needs to be distinguished from TB treatment failure. BCG-related right sided axillary adenitis is the most common IRIS manifestation in BCG-vaccinated infants recently started on highly active anti-retroviral therapy (HAART).

**FNAB in children**

FNAB has been less widely utilized in pediatrics as a diagnostic modality, although the literature demonstrates a steady increase in the use of FNAB in children, achieving sensitivity and specificity rates of over 90% in these studies. The majority of the studies were in pediatric oncology patients, where they play a role in triage, as childhood malignancy can be difficult to diagnose, with signs and symptoms which often mimic other common paediatric conditions especially viral infections. With significant mass lesions, FNAB proved to be the diagnostic tool of choice in the triage of these patients. However, in developing countries with a high burden of infectious diseases such as TB and HIV, FNAB can be of inestimable
value in confirming the diagnosis of mycobacterial infection, permitting early appropriate therapy, as well as a means to obtain specimens for culture, bacterial species determination and sensitivity testing. 60, 62-64

FNAB is a simple and minimally invasive technique, performed at the bedside, and is well tolerated by children. Table 1 provides a summary of the equipment required and how to perform the technique. Children under the age of six years are given are oral or intranasal sedation for amnesia, but children older than six years tolerate aspiration with no sedation. The technique is explained to the child, the caregiver is asked to remain with the child during the procedure and the permission of the child is requested to proceed. If the child understands that this is a very quick minimally painful procedure that will enable them to return home after a short period of observation, they are willing to cooperate.

Most aspirates in children are from axillary or cervical lymph nodes that are easily accessible. If the procedures are performed correctly, with a small gauge needle (22-23 G or smaller) complications such as a small hematoma are rare. If the correct technique is utilized the yield as well as the acceptance of the procedure by the patients will be excellent and nursing sisters can be taught this technique with excellent results. In developing countries where TB is endemic and specimens for bacteriological diagnosis are difficult if not impossible to obtain from children, FNAB provides the means for accurate diagnosis in a significant percentage of children with mycobacterial infection. This is particularly important in countries with a high HIV rate, as well as increasing rates of multi drug-resistant (MDR) and extensively drug-resistant (XDR) TB.

FNAB is a cost-effective diagnostic modality that may be performed at the bedside as an outpatient procedure. 62 It requires no infrastructure or sterile environment and no sophisticated or expensive equipment. Superficial aspirates may be performed by clinicians, pathologists or trained nursing personnel and do not require local or general anesthesia. 61
FNAB is defined as an aspirate done using a cutting needle no larger than 22G. This ensures minimal complications, the commonest of which is a small hematoma and thus FNAB is the safest method of obtaining a tissue diagnosis.

A number of different techniques have been used to obtain a tissue and/or bacteriological confirmation of TB lymphadenitis. These include core needle biopsy, wide needle aspiration biopsy and surgical lymph node biopsy. None are as simple, safe and cost effective to perform, as FNAB (Table 2). Compared to core needle biopsy, the risk of sinus or fistula formation, permanent damage to nerves or seeding of tumor along the needle track is minimal. It permits sampling of the entire mass (node) as multiple excursions of the needle are performed in the lesion without withdrawing the needle, and is no more painful than venepuncture. No more than one ml of suction is applied during the aspiration as the cutting needle will ensure material enters the needle by capillary action, while more suction would increase the risk of bleeding and hematoma formation.

Most aspirates yield a very small amount of material which is expressed onto a glass slide and a monolayer smear prepared. Preparation and staining of the smears can be done in a very basic laboratory using manual methods. Although screening and reporting of the aspirates should be performed by a cytopathologist. The smears are robust and may be transported or even couriered to a central laboratory service for diagnosis and reporting.

FNAB is an effective and safe modality for obtaining a representative sample of material from a lymph node, but laboratory confirmation of mycobacterial adenitis can be made utilising a number of different investigations.

**Laboratory analysis**

**Cytomorphology**

Cytomorphology is simple, but not specific, as other opportunistic infections may present with similar cytological changes and the quality of the smear is operator dependent. A
number of reactions have been described in the literature: epithelioid granulomas without necrosis, epithelioid granulomas with necrosis and necrosis without granuloma formation.\textsuperscript{68}

\textit{Ziehl-Neelson (ZN) staining}

ZN staining, even in good laboratories, is insensitive, identifying organisms in only about 20\% of culture positive samples.\textsuperscript{44} Identification of the mycobacterial organism may be dependent on the immune status of the patient and is reflected in the cytomorphological pattern. Smears with poor granuloma formation are more likely to be positive on Ziehl-Neelsen staining than those showing good epithelioid granulomas without necrosis.\textsuperscript{68}

\textit{Autofluorescence}

Autofluorescence of mycobacteria is simple, sensitive and inexpensive, but is not widely used. It is not highly specific, as the specificity is dependent on the experience of the operator and it cannot differentiate between the various mycobacteria.\textsuperscript{62, 69} The recent development of light-emitting diode (LED) technology provides a cheap and reliable light source with a usable life of more than 50,000 hrs using a fraction of the energy required by conventional light bulbs and can run on batteries or inexpensive low-voltage power supplies. Preliminary studies have indicated that LED-fluorescence microscopy offers a valid alternative to conventional fluorescence microscopy\textsuperscript{70} and studies are underway to investigate the diagnostic utility of LED-auto fluorescence microscopy to detect Mycobacteria in fine needle aspiration samples.

\textit{Culture}

Direct inoculation into MGIT or Bactec tubes (Becton Dickinson, USA) at the bedside gives a high yield,\textsuperscript{62} but it takes 1-6 weeks for a result, and fails to differentiate the various mycobacteria. Additional PCR-based testing is required for accurate speciation.. Direct
inoculation however is not feasible as the MGIT and Bactec collection media are expensive, and are not readily available in clinics and wards for use by clinicians. We have developed an inexpensive transport bottle which is easy to prepare “in house” and does not require refrigeration.\textsuperscript{71} The positive culture yield and the time to positive culture from the transport medium, even after 7 days at room temperature, is statistically no different to direct bedside inoculation. The production and distribution of these bottles to clinics and hospitals, will increase the feasibility of diagnosing tuberculosis at primary health care level.

\textit{Nucleic acid amplification tests (NAATs)}

A small number of recent studies have shown considerable promise in the use of NAATs in the diagnosis of \textit{M. tuberculosis} on FNAB specimens.\textsuperscript{72-75} These studies have been predominantly in adults. There is limited data on the use of FNAB for the diagnosis of mycobacterial disease in children. Rapid and minimally invasive techniques such as FNAB, which can be performed on an outpatient basis, in a primary health care setting, when combined with a rapid and sensitive diagnostic technique such as PCR, may substantially contribute to the effective management of mycobacterial infection in HIV-infected and uninfected children. The implications of a rapid and accurate diagnosis include expediting access to appropriate and adequate therapy as well as potentially limiting further costly investigation.
Conclusion

It is difficult to understand why such an effective, simple and safe diagnostic modality is so underutilized in the diagnostic workup of pediatric tuberculosis. It is ideally suited for widespread use in resource limited countries which carry the dual burden of TB and HIV. FNAB may be perceived as a procedure to be carried out by specialized medical personnel, but it can safely be carried out by junior medical staff and nursing staff, capable of and trusted with inserting intravenous lines. It has been demonstrated that samples collected by nurse aspirators trained during a Burkitt’s lymphoma study conducted in Malawi is of similar quality as those obtained by trained pathologists.

Adequate training is essential to develop confidence in the technique and to ensure a good yield. This can be done through initial outreach teaching and training programs, followed by cascade training to facilitate widespread dissemination of FNAB skills. Pathology departments from South Africa and organizations such as the British Division of the International Academy of Pathology and the Friends of Africa through the United States and Canadian Academy of Pathology (USCAP) offer ongoing education in FNAB and pathology in African and other TB-endemic countries. Most training programs incorporate tuition in the cytopathological diagnosis of mycobacterial infection by pathologists, as the advent of HIV has changed the cytomorphological appearance of these infections.

In addition to making a rapid and definitive tissue diagnosis, widespread use of FNAB would provide ready access to material ideally suited to develop alternative methods for rapid detection of mycobacteria, species determination and drug resistance testing. In order to reduce the massive TB disease burden suffered by children in endemic areas we have to utilize creative and effective means to ensure rapid and accurate diagnosis, such as FNAB, that would facilitate early and appropriate treatment.
REFERENCES


**Table 1**

**Summary of the Fine Needle Aspiration Biopsy Technique**

<table>
<thead>
<tr>
<th>Equipment</th>
</tr>
</thead>
<tbody>
<tr>
<td>22 or 23G cutting needles</td>
</tr>
<tr>
<td>10ml disposable plastic syringes</td>
</tr>
<tr>
<td>Glass cytology slides (ground edges)</td>
</tr>
<tr>
<td>Commercial spray fixative or 95% alcohol</td>
</tr>
<tr>
<td>Non Sterile gloves</td>
</tr>
<tr>
<td>Alcohol swabs</td>
</tr>
</tbody>
</table>

**Procedure**

Clean skin, immobilize the mass, position needle to access entire mass and avoid passing through muscles such as sternocleidomastoid.

Insert needle firmly and apply no more than 1ml suction, applying constant suction throughout aspirate.

Aspirate moving the needle in a fan like fashion through the mass.

When material is in the hub of the needle, release suction and withdraw needle.

Ask parent or assistant to apply pressure to puncture wound.

**Preparation of smears**

Remove needle from syringe, pull 10 ml of air into syringe and reattach needle.

This air is used to express the material in needle onto a glass slide. Place second glass slide face down on first, allow material to spread gently, and keeping both slides together, pull gently apart.

Fix one slide with alcohol and allow the second to air dry.

**Culture**

If liquid culture or transport medium is available, withdraw an aliquot of liquid media into the syringe and then expel it back into the tube or bottle, thereby using the culture media to rinse the syringe and needle in a sterile fashion.
Table 2
Comparison of different superficial mass lesion sample collection methods to achieve a definitive tissue diagnosis

<table>
<thead>
<tr>
<th></th>
<th>Open surgical biopsy</th>
<th>Core needle biopsy</th>
<th>Standard Needle Aspiration (greater than 22G)</th>
<th>Fine Needle Aspiration Biopsy (22G or smaller)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Suitable for small lesions (1x1cm)</td>
<td>Yes</td>
<td>No</td>
<td>Yes</td>
<td>Yes</td>
</tr>
<tr>
<td>Entire mass sampled</td>
<td>Yes, if excision</td>
<td>No</td>
<td>No</td>
<td>Yes</td>
</tr>
<tr>
<td>Complications</td>
<td>Risk of anaesthesia, hospitalization, sinus formation, infection</td>
<td>Sinus formation, infection</td>
<td>Rare</td>
<td>Extremely rare</td>
</tr>
<tr>
<td>Cost</td>
<td>Hospitalization, theatre time</td>
<td>High cost of needle</td>
<td>Minimal</td>
<td>Minimal</td>
</tr>
<tr>
<td>Hospitalisation</td>
<td>Yes</td>
<td>No</td>
<td>No</td>
<td>No</td>
</tr>
<tr>
<td>Anaesthetic required</td>
<td>General</td>
<td>Local</td>
<td>Local</td>
<td>None</td>
</tr>
<tr>
<td>Time for entire procedure</td>
<td>1-2 days</td>
<td>30 minutes</td>
<td>30 minutes</td>
<td>10 minutes</td>
</tr>
<tr>
<td>Tissue diagnosis</td>
<td>Yes</td>
<td>Yes</td>
<td>Yes</td>
<td>Yes</td>
</tr>
<tr>
<td>Microscopy for organism</td>
<td>Yes</td>
<td>Yes</td>
<td>Yes</td>
<td>Yes</td>
</tr>
<tr>
<td>Culture</td>
<td>Yes</td>
<td>Yes</td>
<td>Yes</td>
<td>Yes</td>
</tr>
<tr>
<td>Time for initial result</td>
<td>1-2 days</td>
<td>1-2 days</td>
<td>12-24 hours (Possible in &lt;1 hr)</td>
<td>12-24 hours (Possible in &lt;1hr)</td>
</tr>
</tbody>
</table>
Table 3

Comparison of methods to confirm a mycobacterial diagnosis in children

<table>
<thead>
<tr>
<th>Method</th>
<th>Hospitalization required</th>
<th>Suitable ages</th>
<th>Local/general anaesthetic</th>
<th>Equipment / infrastructure</th>
</tr>
</thead>
<tbody>
<tr>
<td>FNAB</td>
<td>No</td>
<td>All</td>
<td>No</td>
<td>None</td>
</tr>
<tr>
<td>Gastric aspirates</td>
<td>Yes</td>
<td>All</td>
<td>No</td>
<td>None</td>
</tr>
<tr>
<td>Induced sputum</td>
<td>Yes</td>
<td>All</td>
<td>No</td>
<td>Nebulizer, suction, saturation monitor</td>
</tr>
<tr>
<td>Sputum</td>
<td>No</td>
<td>&gt;7-8yrs only</td>
<td>No</td>
<td>None</td>
</tr>
<tr>
<td>Surgical biopsy</td>
<td>Yes</td>
<td>All</td>
<td>Yes</td>
<td>Full theatre</td>
</tr>
</tbody>
</table>
CHAPTER 3

TB LYMPHADENITIS: A COMMON CLINICAL PROBLEM

Tuberculous Lymphadenitis as a Cause of Persistent Cervical Lymphadenopathy in Children From a Tuberculosis-Endemic Area
Marais BJ, Wright CA, Schaaf HS, Gie, RP, Hesseling A, Enarson D, Beyers N.
Paediatric Infectious Diseases Journal 2006; 25:142-146.
Tuberculous Lymphadenitis as a Cause of Persistent Cervical Lymphadenopathy in Children From a Tuberculosis-Endemic Area

Ben J. Marais, MRCP (Paed), FCP (Paed), MMed,* Colleen A. Wright, MMed, FCPath, FRCPath, FIAC,† H. Simon Schaaf, MMed, MD (Paed),* Robert P. Gie, FCP (Paed),* Anneke C. Hesseling, MBChB, MSc,* Don A. Enarson, MD,‡ and Nulda Beyers, FCP (Paed), MSc, PhD*

Background: Cervical lymphadenitis is the most common form of extrapulmonary tuberculosis in children, although its relative contribution as a cause of persistent cervical adenopathy is not well-documented. The aim of this study was to determine the relative contribution of tuberculous lymphadenitis as a cause of persistent cervical adenopathy in a tuberculosis-endemic setting and to document its clinical presentation at the primary health care level.

Methods: A prospective descriptive study was conducted from February 2003 through October 2004 in Cape Town, South Africa. The study included all children younger than 13 years presenting with persistent cervical adenopathy to the local primary health care clinic.

Results: A total of 158 children were evaluated of whom 35 (22.2%) were diagnosed with tuberculous lymphadenitis. Bacteriologic confirmation was achieved in 27 of 35 (77.1%) children; all 35 responded to standard antituberculosis treatment. The majority of those without tuberculous lymphadenitis (105 of 123, 85.4%) had a visible superficial lesion in the area drained by the affected nodes. In children with persistent lymphadenopathy ≥2 × 2 cm, tuberculosis lymphadenitis was diagnosed in 31 of 33 (93.9%); specificity was 98.4%, sensitivity was 88.6% and the positive predictive value was 93.4%.

Conclusion: Children commonly present with persistent cervical adenopathy at the primary health care clinic. The use of a simple clinical algorithm provided an accurate diagnosis of tuberculous lymphadenitis in the study setting. Fine needle aspirations provided a rapid and definitive diagnosis in the majority of children and will have added diagnostic value in settings where alternative diagnoses are more likely.

Key Words: tuberculosis, lymphadenitis, children, persistent cervical adenopathy

Peripheral tuberculous lymphadenitis predominantly involves the cervical lymph nodes and is the most common form of extrapulmonary tuberculosis in children from tuberculosis-endemic areas. However, its relative contribution as a cause of persistent cervical adenopathy in these communities is not well-documented.

Cervical lymphadenitis, caused by Mycobacterium tuberculosis, is generally considered to have its origin in the lymphatic spread of organisms from a primary pulmonary focus, but in a minority of cases it can originate from a primary focus in the mouth, tonsils, oropharynx or tissues of the head and neck. Other mycobacteria can also cause cervical lymphadenitis; the relative contribution of different mycobacteria is influenced by the control of bovine tuberculosis, the use of BCG vaccination, the presence of environmental mycobacteria and the prevalence of tuberculosis within a particular setting.

The diagnosis of tuberculosis in children is often difficult, given that symptoms and signs might be nonspecific, the collection of bacteriologic specimens problematic and bacteriologic yields low. In children with peripheral tuberculous lymphadenitis, however, clinical signs are usually apparent, and fine needle aspiration (FNA) provides excellent bacteriologic yields. Although the diagnostic value of FNA has been demonstrated in resource-limited settings, to date it remains underutilized as a routine diagnostic modality in most endemic areas.

The aim of this study was to determine the relative contribution of tuberculous lymphadenitis as a cause of persistent cervical adenopathy in children from a tuberculosis-endemic area and to document its clinical presentation at the primary health care level.

Methods

A prospective descriptive study was performed from February 2003 through October 2004 in Cape Town, the Western Cape Province, South Africa.
Setting. The study was conducted at 5 primary health care clinics served by one referral hospital. The incidence of all tuberculosis in Cape Town was 678/100,000,14 and the prevalence of human immunodeficiency virus (HIV) infection among women attending public antenatal clinics in the Western Cape Province was 13.1% (95% confidence interval, 8.5–17.7%), in 2003.15 Bovine tuberculosis is well-controlled within the study communities, and children receive routine neonatal Calmette-Guérin bacillus (BCG) vaccination. The study communities rarely use private medical services, and children diagnosed with tuberculosis are routinely referred to the local primary health care clinic, where supervised antituberculosis treatment is provided free of charge. Pediatric services are extended only to children younger than 13 years of age.

Study Population. All children (younger than 13 years of age) who presented with persistent cervical adenopathy; defined as lymph nodes ≥ 1 × 1 cm, persisting for ≥4 weeks despite a course of oral antibiotics (usually amoxicillin), were referred to the investigator for evaluation.

Data Collection and Surveillance. The principal investigator visited each clinic on a weekly basis to screen referred children, whereas a study nurse recorded children referred directly to hospital. The areas surrounding the affected cervical lymph nodes were inspected to exclude a visible superficial lesion within their drainage area (visible local cause), such as impetigo of the scalp, tinea capitis or traction folliculitis. Those with a visible local cause were given appropriate therapy and instructed to return if the lymph nodes persisted or increased in size. Surveillance was continued at all 5 clinics and the referral hospital throughout the study period, and for an additional 3 months after enrollment was stopped, to document any child who subsequently returned with symptom deterioration or a possible diagnosis of tuberculosis.

Children with no visible local cause of cervical lymphadenopathy received a Mantoux (2 tuberculin units of purified protein derivative RT23 intradermal) tuberculin skin test (TST) and a chest radiograph (anteroposterior and lateral). The Mantoux TST was interpreted as positive if induration was ≥ 10 mm. Two independent experts evaluated the chest radiographs for intrathoracic signs of tuberculosis. Symptoms and signs as well as possible tuberculosis contact were documented. Children diagnosed with tuberculous lymphadenitis were offered a rapid HIV test (Determine HIV 1/2; Abbott) after appropriate counseling. Standard 3-drug antituberculosis treatment was initiated,16 and response to antituberculosis therapy was monitored after a period of 3 months. None of the children with a visible local cause had radiographic or other clinical signs indicative of possible tuberculosis, and all children judged not to have tuberculosis on clinical grounds showed symptom resolution in the absence of antituberculosis treatment.

Case Definition. Tuberculous lymphadenitis was defined as cervical lymphadenopathy ≥ 1 × 1 cm persisting for > 4 weeks despite a course of oral antibiotics, in the presence of at least 1 of the following criteria: (1) bacteriologic confirmation: isolation of M. tuberculosis from a lymph node, or microscopically visible acid-fast or autofluorescent bacilli associated with amorphous necrosis on cytology; or (2) clinical diagnosis: significant therapeutic response (lymph node size decreased from ≥ 2 × 2 cm to < 1 × 1 cm after 3 months of standard antituberculosis treatment). All cultures positive for M. tuberculosis complex were routinely speciated by polymerase chain reaction to differentiate M. tuberculosis from Mycobacterium bovis and M. bovis BCG.17

FN A nd/or Biopsy. Of the children enrolled, only those with a positive TST (≥ 10 mm) and/or a suggestive chest radiograph and/or a cervical mass ≥ 2 × 2 cm were referred to the hospital for FNA. FNA was performed in the pediatric outpatient department with sedation and pain relief (chloral hydrate 20–50 mg/kg and paracetamol 10–20 mg/kg, administered orally 30–60 minutes before the procedure). The largest discrete node was identified by careful palpation of the cervical mass. Two needle passes were performed with a 23-gauge needle attached to a 10-mL syringe, applying 2 mL of suction. Aspirated material was placed on standard microscope slides, thinly smeared and air-dried or fixed with commercial spray fixative for Giemsa and Papanicoula stains, before flushing the syringe and needle directly into a MGIT (Becton Dickinson, Cockeysville, MD) mycobacterial culture bottle. Microscopic evaluation included cytology, acid-fast staining and autofluorescence.18 Excision biopsies were occasionally performed if children presented with persistent cervical adenopathy to the referral hospital without a letter from the investigator or if the FNA was inconclusive.

Statistical Analysis. Descriptive analysis was conducted with SPSS (version 11.5). The sensitivity and specificity of a simple clinical algorithm were calculated with the use of either bacteriologic confirmation and/or the clinical case definition (see case definition) as the “reference” test.

Parents gave written informed consent for study participation and ethics approval was obtained from the Institutional Review Board of Stellenbosch University, the City of Cape Town Health Department and local health committees.

RESULTS

Persistent Cervical Adenopathy. Of the 167 children who were identified with persistent cervical adenopathy, 9 (5.4%) did not return to the clinic for evaluation by the investigator (Fig. 1). Of the 158 children evaluated, 53 had no visible cause of whom 40, with either a positive TST or a cervical mass ≥ 2 × 2 cm, were referred to hospital to establish a histologic diagnosis. None of the 13 TST-negative children had radiographic or other clinical signs indicative of possible tuberculosis, and all children judged not to have tuberculosis on clinical grounds showed symptom resolution in the absence of antituberculosis treatment.

Table 1 summarizes the demographics and etiology of persistent cervical adenopathy in the 158 children who were evaluated. The majority of children (105, 66.5%) had lymph nodes < 2 × 2 cm with a visible local cause; 28 returned for reevaluation within 1 month; the lymphadenopathy resolved in 18 and decreased to < 1 × 1 cm in 9. Multiple discrete lymph nodes, 1–2 cm in diameter, remained in 1 child who was TST-negative and asymptomatic. The lymphadenopathy showed slow resolution, decreasing to < 1 × 1 cm within 3 months. None of the children with a visible local cause returned to the clinic with symptom deterioration or with a possible tuberculosis diagnosis from any hospital, during the study period.
Of the 53 (33.5%) children without a visible local cause, tuberculous lymphadenitis was diagnosed in 35 (66.0%). Of those with a cervical mass ≥2 × 2 cm, tuberculous lymphadenitis was diagnosed in 31 of 33 (93.9%). Of the 2 remaining children, 1 had an unidentified chronic inflammatory process diagnosed through excision biopsy, and the other developed a chronic bacterial abscess after repeat courses of oral antibiotics, requiring eventual incision and drainage. Two children presented with acute bacterial adenitis; both had a history of persistent lymph node enlargement that preceded the acute event. Both these children were TB-positive and were diagnosed with secondary bacterial infection, as *M. tuberculosis* was cultured after incision and drainage. One had signs suggestive of tuberculosis on the chest radiograph and reported a prolonged cough and night sweats, whereas the other had no additional suspect symptoms or signs apart from the cervical mass and a positive TST.

**Tuberculous Lymphadenitis.** Of the 35 children diagnosed with tuberculous lymphadenitis (Fig. 1), bacteriologic confirmation was achieved in 27 (77.1%) children. Of the 8 children without bacteriologic confirmation, 7 failed to attend the referral hospital, and 1 refused permission for FNA. All 8 had a TST response ≥15 mm and showed excellent response to standard antituberculosis treatment. In the 27 children with bacteriologic confirmation; FNA was performed in 21 children and formal biopsies in 6. No cases caused by *M. bovis, M. bovis* BCG, infection with environmental mycobacteria such as *Mycobacterium avium-intracellulare* complex or *Mycobacterium scrofulaceum* or malignancies were identified.

Of the 21 children with tuberculous lymphadenitis in whom FNA was performed, 16 of 21 (76.2%) were acid-fast or autofluorescent smear-positive, which allowed rapid and definitive diagnosis. Culture confirmation was achieved in 19 of 21 (90.5%), of whom 2 had no microscopic features indicative of tuberculosis. One of the 2 children who were culture-negative had received antituberculosis treatment before FNA was performed. Either a positive culture or typical microscopic features were present in all 21 cases. One of the 2 children, who had no microscopic features indicative of tuberculosis on FNA, had an excision biopsy performed that established the diagnosis, before the FNA culture result became known. No immediate complications relating to the FNA procedure, apart from minimal bleeding, were noted. No long term complications, such as sinus formation, were recorded during the 3-month follow-up period.

Table 2 reflects the lymph node characteristics and associated findings in the 35 children diagnosed with tuberculous lymphadenitis. Using the clinical algorithm of a persistent cervical mass ≥2 × 2 cm, without a visible local cause or response to antibiotics, accurately identified children with tuberculous lymphadenitis; sensitivity was 88.6%, specificity was 98.4% and the positive predictive value of this clinical algorithm was 93.4%.

Tuberculous lymphadenitis occurred in children of all ages, except in infants. In 18 (51.4%), lymph nodes occurred in the anterior triangle, with involvement of multiple regions in 5 (14.3%). Other regions involved were: posterior triangle, 8 (22.9%); submandibular, 2 (5.7%); and supraclavicular, 2 (5.7%). Lymph nodes occurred more regularly on the right side of the neck (36.7%), lower in the neck (11.4%), and upper in the neck (4.6%).

**TABLE 1.** Demographics and Etiology of Persistent Cervical Adenopathy in Children (n = 158)

<table>
<thead>
<tr>
<th>Demographics</th>
<th>No. of Instances</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gender</td>
<td></td>
</tr>
<tr>
<td>Male</td>
<td>69 (43.7) *</td>
</tr>
<tr>
<td>Female</td>
<td>89 (56.3)</td>
</tr>
<tr>
<td>Age groups</td>
<td></td>
</tr>
<tr>
<td>&lt;5 yr</td>
<td>93 (58.9)</td>
</tr>
<tr>
<td>5–9 yr</td>
<td>51 (32.2)</td>
</tr>
<tr>
<td>≥10 yr</td>
<td>14 (8.9)</td>
</tr>
<tr>
<td>Etiology</td>
<td></td>
</tr>
<tr>
<td>Visible local cause</td>
<td>105 (66.5)</td>
</tr>
<tr>
<td>Bacterial infection</td>
<td>26 (16.5)</td>
</tr>
<tr>
<td>Tinea capitis (with secondary infection)</td>
<td>34 (21.5)</td>
</tr>
<tr>
<td>Traction folliculitis</td>
<td>44 (27.8)</td>
</tr>
<tr>
<td>Otitis externa</td>
<td>1 (0.6)</td>
</tr>
<tr>
<td>No visible local cause</td>
<td>53 (33.5)</td>
</tr>
<tr>
<td>Tuberculous lymphadenitis</td>
<td>35 (22.2)</td>
</tr>
<tr>
<td>Reactive nodes</td>
<td>13 (8.2)</td>
</tr>
<tr>
<td>Nonspecific inflammation</td>
<td>4 (2.5)</td>
</tr>
<tr>
<td>Nonacute bacterial abscess</td>
<td>1 (0.6)</td>
</tr>
<tr>
<td>Malignancy</td>
<td>0</td>
</tr>
</tbody>
</table>

*Numbers in parentheses, percent.

*Cervical mass <2 × 2 cm, tuberculin skin test-negative and natural symptom resolution.*
TABLE 2. Clinical Characteristics of Children With Tuberculous Lymphadenitis (n = 35)

<table>
<thead>
<tr>
<th>Character</th>
<th>No. of Instances</th>
</tr>
</thead>
<tbody>
<tr>
<td>Lymph node characteristics</td>
<td>35 (100)*</td>
</tr>
<tr>
<td>Persistence (present for &gt;4 wk, no response to antibiotics)</td>
<td></td>
</tr>
<tr>
<td>Size†</td>
<td></td>
</tr>
<tr>
<td>&lt;2 × 2 cm</td>
<td>4 (11.4)</td>
</tr>
<tr>
<td>(2–4) × (2–4) cm</td>
<td>25 (71.5)</td>
</tr>
<tr>
<td>&gt;4 × 4 cm</td>
<td>6 (17.1)</td>
</tr>
<tr>
<td>Character</td>
<td></td>
</tr>
<tr>
<td>Single</td>
<td>5 (14.3)</td>
</tr>
<tr>
<td>Multiple</td>
<td></td>
</tr>
<tr>
<td>Discreet</td>
<td>14 (40.0)</td>
</tr>
<tr>
<td>Matted</td>
<td>16 (45.7)</td>
</tr>
<tr>
<td>Solid</td>
<td>28 (80.0)</td>
</tr>
<tr>
<td>Fluctuant</td>
<td></td>
</tr>
<tr>
<td>Without secondary bacterial infection</td>
<td>5 (14.3)</td>
</tr>
<tr>
<td>With secondary bacterial infection (red and warm)</td>
<td>2 (5.7)</td>
</tr>
<tr>
<td>Associated findings</td>
<td></td>
</tr>
<tr>
<td>Tuberculin skin test</td>
<td></td>
</tr>
<tr>
<td>0 mm</td>
<td>2 (5.7)</td>
</tr>
<tr>
<td>1–9 mm</td>
<td>0</td>
</tr>
<tr>
<td>≥10 mm</td>
<td>33 (94.3)</td>
</tr>
<tr>
<td>≥15 mm</td>
<td>32 (91.4)</td>
</tr>
<tr>
<td>Mean response 19.1 mm (standard deviation 2.9 mm)</td>
<td></td>
</tr>
<tr>
<td>Constitutional symptoms</td>
<td></td>
</tr>
<tr>
<td>Any symptom</td>
<td>21 (60.0)</td>
</tr>
<tr>
<td>Fever</td>
<td>7 (20.0)</td>
</tr>
<tr>
<td>Cough</td>
<td>9 (25.7)</td>
</tr>
<tr>
<td>Night sweats</td>
<td>8 (22.8)</td>
</tr>
<tr>
<td>Fatigue†</td>
<td>19 (54.3)</td>
</tr>
<tr>
<td>Failure to thrive†</td>
<td>10 (28.6)</td>
</tr>
<tr>
<td>Chest radiograph</td>
<td></td>
</tr>
<tr>
<td>Suggestive of tuberculosis</td>
<td>13 (37.1)</td>
</tr>
<tr>
<td>Lymph node disease</td>
<td></td>
</tr>
<tr>
<td>Uncomplicated</td>
<td>8 (22.8)</td>
</tr>
<tr>
<td>With airway compression</td>
<td>1 (2.9)</td>
</tr>
<tr>
<td>With parenchymal consolidation</td>
<td>4 (11.4)</td>
</tr>
</tbody>
</table>

Numbers in parentheses, percent.
†Transverse diameter of the largest cervical mass.
‡Less playful and active since the mass was first noted.
§Numbers in parentheses, percent.
© 2006 Lippincott Williams & Wilkins

DISCUSSION

Children commonly presented with persistent cervical adenopathy to primary health care clinics in the study setting. The majority of these children had visible local lesions such as crusts of impetigo, infected tinea capitis or traction folliculitis. The hair of most small girls in this community is tightly braided, which often leads to irritation and bacterial infection of the hair follicles. This also explains the overrepresentation of girls in the study population. It is understandable that none of these conditions responded to a short course of oral antibiotics, in the absence of etiology-specific systemic and/or local treatment.

A simple clinical algorithm that identified children with persistent (>4 weeks) cervical lymphadenopathy, no visible local cause or response to antibiotics and a cervical mass >2 × 2 cm showed excellent diagnostic accuracy within the study setting. The addition of a positive TST may be of value in settings where infection with M. tuberculosis is less common or where conditions other than tuberculosis, such as malignant lymph node involvement or other chronic infections, are more common and may present with a similar clinical picture. The fact that accurate clinical diagnosis is possible at the primary health care level might allow the initiation of antituberculosis treatment without hospital referral, which should improve access to care for children in extremely resource-limited settings. However, regular follow-up (at least monthly) is essential so that children who do not respond to standard antituberculosis treatment are referred as soon as possible to establish a definitive diagnosis. The value of this clinical approach requires further evaluation.

FNA proved to be a robust and simple technique, which provided an excellent bacteriologic yield. No significant side effects were noted. FNA provided a definitive bacteriologic diagnosis with the ability to speciate M. tuberculosis complex and to perform drug susceptibility testing. A definitive diagnosis is always desirable, but the diagnostic value of FNA will be even greater in settings where a clinical diagnosis is expected to be less accurate, such as areas where malignant lymph node involvement and/or other chronic infections are more common.11,12

The lymph node characteristics and constitutional symptoms recorded in this study correlate well with findings from Papua New Guinea and India.3,4,19 The most distinctive features were the chronic persistent course, the lymph node size (>2 × 2 cm) and the involvement of multiple, discrete or matted, lymph nodes. The study also emphasizes the value of
clinical follow-up and serial measurement of enlarged lymph nodes if the diagnosis is not apparent.\textsuperscript{20}

Only 1 child with tuberculous lymphadenitis was HIV-infected. This reflects the low HIV prevalence among children in the study setting but might also illustrate a decreased tendency in HIV-infected children to develop peripheral tuberculous lymphadenitis, as suggested in a comparative study from Zambia.\textsuperscript{21} FNA may have increased diagnostic value in HIV-infected children, because the relative contribution of tuberculous lymphadenitis to persistent cervical adenopathy in this group is expected to be smaller.

It is interesting that no environmental mycobacteria were isolated, although it is reported to be the most common cause of persistent cervical adenopathy in the developed world.\textsuperscript{22} Paucity of disease caused by environmental mycobacteria has also been described in other tuberculosis-endemic countries, such as India.\textsuperscript{23,24} This may result from the protective effect afforded by routine neonatal BCG vaccination,\textsuperscript{8} whereas natural infection with \textit{M. tuberculosis} might provide additional protection against disease caused by environmental mycobacteria.

An important study limitation is that bacteriologic confirmation was achieved in only 77.1\% of the study patients, as it was not attempted in 8 children. However, we are confident that the diagnosis was accurate, given the fact that these 8 children all had a TST response $\geq$ 15 mm and showed excellent clinical response to standard antituberculosis treatment.

In conclusion, the use of a simple clinical algorithm identified tuberculous lymphadenitis with a high degree of accuracy in the study setting, whereas FNA provided a rapid and definitive diagnosis in the majority of children.

\textbf{ACKNOWLEDGMENTS}

We thank the primary health care clinics involved, Dr Ivan Toms (City of Cape Town Health Department), the patients and their parents for their kind assistance.

\textbf{REFERENCES}

CHAPTER 4

DIAGNOSTIC UTILITY AND FEASIBILITY OF FINE NEEDLE ASPIRATION BIOPSY

Fine needle aspiration biopsy: diagnostic utility in resource-limited settings
C A Wright, J P Pienaar, Ben J Marais

Fine Needle Aspiration Biopsy - A First Line Diagnostic Procedure in Pediatric Tuberculosis suspects with Peripheral Lymphadenopathy?
Colleen A Wright, MD FRCPath; Anneke C Hesseling, MD, PhD; Colleen Bamford, MD, FCPATH; Steven Michael Burgess, PhD; Rob Warren, PhD; and Ben J Marais, MD, PhD
International Journal of Tuberculosis and Lung Disease-in press
Fine needle aspiration biopsy: diagnostic utility in resource-limited settings

C. A. WRIGHT, J. P. PIENAAR & B. J. MARAIS*

Discipline of Anatomical Pathology, Department of Pathology and NHLS Tygerberg and *Department of Paediatrics & Child Health and Ukwanda Centre for Rural Health, University of Stellenbosch, Tygerberg, South Africa

(Accepted November 2007)

Abstract
Background: Little information is available on the value of fine needle aspiration biopsy (FNAB) in routine paediatric practice in resource-limited settings.

Aim: To provide an overview of all paediatric FNAB samples received at Tygerberg Hospital, Cape Town, South Africa over a 3-year period, including the determinants of sample adequacy and the diagnoses.

Methods: Samples were analysed from three locations: Tygerberg Hospital (TBH) where pathologists performed all the procedures, surrounding clinics where aspirates were mostly performed by doctors with no formal training in FNAB technique, and Queen Elizabeth Hospital, Blantyre, Malawi where FNABs were performed by trained nurse aspirators.

Results: A total of 830 aspirates were reviewed: 464 (56%) from TBH, 264 (32%) from local clinics and 102 (12%) from Blantyre. The main diagnoses at TBH were mycobacterial infection (31%), normal/reactive tissue (27%) and malignancy (14%); malignancy dominated (74%) in the select group from Blantyre. Sample adequacy rates were similar between pathologists and nurse aspirators [399/464 vs 82/102, odds ratio (OR) 1.4, 95% confidence interval (CI) 0.8–2.6]. Results were significantly better in the group who received formal training (TBH and Malawi) than in the clinics where clinicians had no formal training (481/566 vs 171/264, OR 3.1, 95% CI 2.2–4.4).

Conclusions: FNAB provides a definitive tissue diagnosis in the majority of patients. Well-trained nurse aspirators perform as well as pathologists, indicating the feasibility of FNAB in resource-limited settings.

Introduction

Fine needle aspiration biopsy (FNAB) has been used as a diagnostic modality for more than a century. It was initially applied to infective and inflammatory lesions only, but the advent of cytological investigation (around 1925) expanded its diagnostic application to confirm or exclude malignancy also. FNAB is well established as the diagnostic procedure of choice for superficial mass lesions. However, few data are available on its routine use in paediatric practice, especially in resource-limited settings.

FNAB provides a simple, fast, cost-effective and safe method of obtaining a definitive tissue diagnosis. In experienced hands, a high level of diagnostic accuracy is possible, averting the need for a formal biopsy and all its associated risks. Apart from providing a cytological diagnosis, FNAB also provides tissue for ancillary tests, such as microbiological cultures, flow cytometry, cytogenetics, electron microscopy and polymerase chain reaction (PCR).
If the procedure is performed according to established protocols, the disadvantages are minimal.\textsuperscript{9} Potential complications include pain, bleeding, infection, vasovagal reactions, haemothorax, pneumothorax, air embolism, seeding along the needle tract and perforation of organs.\textsuperscript{9} As a general rule, the rate of complications rises with use of larger calibre needles and deep organ aspiration. Contra-indications for FNAB are few but include an abnormal clotting profile (especially for deep organ aspirations) and suspected vascular lesions, and aspiration in the region of the thorax is best avoided in patients with cardiovascular and/or respiratory dysfunction.\textsuperscript{9} The risk associated with the aspiration of superficial masses using small gauge needles (22G or less) is minimal.\textsuperscript{2,9}

Previously at Tygerberg Hospital, clinicians without formal training in the correct technique performed routine FNAB, but since July 2001 trained pathologists perform all paediatric fine needle aspirates. The study aims were to provide an overview of all diagnoses made during the 1st 3 years of this new policy and to investigate possible determinants of sample adequacy.

**Methods**

*Study population*

This retrospective descriptive study reviewed all paediatric FNAB samples received by the cytology laboratory at Tygerberg Hospital (TBH) from 1 July 2001 to 30 June 2004. These included samples received from TBH and surrounding clinics and specimens couriered from Queen Elizabeth Hospital, Blantyre, Malawi. The cases referred from Malawi were all enrolled in a paediatric oncology study and represent a very select group. Children were recruited into this study and underwent FNAB if they presented with a mass lesion suspected of being Burkitt’s lymphoma.\textsuperscript{10}

**Fine needle aspiration biopsy**

FNAB were usually performed in the clinic, in the paediatric outpatient department or in the ward. Children received sedation and pain relief (usually chloral hydrate 20–50 mg/kg and paracetamol 10–20 mg/kg) orally 30–60 minutes before the procedure. Before performing the aspiration, the largest discrete mass was identified by careful palpation, (usually using a 23G-needle attached to a 10-ml syringe and applying 2 ml of suction). Aspirated material was placed on standard microscope slides, thinly smeared and air-dried or fixed with commercial spray fixative for Giemsa and Papanicolaou stains. If tuberculosis was clinically suspected, the needle and syringe were rinsed in mycobacterial growth tube indicator (MGIT, Beckton Dickinson, USA) medium for mycobacterial culture. All mycobacterial isolates were identified as *Mycobacterium tuberculosis*, *Mycobacterium bovis* BCG or non-tuberculous mycobacteria (NTM) by polymerase chain reaction (PCR) testing.

**Data collection and analysis**

Cytology results were captured in a computerised database. We reviewed the results of all FNAB samples received during the 3-year study period from patients ≤16 years of age. Relevant information including patient demographics, specimen characteristics and final diagnosis was transferred to a Microsoft Excel spreadsheet. Descriptive statistical analysis was performed using Statistica 7.

Written, informed consent to the FNAB procedure was obtained from the parent or legal guardian. Ethics clearance was obtained from the institutional review board of the University of Stellenbosch.

**Results**

A total of 830 aspirates were reviewed, 464 (56%) from TBH, 264 (32%) from local
clinics and 102 (12%) from Malawi. In general, the age and gender distribution was uniform (406, 51% female), but Malawi had more males (63% male vs 37% female) and the clinics had more females (57% female vs 43% male).

Peripheral lymph nodes were most frequently aspirated (376, 45%); of the lymph nodes specified, 70% were cervical and 30% axillary. Aspiration sites for the remaining 454 samples were distributed as follows: head and neck (124, 15%), chest and abdomen (98, 12%), breast (66, 8%), site not specified (166, 20%). All 66 breast tissue samples were collected from pubertal girls and came from the clinics, constituting 25% of clinic samples. In Malawi where all the children were clinically suspected of having Burkitt’s lymphoma, the bulk of aspirates (44, 43%) came from the head and neck area, 28 (27%) from the chest, abdomen or pelvis and in 30% of cases the site was not specified.

The cytological diagnosis was assessed in conjunction with the clinical and radiological findings and all clinically or cytologically suspected lymphomas at TBH were aspirated for flow cytometry. Where appropriate, immunocytochemistry was performed on neoplasms. All solid neoplasms were managed according to SIOP (International Society for Paediatric Oncology) protocols, and histology was done after chemotherapy. Where there was any uncertainty as to the precise diagnosis after FNAB, the mass was excised or biopsied prior to chemotherapy.

All cases of suspected mycobacterial infection received bedside inoculation for mycobacterial culture and subsequent speciation.

All aspirates from Malawi were referred after clinical assessment as Burkitt’s lymphoma and, when confirmed by cytology, patients were commenced on limited chemotherapy according to the study protocols and response to treatment was carefully monitored.

The most common diagnostic category was normal/reactive tissue, followed by mycobacterial infection and lymphoma, in particular Burkitt’s lymphoma. However, the distribution of diagnostic categories showed marked variation between the different centres. Mycobacterial disease was the most common diagnosis in children from TBH and Burkitt’s lymphoma in the select group from Malawi. Among the children with mycobacterial infection, *M. bovis* BCG was identified in 20 (22.5%) cases. All these children had right-sided axillary lymph node enlargement and were <2 years of age (average age <1 y). *M. tuberculosis* was identified in 67 (75.3%) cases, mostly from cervical lymph nodes, and was equally prevalent in all age groups (average age 5.7 y). In Malawi, Burkitt’s lymphoma was seen predominantly between the ages of 2 and 12 years (average age 6.3), corresponding to the age of maximum prevalence of endemic Burkitt’s lymphoma.11 No significant seasonal or other time-related variation was recorded on reviewing the temporal profile of cases with TB or Burkitt’s lymphoma.

Of all aspirates received, 464 (56%) were performed by pathologists (TBH), 264 (32%) by clinicians (clinics) and 102 (12%) by nurse aspirators (Malawi). Pathologists and nurse aspirators received formal training in FNAB technique but few of the clinicians. Fig. 1 indicates the adequacy of samples received; in total, 174 (21%) samples were regarded as inadequate. The proportion of inadequate samples varied widely between centres, TBH 14%, Malawi 20% and clinics 35%. This reflects differences between the aspirator groups, adequate samples being 86% in the pathologist group, 80% in the nurse aspirator group and 65% in the untrained clinician group. Sample adequacy rates were similar between pathologists and nurse aspirators (399/464 vs 82/102; OR 1.4, 95% CI 0.8–2.6) but were significantly better in the group who received formal training (TBH and Malawi) than in the clinics where clinicians did not receive formal training (481/566 vs 171/264, OR 3.1, 95% CI
2.2–4.4). The sample adequacy rate also differed according to the number of passes performed, single pass 75% and more than one pass 85% (85% vs 75%, OR 1.9, 95% CI 1.3–2.8).

Discussion

FNAB is widely used as a first-line diagnostic procedure for the diagnosis of mass lesions in adults, but it is used less frequently in children, especially so in resource-limited settings where it has the greatest diagnostic potential. This study demonstrates that FNAB is a feasible option in resource-limited settings; well trained nurse aspirators performed as well as trained pathologists and the procedure can be done on an outpatient basis.

Nurse aspirators from Malawi who attended a short training course at TBH and received clear operating procedure guidelines achieved an adequacy rate of 80%, despite the fact that multiple needle passes were restricted. The technique shown was subsequently implemented within the units. This was a highly select patient group with clinically diagnosed Burkitt’s lymphoma who were enrolled in a trial that investigated cost-effective management of Burkitt’s lymphoma in resource-limited countries. In the absence of a local cytopathologist, the slides were fixed and sent to TBH for review. This trial demonstrated that diagnostic slides can be transported easily and safely using standard courier services, enabling cytopathologists to provide a long-distance diagnostic service for countries without cytopathology services.

The diagnostic value of FNAB is completely dependent on the quality of the sample presented to the cytopathologist. Optimal FNAB technique together with adequate slide preparation and preservation is taught poorly, if at all, to medical students in South Africa. The technique is usually acquired by observing and assisting senior clinicians, most of whom receive no formal training, resulting in the propagation of incorrect techniques and a high percentage of inadequate samples. Samples are frequently bloody, inadequately spread and poorly fixed, making it impossible for the pathologist to interpret the smears. The resulting non-diagnostic reports promote the misconception that FNAB is a technique with a poor diagnostic yield.

Before January 2001, FNAB was not widely used by clinicians at TBH. This was partly due to inadequate training in FNAB technique and under-recognition of the valuable contribution that FNAB could make to the management of infective and neoplastic lesions in children. Since 2001, cytopathologists have provided the FNAB service at TBH, and the diagnostic use and yield of FNAB in children has increased; an outreach programme has been established offering short practical tutorial sessions to
interested clinicians, combined with follow-up of subsequent aspirates received, providing positive feedback and constructive advice.

The advanced training and experience of the cytopathologists might partly explain the excellent results achieved, although nurse aspirators in Malawi who received training in the correct technique achieved comparable results. The performance of more than one needle pass also improved sample adequacy and diagnostic yield. In total, more than one needle pass was performed in 40% of patients, 59% at TBH, 18% in the clinics and 11% in Malawi. In Malawi, multiple needle passes were restricted in an attempt to minimise the risk of needle tract spread, a precaution recommended in children suspected of having a malignant neoplasm.

This lack of training is not unique to developing countries. In a study of breast aspirates, Ljung showed that physicians trained in FNAB technique missed significantly fewer malignant lesions than physicians who had received no training (2% vs 25%, \( p<0.0001 \)).\(^{12}\) This was not affected by the number of aspirates performed by individual aspirators.

Few studies have reported on the value and limitations of FNAB in routine paediatric practice. In the USA, Wakely reported a sensitivity and specificity rate of 97% and a positive predictive value of 95% throughout the entire age range of infancy and childhood.\(^{3}\) Handa showed FNAB to be an excellent procedure for the triage of patients with significant lesions requiring treatment or referral to specialised services in India.\(^{8}\) A paediatric oncology study in the USA demonstrated that FNAB is an excellent tool for the diagnosis of primary and recurrent malignant neoplasms.\(^{6}\) These results were supported by a South African study of paediatric oncology patients where FNAB showed a sensitivity of 96.1% and a specificity of 100%.\(^{13}\)

FNAB is used increasingly for the diagnosis of infectious disease, particularly tuberculosis.\(^{14,15}\) Poor countries bear the brunt of paediatric tuberculosis,\(^{16}\) necessitating optimal use of limited resources. It has been reported that cervical lymphadenitis is the most common extra-thoracic manifestation of paediatric tuberculosis in India and South Africa,\(^{17-19}\) but differentiation from other infections such as \( M. \) \textit{bovis} BCG, non-tuberculous mycobacteria, fungal infections or malignancies such as Burkitt’s lymphoma or Kaposi’s sarcoma is important.\(^{20,21}\) FNAB is a minimally invasive procedure that offers a definitive tissue diagnosis and can be performed by well trained nurses on an outpatient basis. It has particular diagnostic value in HIV-infected children in whom the diagnosis of intrathoracic tuberculosis is often complicated by atypical manifestations and other HIV-associated conditions.\(^{22}\) In addition to providing rapid diagnostic confirmation by cytomorphology and/or direct visualisation of the infecting organism, FNAB samples may be inoculated into relevant culture that allows more accurate organism speciation and drug sensitivity testing.\(^{20,23}\)

This study confirms the diagnostic value of FNAB in routine paediatric practice. It also demonstrates that FNAB, using a small-gauge needle, is rapid and safe even in resource-limited settings. Well trained nurse aspirators can perform superficial aspirates as an outpatient procedure, reducing the need for hospitalisation. Cytology slides, once prepared and fixed, can be transported easily for expert evaluation elsewhere. FNAB is of particular value to countries with limited resources and a high prevalence of diseases such as tuberculosis which can be treated successfully at primary health care level, while identifying patients who require referral to a regional or tertiary care centre.

**Acknowledgments**

We are grateful to Dr M. Kidd, Department of Statistics, University of Stellenbosch, for assistance with the statistical analysis and to

---

*Fine needle aspiration biopsy* 69
Mr D. Geiger for preparing the Microsoft Excel spreadsheet.

References
Fine Needle Aspiration Biopsy - A First Line Diagnostic Procedure in Pediatric Tuberculosis suspects with Peripheral Lymphadenopathy?

Colleen A Wright, MD FRCPathª; Anneke C Hesseling, MD, PhDb; Colleen Bamford, MD, FCPathc; Steven Michael Burgess, PhDd; Rob Warren, PhDc; and Ben J Marais, MD, PhDb

ªDivision of Anatomical Pathology, Department of Pathology, University of Stellenbosch and NHLS Tygerberg Hospital, Tygerberg, South Africa

¬Desmond Tutu TB Center and/or the Department of Pediatrics and Child Health, Stellenbosch University, Tygerberg, South Africa

⁵Division of Medical Microbiology, Department of Pathology, University of Stellenbosch and NHLS Tygerberg Hospital, Tygerberg, South Africa

⁴Graduate School of Business, University of Cape Town, Cape Town, South Africa

⁶NRF Centre of Excellence in Biomedical Tuberculosis Research / MRC Centre for Molecular and Cellular Biology, Division of Molecular Biology and Human Genetics, University of Stellenbosch, South Africa

SUMMARY

OBJECTIVE To evaluate the diagnostic yield and time to diagnosis of fine needle aspiration biopsy (FNAB) versus routine respiratory specimens collected from children with a palpable peripheral lymph node mass and symptoms suspicious of tuberculosis.

DESIGN We performed a retrospective review of laboratory records at Tygerberg Hospital over a 4-year period, from January 2003 to December 2006. All children (<13 years) in whom an FNAB and other mycobacterial specimens were collected as part of their diagnostic workup were included.
RESULTS In 95 children the following specimens were collected; FNAB 95, gastric aspirates 142, other respiratory specimens 36, non-respiratory specimens 26. Mycobacterial disease was diagnosed in 70/95 (73.3%) patients. Children without respiratory specimens (n = 6) and/or with *M. bovis BCG* disease (n = 15) were excluded from comparative analysis. In the remainder, FNAB was positive in 45/74 (60.8%) versus any respiratory specimen in 29/74 (39.2%; p < 0.001). The mean time to bacteriologic diagnosis with FNAB was 7.1 days (95% CI 4.2-10.1) compared to 22.5 days (95% CI 15.8-29.1) for any respiratory specimen.

CONCLUSION FNAB is a simple, rapid and effective modality to achieve confirmation of mycobacterial disease in pediatric tuberculosis suspects with a palpable peripheral lymph node mass.

INTRODUCTION

Pediatric tuberculosis (TB) contributes significantly to the global TB disease burden.\(^1\)\(^2\) Children suffer severe TB-related morbidity and mortality in endemic areas, but contribute little to disease transmission and the maintenance of the TB epidemic.\(^1\) Consequently, treatment of children has been a low priority in global TB control efforts, but the World Health Organization (WHO) produced guidance for the management of childhood TB in 2006 calling for the evaluation of new techniques to improve the diagnosis.\(^3\) The Global Drug Fund made child-friendly TB treatment formulations available to poor countries since 2008.\(^4\)

The accurate diagnosis of pediatric TB remains a significant challenge, due to the low specificity of signs and symptoms, especially in human immunodeficiency virus (HIV)-infected children, the difficulty of obtaining bacteriologic specimens and the paucibacillary nature of pediatric disease.\(^5\) New diagnostic tests for TB have been developed, such as T cell assays and nucleic amplification tests, but these are all poorly validated in children.\(^6\) The
ability to distinguish latent infection from active disease remains problematic, particularly in endemic areas with heavy disease burdens.

An accurate bacteriologic diagnosis in children with TB is needed to initiate effective therapy and has become even more important with the transmission of multidrug-resistant (MDR) and extensively drug-resistant (XDR) TB to children. Obtaining bacteriologic specimens from children is difficult and diagnostic yield is usually low. Sputum smear microscopy is positive in less than 10-15% of children with probable pulmonary TB, with reported culture yields between 30-40%. Various methods have been described to obtain respiratory specimens from children who are unable to expectorate, with no international consensus on what is best.

TB lymphadenitis is the most common extrapulmonary manifestation of TB in children from endemic areas, accounting for almost 50% of extrapulmonary disease. Fine needle aspiration biopsy (FNAB) is the diagnostic modality of choice in children with persistently enlarged cervical nodes in TB endemic areas. FNAB is increasingly utilized in paediatric oncology patients, and multiple studies have reported the value of FNAB for TB diagnosis in adults, but it remains underutilized as a diagnostic tool in TB suspects. The aims of this study were to compare the diagnostic yield and time to diagnosis of FNAB compared to routine respiratory specimens collected in child TB suspects with a palpable peripheral lymph node mass.

**METHODS**

We performed a retrospective review of laboratory records at Tygerberg Hospital, Cape Town South Africa, from January 2003 to January 2007. Only children less than 13 years of age (eligible for access to pediatric services) were included. Since this was a laboratory-based study we were unable to review clinical information and all diagnostic tests were requested at
the discretion of the attending clinician. Standard practice at our institution is to refer children with a persistent peripheral lymph node mass (>1-2x1-2cm) for FNAB, while respiratory specimens are collected in most pediatric TB suspects.

Children who underwent FNAB were identified using laboratory records of the National Health Laboratory Services. Those in whom another specimen for mycobacterial culture was obtained within a 30 day time window either side of the FNAB collection date were included in the analysis. Cases with *Mycobacterium bovis* bacille Calmette-Guerin (BCG) disease were excluded from the comparative analysis. These children typically present with right axillary adenitis ipsilateral to the vaccination site, which is nearly pathognomonic. Inclusion of these children would have biased results in favour of FNAB.

FNAB was done in the ward or as an outpatient procedure by a trained pathologist following standard protocol. The lymph node mass was stabilized by the pathologist performing the aspiration biopsy using a 23g or 25g needle attached to a 10ml syringe, applying constant suction of no more than 2ml. Two smears were prepared from each aspirate: one fixed with commercial cytology fixative for Papanicolaou staining and the other air-dried for Giemsa and ZN staining. Thereafter the needle and syringe was rinsed by withdrawing an aliquot of liquid growth media into the syringe and discharging the contents back into a mycobacterial growth indicator tube (MGIT, Beckton Dickinson, USA). The inoculated MGIT tubes were transported to the microbiology laboratory within 2 hours, where standard PANTA supplement was added and the tubes incubated for 42 days. Positive cultures were identified as *M. tuberculosis* complex by polymerase chain reaction (PCR).

In the cytology laboratory, smears were evaluated for adequacy, defined as sufficient cells and/or necrotic material to allow a definitive diagnosis. A modified ZN stain was used on the Giemsa stained smears to detect acid fast bacilli. Fluorescence microscopy was carried out on Papanicolaou stained smears using a Zeiss Axiophot microscope with a
fluorescent attachment and a wide-band blue excitation filter (450–480 nm). Papanicolaou stained mycobacteria fluoresce as brilliant yellow bacilli, thin and slightly curved with polar enhancement and a length of 2.0 –2.7 microns.\textsuperscript{26} \textit{M. tuberculosis}, \textit{M. bovis}, \textit{M. bovis} BCG and non-tuberculuous mycobacteria (NTM) are morphologically indistinguishable, but in this TB-endemic setting \textit{M. bovis} is hardly ever detected on culture, \textit{M. bovis} BCG usually has a clinical pathognomonic presentation,\textsuperscript{27} and disease caused by NTM is relatively rare.

Early morning gastric aspirates were obtained following overnight admission to the pediatric ward. A nasogastric tube was passed into the stomach and the contents aspirated; 5-10 ml of normal saline inserted if required until a total volume of at least 5ml was aspirated. Gastric aspirates were placed in a sterile tube and neutralized with an equal volume of sodium carbonate. Induced sputa and/or nasopharyngeal aspirates were collected by a trained physiotherapist, but this was rarely done. Sputum induction was undertaken after 3 hours of fasting. Children were pre-treated with 200ug of salbutamol via metered dose inhaler and spacer. A jet nebulizer attached to oxygen at a flow rate of 5 l/min delivered 5 ml of 5% sterile saline for 15 minutes. Thereafter chest physiotherapy was applied before obtaining sputum by expectoration or by nasopharyngeal suctioning, using a mucus extractor. Oropharyngeal suction was used if the child had excessive nasal secretions.

All microbiology specimens were submitted to the laboratory within 6 hours. Smear microscopy, using either Ziehl-Neelsen or auramine stains, was only performed on expectorated sputum samples. Following N-acetyl-L- cysteine - sodium hydroxide decontamination, sputum specimens were inoculated into MGIT and incubated for 42 days. Further procedures for identification of mycobacteria were as described for the FNAB samples. All procedures were carried out according to standard operating procedures in a Biosafety Level 2 laboratory.
The reference standard for disease on FNAB was cytomorphology consistent with mycobacterial infection plus mycobacteria visualized with ZN stain and/or autofluorescence, and/or if \textit{M. tuberculosis} was identified on culture. Given the low rates of NTM in the study population culture negative cases with cytomorphological and mycobacterial proof of disease were included in the TB group. For respiratory specimens the reference standard was acid fast bacilli (AFB) detected on sputum smear microscopy or identification of \textit{M. tuberculosis} by culture in at least one specimen. For comparative purposes multiple respiratory specimens were regarded as a single test procedure, since we aimed to compare the value of routine respiratory specimen collection as a diagnostic procedure to FNAB.

We assessed whether the comparative bacteriological yield of FNAB (cytology and/or culture) was higher than other methods using Pearson’s chi-square ($X^2$) and Fisher’s exact tests. All analyses were conducted using Statistica Version 8. Ethics approval was obtained from the Institutional Review Board of Stellenbosch University (N08/09/240).

**RESULTS**

FNAB and additional mycobacterial specimens were collected from 95 children, ranging in age from 2 months to 11 years. Figure 1 provides a flow diagram of patients and specimens included in the study. There was no significant difference in gender distribution (49 male vs. 46 females) and 48% of children were less than one year of age. The HIV-infection status was unknown in 28 (29.5%) children while 33 (34.7%) were HIV-infected and 34 (35.8%) were HIV-uninfected. Mycobacteria were identified in 70 (73.7%) children. Table 1 reflects the biopsy site, bacteriologic yield and mycobacterial species identified on FNAB; no NTM were identified.

Table 2 reflects the bacteriologic yield and mycobacterial species identified for all specimen types; of the 57 specimens speciated 41 (71.9%) were \textit{M. tuberculosis}, 15 (26.3%)
M. bovis BCG and 1 (1.8%) NTM. A total of 143 gastric aspirates were collected from 80 (84.2%) children, with only 11 children (13.8%) submitting the desired 3 specimens. Additional respiratory specimens were collected in 15 patients; of whom 7 (mean age 9.3 years) provided expectorated sputum. Non-respiratory specimens, including cerebrospinal fluid, pus swabs and blood, were collected in 21 patients (26 specimens). All non-respiratory specimens, including 6 patients in whom no respiratory specimens were collected, and M. bovis BCG cases were excluded from the comparative analysis. M. bovis BCG was isolated in 15 children; in 2 cases the site of aspiration was soft tissue rather than an axillary lymph node. One third of these children (5 cases) had distant or disseminated BCG disease as evidenced by the isolation of M. bovis BCG from gastric and/or nasopharyngeal aspirates. All were immunocompromised: 4 were HIV-infected and the other had Severe Combined Immune Deficiency.

Of the 45 patients diagnosed with TB using the defined FNAB reference standard, 36 (80.0%) were confirmed to be M. tuberculosis on culture. Of the 30 diagnosed from respiratory specimens, 29 (96.7%) were confirmed to be M. tuberculosis on culture and 1 adolescent child was sputum smear (AFB) positive. For comparative purposes the collection of multiple respiratory specimens was regarded as a single test procedure. The bacteriological yield of FNAB compared to any combination of respiratory specimens is reported in Table 3; being higher compared to gastric aspirates (p < 0.001) and all respiratory specimens combined (p < 0.001).

We also assessed differences in the mean time to diagnosis for FNAB versus the other specimen types (where time to diagnosis is time to bacteriologic confirmation from onset of the test procedure). Whether the timing of additional mycobacterial cultures occurred before, after or simultaneously with FNAB collection did not influence the significance of the yield or time to diagnosis comparisons. Collecting multiple respiratory specimens was regarded as
a single test procedure. Accepting an average time to microscopy diagnosis of 2 days, FNAB (7.1 days, 95% confidence interval = 4.2 - 10.1 days) requires less than one-third the time of respiratory cultures (22.5 days, 95% CI = 15.8 – 29.1 days). This difference is illustrated in Figure 2. The one outlier is the child whose TB was confirmed by sputum smear-microscopy. The mean time to diagnosis for gastric aspirate culture (the most commonly performed test) was 21.4 days (95% CI = 15.2 – 27.6 days).

**DISCUSSION**

In the current study FNAB provided a better yield in child TB suspects with palpable peripheral lymph node masses than the collection of multiple respiratory specimens. Children referred with suspected mycobacterial lymphadenitis routinely have a tuberculin skin test (TST) performed and return to the clinic or hospital for this to be read within 48–72 hours. If FNAB is performed with the initial visit, microscopy results should be available at the time of the return visit 48-72 hours later, enabling the commencement of TB therapy pending the results of culture and speciation.

The true extent of the pediatric TB disease burden is poorly documented, since more than 80% of children with TB are sputum smear-negative and direct smear microscopy is often the only diagnostic modality available in countries with limited resources. The diagnostic dilemma is further compounded by the HIV pandemic, HIV-associated disease and TB may have common clinical and radiological presentations, in addition tests for TB infection such as the TST and newer T-cell assays have poor sensitivity in immunocompromised children. Two to three fasting gastric aspirates or a single induced sputum specimen have traditionally been advised for bacteriological confirmation of TB. However, the yields observed in everyday practice deviate considerably from those achieved under trial conditions, which emphasizes the value of “real-life” retrospective analysis.
The low number of patients in whom gastric aspirates were collected on 3 consecutive days reflects practical difficulties. Severe limitations of in-patient beds and staff and the reluctance of parents to remain in hospital with their children for more than 1-2 days, since they often have other children at home to care for. In 5 children induced sputa and/or nasopharyngeal aspirates were collected as part of a separate specimen collection study, comparing the yield of gastric aspirates, nasopharyngeal aspirates and induced sputum. We included these specimens to provide a comprehensive overview of all the respiratory specimens collected. Inclusion of these specimens may have introduced bias, but numbers were small and did not influence the outcome. If anything these additional specimens would have favored the bacteriological yield of routine respiratory specimens. Development of improved respiratory specimen collection techniques that can be performed on an outpatient basis require further exploration. However our study demonstrates the value of FNAB as an outpatient diagnostic procedure in TB suspects with a palpable peripheral lymph node mass.

Enlarged peripheral lymph nodes provide a valuable opportunity to establish a definitive diagnosis of TB in HIV-infected children. The majority of HIV-infected children with persistent lung disease have concurrent persistent peripheral lymphadenopathy, which provides a neglected sample collection opportunity in this problematic group. In a recent South African study, 22 of 95 FNAB performed in mass lesions in HIV–infected children were positive for mycobacterial infection. FNAB has been established as an easy and reliable outpatient procedure for the diagnosis of palpable superficial masses in children, and is ideally suited for use in resource limited settings. It is simple, inexpensive, requires limited infrastructure and can be taught to medical and nursing staff. More widespread utilization is limited by a lack of experienced pathologists, but slides are easily transportable to regional or even international centers for diagnosis.
If the correct technique is followed, it is associated with minimal complications. There were no adverse events or complications recorded with any of the FNAB’s performed during this study. In a previous study we demonstrated that using FNAB with cytomorphology, autofluorescence and culture, provided a rapid and definitive diagnosis in the vast majority of children. Culture alone produced a significantly lower yield, since it is influenced by factors such as inefficient sampling, contamination, variable bacterial load, and prior commencement of TB therapy. Rapid microscopy confirmation was achieved in 34/45 (75.5%) patients ultimately diagnosed with *M. tuberculosis* using a combination of microscopy and/or culture positivity. Previous FNAB studies from this TB endemic setting, as well as the current study, demonstrated no NTM disease, while *M. bovis BCG* was only cultured in children less than 2 years age with a clinically pathognomonic presentation. Although rapid microscopy diagnosis allows a fairly accurate diagnosis and permits immediate commencement of therapy, culture is advisable as this enables speciation and drug susceptibility testing.

Compared to conventional respiratory specimens FNAB provided superior diagnostic yields and a significant reduction in time to TB diagnosis. Rapid bacteriological diagnosis has important benefits for patient management, also in settings not endemic for TB where NTM adenitis may be more common. This study reviewed laboratory records only, therefore, we are unable to provide detailed clinical descriptions or explore potential differences in patient characteristics that may have influenced the sequencing and/or yield of various diagnostic tests. However, it provides an assessment of every day (“real life”) practice without any intentional bias. Despite these limitations, we believe FNAB should be regarded as a first line diagnostic modality in child TB suspects with a palpable peripheral lymph node mass.
ACKNOWLEDGEMENTS

The authors would like to thank Professor Martin Kidd (Stellenbosch University) for statistical analysis. This study is in partial fulfillment of a PhD thesis.
REFERENCES


### Table 1

**Mycobacteria isolated on culture and site of fine needle aspiration biopsy (FNAB)**

<table>
<thead>
<tr>
<th>Mycobacteria isolated&lt;sup&gt;a&lt;/sup&gt;</th>
<th>Number (%)</th>
<th>Site of FNAB</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Axillary node</td>
</tr>
<tr>
<td></td>
<td></td>
<td>R</td>
</tr>
<tr>
<td><strong>All</strong></td>
<td>51/95 (54)</td>
<td>11</td>
</tr>
<tr>
<td></td>
<td></td>
<td>8</td>
</tr>
<tr>
<td><strong>M. Tuberculosis</strong></td>
<td>36/51 (71)</td>
<td>5</td>
</tr>
<tr>
<td></td>
<td></td>
<td>8</td>
</tr>
<tr>
<td><strong>M. bovis BCG</strong></td>
<td>15/51 (29)</td>
<td>6</td>
</tr>
<tr>
<td></td>
<td></td>
<td>0</td>
</tr>
</tbody>
</table>

<sup>a</sup> No M. bovis or Non Tuberculous Mycobacteria (NTM) were isolated

<sup>b</sup> 13 (87%) of children <1 year of age
Table 2
Specimen type, bacteriological yield and organisms cultured in 95 patients

<table>
<thead>
<tr>
<th>Specimen type</th>
<th>Total number</th>
<th>Consecutive specimens No (%)</th>
<th>Culture positive No (%)</th>
<th>M. <em>tb</em>&lt;sup&gt;a&lt;/sup&gt; No (%)</th>
<th>BCG&lt;sup&gt;b&lt;/sup&gt; No (%)</th>
<th>NTM&lt;sup&gt;c&lt;/sup&gt; No (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>FNAB</td>
<td>95</td>
<td>95 0 0</td>
<td>51 (54)</td>
<td>36 (71)</td>
<td>15 (30)</td>
<td>0</td>
</tr>
<tr>
<td>Gastric aspirates</td>
<td>143</td>
<td>80 52 (65) 11 (14)</td>
<td>39 (27)</td>
<td>30 (77)</td>
<td>8 (21)</td>
<td>1 (3)</td>
</tr>
<tr>
<td>Induced sputum</td>
<td>15</td>
<td>10 5 (50) 0</td>
<td>6 (40)</td>
<td>6 (100)</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>Expectorated sputum</td>
<td>11</td>
<td>7 3 (43) 1 (14)</td>
<td>5 (45)</td>
<td>5 (100)</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>Nasopharyngeal aspirates</td>
<td>10</td>
<td>7 3 (43) 0</td>
<td>5 (50)</td>
<td>4 (80)</td>
<td>1 (20)</td>
<td>0</td>
</tr>
<tr>
<td>Non–respiratory</td>
<td>26&lt;sup&gt;d&lt;/sup&gt;</td>
<td>26 0 0</td>
<td>3 (12)</td>
<td>2 (67)&lt;sup&gt;e&lt;/sup&gt;</td>
<td>1 (33)&lt;sup&gt;f&lt;/sup&gt;</td>
<td>0</td>
</tr>
</tbody>
</table>

<sup>a</sup>*M. tb* – *Mycobacterium tuberculosis*;
<sup>b</sup>BCG – *Mycobacterium bovis* bacilli Calmette-Guerin;
<sup>c</sup>NTM – Non-tuberculous mycobacteria
<sup>d</sup>21 patients, pus swabs, urine, CSF, blood cultures;
<sup>e</sup>Cerebrospinal fluid;
<sup>f</sup>Pus swab.
Table 3
Bacteriologic yield of fine needle aspiration biopsy (FNAB) compared to routine respiratory specimens (*M. bovis* BCG excluded)

<table>
<thead>
<tr>
<th>Routine respiratory specimens</th>
<th>FNAB&lt;sup&gt;a&lt;/sup&gt; negative</th>
<th>FNAB&lt;sup&gt;a&lt;/sup&gt; positive</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td>All gastric aspirates negative</td>
<td>25</td>
<td>19</td>
<td>44</td>
</tr>
<tr>
<td>Any gastric aspirate positive</td>
<td>3</td>
<td>21</td>
<td>24</td>
</tr>
<tr>
<td>Total</td>
<td>28</td>
<td>40</td>
<td>68</td>
</tr>
<tr>
<td>All respiratory specimens&lt;sup&gt;b&lt;/sup&gt; negative</td>
<td>26</td>
<td>19</td>
<td>45</td>
</tr>
<tr>
<td>Any respiratory specimen positive</td>
<td>3</td>
<td>26</td>
<td>29</td>
</tr>
<tr>
<td>Total</td>
<td>29</td>
<td>45</td>
<td>74</td>
</tr>
</tbody>
</table>

<sup>a</sup>Cytology and/or culture

<sup>b</sup>Includes gastric and nasopharyngeal aspirates, as well as sputum and induced sputum specimens
Figure 1
Breakdown of patients and specimens included in the analysis

95 patients with FNAB* and another specimen

143 gastric aspirates
36 other respiratory specimens
26 non-respiratory specimens (excluded)

Excluded

15 patients with M. bovis BCG
6 patients, non-respiratory specimens only

74 patients with FNAB* and another respiratory specimen

68 patients with gastric aspirates
12 patients with other respiratory specimens
74 patients with FNAB*

*FNAB – Fine needle aspiration biopsy
Figure 2
Time to diagnosis Fine Needle Aspiration Biopsy (FNAB) versus All respiratory specimens

Y-axis – specify units “Days”
X-axis – only “FNAB” & “All respiratory”
CHAPTER 5

OPTIMISING CYTOPATHOLOGICAL DIAGNOSIS

Mycobacterial Autofluorescence in Papanicolaou-Stained Lymph Node Aspirates: a Glimmer in the Dark?
Colleen A Wright, Yvonne van Zyl, Steven M Burgess, Lucille Blumberg, Gladwyn Leiman.

Diagnosing Mycobacterial Lymphadenitis in Children Using Fine Needle Aspiration Biopsy: Cytomorphology, ZN Staining and Autofluorescence-making more of less.
Colleen A Wright, Steven M Burgess, Mirjam van der Burg, D Geiger, Jeroen G Noordzij, Ben J Marais
Diagnostic Cytopathology: 2008; 36: 245-251
Mycobacterial Autofluorescence in Papanicolaou-Stained Lymph Node Aspirates: A Glimmer in the Dark?

Colleen A. Wright, FRCPath,1* Yvonne van Zyl, C.T.1 Steven M. Burgess, Ph.D.,2 Lucille Blumberg, M.D.,3 and Gladwyn Leiman, FRCPath1

This study was undertaken to determine the value of incorporating fluorescence into cytopathological evaluation of lymph node fine-needle aspiration (FNA) specimens suspected of harboring mycobacterial species. The study population consisted of 1,044 HIV-positive and -negative patients referred for FNA to the cytopathology unit of a South African medical school located in a very high HIV prevalence region. Each aspirate was assessed on routine Papanicolaou-stained slides for morphologic characteristics of mycobacterial infection. The same glass slides were then viewed under fluorescent microscopy to determine the presence or absence of mycobacterial autofluorescence. Using multivariate analysis, results of both cytology and fluorescence were compared with mycobacterial culture as the final arbiter of the presence of organisms. In this large clinical study, compared with culture, cytomorphology showed sensitivity of 84.9%, but low specificity of only 50.9%. Fluorescence demonstrated lower sensitivity of 65.9%, but improved specificity of 73.0%. Taken together, positivity of both cytology and fluorescence improved specificity to 81.8%. Fluorescent microscopy is rapid, inexpensive, and cost-effective; neither radioactive materials nor further staining are required. It is felt that this methodology would be of diagnostic benefit if used on morphologically suspicious samples in areas with a high prevalence of HIV and mycobacterial infections. Appropriate therapy could be commenced within hours of FNA, with reduction in the current number of patients lost to follow-up while awaiting results of culture. The technique is readily extended to other FNA types such as deep organ aspirates. Autofluorescence of organisms specifically requires usage of Papanicolaou staining; the technique cannot be used in histopathologic specimens stained with hematoxylin-eosin. Diagn. Cytopathol. 2004;30:257–260. © 2004 Wiley-Liss, Inc.

Key Words: mycobacteria; tuberculosis; fluorescence; cytodiagnosis; HIV

The human immunodeficiency virus (HIV) epidemic in South Africa has attracted considerable attention. Prevalence figures in the general population are unknown, but 24.5% of women attending antenatal clinics nationwide are HIV-positive; the corresponding figure for the province in which this study was undertaken is 29.4% (Department of Health National HIV and Syphilis seroprevalence survey of women attending public antenatal clinics in South Africa, 2000; www.doh.gov.za/docs/reports/2000/hivreport.html). It is estimated that more than 60% of medical admissions in academic hospitals are due to opportunistic infections in patients with acquired immunodeficiency disease (AIDS). The most common of these opportunistic infections in South African adults with AIDS is Mycobacterium tuberculosis (MTB). In the year 2000, 150,696 new MTB cases were reported to the national registry, including 31,058 extrapulmonary infections (personal communication, Tuberculosis Directorate, SA National TB Control Program). By 2001, MTB incidence in the general population was approximately 300 per 100,000; at least 50% of new MTB cases were in retroviral-positive patients (personal communication, SA National Department of Health). Dual HIV and

1Cytopathology Unit, School of Pathology of the South African Institute for Medical Research and University of the Witwatersrand, Johannesburg, South Africa
2Department of Commerce, University of the Witwatersrand, Johannesburg, South Africa
3Department of Microbiology, School of Pathology, University of the Witwatersrand, Johannesburg, South Africa

Colleen A. Wright’s current address is Department of Anatomical Pathology, Stellenbosch University, Cape Town, South Africa.

Steven M. Burgess’s current address is Graduate School of Business, University of Cape Town, Cape Town, South Africa.

Lucille Blumberg’s current address is National Institute for Communicable Diseases, National Health Laboratory Services, Johannesburg, South Africa.

Gladwyn Leiman’s current address is Department of Pathology, University of Vermont, Burlington, Vermont.

The South African Institute for Medical Research is now the National Health Laboratory Service of South Africa.

*Correspondence to: Colleen A. Wright, Department of Anatomical Pathology, University of Stellenbosch, P.O. Box 19063, Tygerberg 7505, South Africa. E-mail: cawr@gerga.sun.ac.za

Received 30 May 2003; Accepted 29 September 2003

DOI 10.1002/dc.20009

Published online in Wiley InterScience (www.interscience.wiley.com).
MTB epidemics place overwhelming demands on health care resources in any developing country. Expected life span is reduced by AIDS (down from 68 years to 48 years in South Africa between 1990 and 2000).\textsuperscript{1,2} By dominating hospital admissions, the HIV/MTB population limits investigation and treatment of non-HIV patients with potentially curable disease entities compatible with normal longevity. MTB diagnosis may be challenging, even in high prevalence areas; it is modified by HIV status and compounded by atypical symptoms, overlap in presentation of other opportunistic infections, frequency of extrapulmonary involvement, and rapid progression.\textsuperscript{3}

Traditionally, laboratory diagnosis of mycobacterial infection has been the purview of the microbiologist; it has relied on positive culture or microscopic recognition of the organism by Ziehl-Nielsen staining.\textsuperscript{4} Fluorescence microscopy using auramine-rhodamine was described as early as 1937 and generally has been shown to be superior to Ziehl-Nielsen staining.\textsuperscript{5–7} but involves the use of toxic and carcinogenic substances. A substantial benefit would derive from any new, inexpensive, exposure-free diagnostic procedure, enabling early commencement of anti-MTB treatment. Mycobacteria (as well as \textit{Pneumocystis carinii}, fungi, and certain bacteria) show brilliant fluorescence in Papanicolaou-stained smears.\textsuperscript{8–10} The method is quick and inexpensive and can be focussed on those specimens felt to harbor the above infections on morphologic grounds. This suggests that fluorescence microscopy of Papanicolaou-stained smears could provide a rapid, safe, and inexpensive technique for confirmation of mycobacterial infection in cytological specimens. The present study explores this opportunity by investigating the ability of cytomorphology and fluorescence microscopy of Papanicolaou-stained smears from superficial lymph nodes obtained by fine-needle aspiration (FNA), individually and together, to identify mycobacterial infections.

Materials and Methods

A prospective study was performed in the teaching hospitals of the University of the Witwatersrand in Johannesburg, South Africa, during the period January 1998 to December 2001. The sample consisted of 1,044 superficial lymph node aspirates from patients with clinically suspected mycobacterial lymphadenopathy. In this center, retroviral serology is not performed without patient consent; serology was thus not available in many cases. In the 330 cases in which retroviral status was known, 318 (96%) were positive; a high proportion of the remaining patients were clinically suspected to be retrovirus-positive. Patients were referred to FNA clinics operated by the cytopathology unit at four teaching hospitals. Procedures were performed using 22 gauge needles attached to 10 ml syringes. At least two needle passes were performed on each node aspirated. Direct smears were made and were either air-dried for Diff-Quik staining or fixed with commercial spray fixative for Papanicolaou staining. Onsite rapid staining was not undertaken. The needle was rinsed in BACTEC 12B medium (Becton-Dickinson, Mountain View, CA), a commercially manufactured mycobacterial culture system. Slides were returned to the central cytopathology laboratory for same-day processing.

The air-dried Diff-Quik-stained and alcohol-fixed Papanicolaou-stained smears were screened and evaluated for adequacy and content. All neoplastic samples were excluded from the study, as were inadequate slides and those showing benign nonreactive, noninflammatory morphology. The remainder were reported as cytologically suggestive (positive) or nonsuggestive (negative) of mycobacterial lymphadenitis. This cytomorphologic evaluation was made on previously established criteria: amorphous granular necrotic debris, epithelioid histiocytes, and multinucleated Langhans-type giant histiocytes, in a background population of reactive lymphocytes and plasma cells. In the presence of AIDS, the cytopathological picture is more frequently that of necrosis only, or necrosis associated with an acute inflammatory infiltration, without attendant lymphocytic or histiocytic components.

Without any additional processing, the Papanicolaou-stained smears were then screened by one author (Y.v.Z.) using an Olympus BX40 microscope with a fluorescent attachment and the following filters: wide-band blue excitation filter (450–480 nm), dichromatic splitter (500 nm), and barrier filter (515 nm). Slides were viewed at ×400 magnification. This displayed the MTB organisms as brilliant yellow fluorescent rods, which were thin and slightly curved with light ends and a uniform length of 2.0–2.7 microns (Fig. C-1). \textit{Mycobacterium tuberculosis} is differentiated from \textit{Mycobacterium avium}, which is shorter (1.2–1.8 microns) and variably fluorescent.
Table I. Comparative Results of MTB Detection by Cytodiagnosis and by Culture*

<table>
<thead>
<tr>
<th>Cytodiagnosis</th>
<th>Negative</th>
<th>Positive</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td>Negative</td>
<td>228 (22%)</td>
<td>90 (8%)</td>
<td>318 (31%)</td>
</tr>
<tr>
<td>Positive</td>
<td>220 (21%)</td>
<td>506 (49%)</td>
<td>726 (70%)</td>
</tr>
<tr>
<td>Total</td>
<td>448 (43%)</td>
<td>596 (57%)</td>
<td>1,044 (100%)</td>
</tr>
</tbody>
</table>

*Sensitivity, 84.9%; specificity, 50.9%; negative predictive value, 71.7%; positive predictive value, 69.7%; accuracy, 69.3%.

Table II. Comparative Results of Detection of MTB by Autofluorescence and by Culture*

<table>
<thead>
<tr>
<th>Fluorescence</th>
<th>Negative</th>
<th>Positive</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td>Negative</td>
<td>327 (31%)</td>
<td>203 (19%)</td>
<td>530 (50%)</td>
</tr>
<tr>
<td>Positive</td>
<td>121 (12%)</td>
<td>393 (38%)</td>
<td>514 (50%)</td>
</tr>
<tr>
<td>Total</td>
<td>448 (43%)</td>
<td>596 (57%)</td>
<td>1,044 (100%)</td>
</tr>
</tbody>
</table>

*Sensitivity, 65.9%; specificity, 73.0%; negative predictive value, 67.7%; positive predictive value, 76.5%; accuracy, 69.3%.

Table III. Summary of Statistical Analyses for Detection of MTB by Cytodiagnosis and by Autofluorescence, Separately and Together (%)

<table>
<thead>
<tr>
<th></th>
<th>Cytodiagnosis</th>
<th>Autofluorescence</th>
<th>Both</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sensitivity</td>
<td>84.9</td>
<td>65.9</td>
<td>56.7</td>
</tr>
<tr>
<td>Specificity</td>
<td>50.9</td>
<td>73.0</td>
<td>81.8</td>
</tr>
<tr>
<td>Negative predictive value</td>
<td>71.7</td>
<td>61.7</td>
<td>58.7</td>
</tr>
<tr>
<td>Positive predictive value</td>
<td>69.7</td>
<td>76.5</td>
<td>80.5</td>
</tr>
<tr>
<td>Accuracy</td>
<td>69.3</td>
<td>69.3</td>
<td>69.4</td>
</tr>
</tbody>
</table>

Results

Retroviral status (where known) and the diagnostic outcome of each of the three tests (i.e., cytomorphology, fluorescence, and culture) were code-captured in a standard portable computer spreadsheet using a standard indicator coding scheme (1 = positive; 0 = negative). The laboratory’s unique case reference number was also captured for quality control purposes. Detailed results appear in Tables I–III. The results indicate a strong and highly significant association between cytology and culture diagnoses (Cramer’s V = 0.59; P < 0.001) and a virtually equivalent association between fluorescence and culture (Cramer’s V = 0.57; P < 0.001). Thus, the overall accuracy of the techniques, separately and together, is virtually identical at almost 70%. Cytology and fluorescence offer differing benefits as regards sensitivity and specificity. Cytology is suggestive of 84.9% of culture-positive cases, yields few false negatives, but has low specificity of 50.9%. Fluorescence has lower sensitivity of 65.9%, yields few false positive readings, and thereby raises specificity to 73.0%. Taken together, cytology and autofluorescence refine the diagnostic group, incorporating lower sensitivity at 56.7%, but significantly improving specificity to 81.8%. Clinical implications are that three of five TB patients undergoing FNA could be placed on appropriate and immediate antituberculous therapy. At a later stage of patients so medicated, one of four would not be confirmed on traditional culture as harboring active mycobacterial infection.

Discussion

The diagnosis of tuberculosis by cytomorphology is not new. It is a necrotizing granulomatous infection, which cytopathologically demonstrates the microscopic equivalent of caseous necrosis, i.e., a granular-appearing necrotic background, together with mature lymphocytes, tangles of epithelioid histiocytes, and giant multinucleated Langhans-type histiocytes. This cytopathological appearance, however, is neither sensitive nor specific for MTB. In recent years, the advent of HIV-associated tuberculosis has altered the characteristic cytopathologic picture, mainly by paucity or even absence of cellular response, i.e., fewer epithelioid and multinucleated histiocytes. Recent literature has shown that FNAs, particularly from large nodes with necrotic centers, may contain no cellular response at all, i.e., no epithelioid or multinucleated histiocytes, and may even lack lymphocytes if the entire node has undergone necrosis. Aspirates may then be entirely necrotic or may contain debris together with a secondary influx of neutrophils. The cytology then resembles an abscess more than it does classic tuberculosis. However, in high-incidence regions, sufficient experience has now been gained with new and varied presentations of tuberculosis in lymph node aspirates to raise the cytological suspicion of MTB in appropriate clinical settings. Ziehl-Nielsen staining may be performed on FNAs; while very specific, this test lacks sensitivity, as organisms may be extremely sparse and difficult to visualize. Therefore, it has become routine to submit needle rinses or separate needle passes in appropriate media to microbiology, where direct microscopy is performed, together with traditional culture. If all tests are negative prior to culture becoming positive, definitive diagnosis may be delayed for up to 6 weeks. In a country or region in which MTB is so prevalent, treatment delays of this nature are unacceptable, as they result in continued presence of infected and infectious patients in their communities, where further spread is likely.

To alleviate this, fluorescence, utilizing fixed Papanicolaou-stained cytologic material, becomes appealing for rapid and cost-effective diagnosis of MTB. When judged against traditional culture, the sensitivity in this study of cytodagnosis (84.9%), and that of autofluorescence of Papanicolaou-stained material (65.9%), may not reach levels of sensitivity as attained in the diagnosis of malignancy by FNA. These sensitivity values are, however, higher than those quoted by Kupper et al. in their work on Mycobacterium kansasii. In terms of providing clinicians in overcrowded hospital settings with same-day working diagnoses on which immediate antituberculous treatment can be commenced, it is our feeling that the routine addition of fluo-
rescence to microscopy in FNAs has much to offer. It can be regarded as a one-stop test, performed if necessary on one glass slide, seen in one laboratory unit, with the final report available within hours of the FNA procedure. Diagnosis is accomplished without reprocessing from transport medium for direct evaluation and for culture, as occurs in microbiology. It also avoids the use of radioactive substances that, as indicated previously, carries some degree of morbidity for laboratory workers. The percentage of cytology/fluorescence positive patients subsequently found to be culture-negative (18.2% of those who might be placed on anti-MTB therapy using this dual positivity as an indication; 8% of the entire group referred for FNA) would appear to justify the immediate commencement of such therapy, if clinically consistent. Antituberculous therapy can be stopped if culture is found to be negative, or if clinicoradiological assessment deems the disease process not to be tuberculous in nature.

Culture was used in this study as the final arbiter of MTB infection. However, while it is the most accurate means of diagnosis available, it too is beset by an irreducible false negative rate. It is thus possible that some patients who are positive by cytology and/or fluorescence in this study, but negative by subsequent culture, did in fact have tuberculosis. Only a long-term prospective clinicoradiologic and cytopathologic study incorporating review of patient records and ultimate response to therapy over time could answer this dilemma. An investigation of that nature is outside the capabilities of this laboratory-based study.

There are ways in which the sensitivity of diagnostic fluorescence could be improved. A recent study utilized automated Papnet screening technology for the identification of small rare mycobacterial organisms stained by the auramine-rhodamine method. However, for most laboratories, an innovation of that type would be financially unattainable. On the other hand, the mere addition of careful dark-field illumination, searching for fluorescence as reported here, is feasible in most laboratories.

Compared with culture techniques, fluorescence yielded a number of false positive cases in this study. Bacterial forms other than MTB are known to exhibit autofluorescence; it is thus possible that incorrect identification of these very small organisms will be made. Kupper et al. has suggested that a minimum number of eight organisms be required for positive diagnosis. In our experience in retroviral positive patients, MTB organisms are profuse and theoretically would thus possible that incorrect identification of these very small organisms will be made. Kupper et al. has suggested that a minimum number of eight organisms be required for positive diagnosis. In our experience in retroviral positive patients, MTB organisms are profuse and theoretically would be easier to identify. The fluorescent staining properties of Mycobacterium avium intracellulare are more variable than those of MTB. In 596 culture-positive cases in this study, only 6 were mycobacteria other than Mycobacterium tuberculosis. There is, therefore, no replacement for experience and familiarity with the morphology of the organisms and scrupulous attention to detail when evaluating fluorescent bacteria.

It cannot be sufficiently emphasized that fluorescence of organisms is appropriate only to material stained with the Papanicolaou stain. The results of the above study cannot, for example, be transferred to cell block or histopathologic sections stained with hematoxylin-eosin. The particular ingredient of the Papanicolaou stain that is thought to permit autofluorescence is most likely EA50 or EA65. Because of the capacity of the Papanicolaou stain to permit autofluorescence, cytopathologists find themselves in the unique position of extending the usual capability of cytdiagnosis to include identification of fluorescent organisms, such as Mycobacterium tuberculosis, Pneumocystis, and Cryptococcus. In high-incidence areas of tuberculosis and of AIDS, this method may prove to be diagnostically useful on a scale larger than previously anticipated.

Acknowledgments

The authors thank Mrs. Lily Battaglia of the library at the South African Institute for Medical Research (now National Health Laboratory Service) for her ready and invaluable assistance in locating scarce reports on the statistics of communicable disease in South Africa.

References

Diagnosing Mycobacterial Lymphadenitis in Children Using Fine Needle Aspiration Biopsy: Cytomorphology, ZN Staining and Autofluorescence—Making More of Less

Colleen A. Wright, F.R.C.Path.,1* Mirjam van der Burg, Ph.D.,1 D. Geiger, M.Sc.,1 Jeroen G. Noordzij, Ph.D.,2† Steven M. Burgess, Ph.D.,3 and Ben J. Marais, Ph.D.2

Although the incidence of TB has stabilized or declined in most world regions, it is increasing in Africa, Southeast Asia, and the Eastern Mediterranean, fuelled by the HIV pandemic. More than 4,000 people died daily from TB-related illnesses in 2005. TB is a major cause of childhood morbidity and mortality in these developing countries, and there is an urgent need for rapid and definitive modalities for mycobacterial diagnosis in children. This prospective study in Tygerberg Hospital, Cape Town, South Africa, evaluates the ability of fine needle aspiration biopsy (FNAB) to diagnose mycobacterial lymphadenitis in children, using cytomorphology, autofluorescence on Papanicolaou stained smears, Ziehl-Nielsen (ZN) staining and/or culture. FNABs were performed on 200 children, and 25 (12.5%) aspirates were inadequate. Cultures were positive in 79/175 (45%); Mycobacterium tuberculosis was identified in 61 and Mycobacterium bovis BCG in 18 cases. Using culture as the gold standard, the concordance of the different techniques was as follows: cytology 70%, ZN staining 73%, and autofluorescence 68%. Using an alternative gold standard (culture positive and/or suggestive cytology plus positive autofluorescence or ZN smear), the “true” diagnostic performance of the various techniques was as follows: cytology—sensitivity 78%, specificity 91%, positive predictive value (PPV) 93%, ZN staining—sensitivity 82%, specificity 97%, PPV 97%; autofluorescence—sensitivity 67%, specificity 97%, PPV 97%; and culture—sensitivity 75%, specificity 100%, and PPV 100%. FNAB was shown to provide a rapid and definitive diagnosis in the majority of cases of suspected tuberculous lymphadenitis in children, based on cytomorphology and identification of the organism. Diagn. Cytopathol. 2008;36:245–251. © 2008 Wiley-Liss, Inc.

Key Words: fine needle aspiration biopsy; children; mycobacteria; tuberculosis; lymph node; HIV

Tuberculosis (TB) was declared a global emergency by the World Health Organization (WHO) in 1993. Since then, the incidence of TB has stabilized or declined in most world regions, except Africa, Southeast Asia, and the Eastern Mediterranean, where the impact of the pandemic is staggering. Between 2000 and 2005, new TB cases increased worldwide by 6%, to more than 24,000 new TB cases daily. More than 84% of these cases were located in sub-Saharan Africa and Asia. Even though TB-related deaths declined by 11% between 2000 and 2005, more than 4,000 people died daily from TB-related illnesses in 2005. It is fair to say that TB remains an emergency in those countries that can least afford the costs of diagnosing and treating its victims.

TB is a major cause of childhood morbidity and mortality in developing countries. Children below 3 years of age are particularly vulnerable because their immature immune systems make them susceptible to more severe disease manifestations, such as miliary TB and TB meningitis.

In 1989, the WHO estimated the worldwide incidence of TB in children to be 1.3 million per year with 450,000 children dying from TB-related illnesses in that year. In 2000 there were 8.3 million new cases of TB estimated worldwide, with 11% occurring in children younger than 15 years of age. As alarming as these estimates are, they...
may grossly underestimate the incidence of TB in children because the WHO estimates are based on smear positive cases and more than 80% of children with TB are not smear positive.9 In South Africa, as many as 40% of all cases of TB occur in children.3

The diagnosis of TB in children is particularly challenging for three reasons.10–12 First, sputum and alternative specimens (e.g., gastric aspirates) are difficult to obtain and have a poor diagnostic yield (i.e., typically as low as 30–40% even with more sensitive mycobacterial culture).9 Second, culture has limitations. Although direct culture inoculation at the bedside gives a high yield,13 it takes 1–6 weeks to deliver a final result. Culture also fails to differentiate between the various mycobacteria and, thus, requires additional polymerase chain reaction (PCR)-based testing for accurate speciation. Third, although chest radiography is considered reasonably accurate and practical for diagnosis of intrathoracic TB in children with suspicious symptoms, it has many limitations. Most importantly, it is not helpful in the diagnosis of extrathoracic TB without concurrent intrathoracic manifestations, a combination common in children.2,10 For example, a recent South African national study reported that 39,739/270,178 (14.7%) of new TB cases in 2005 were extrapulmonary TB.2 A Cape Town study reported that 72/439 (16.4%) of children treated for active TB had extrathoracic disease and 65.4% of the positive cases had no concurrent intrathoracic disease manifestations.11 TB lymphadenitis was the most common form of extrathoracic TB in the Cape Town study and nearly all cases with peripheral TB lymphadenitis presented with enlarged cervical lymph nodes,11 consistent with previous research.12,14 TB lymphadenitis is the most common cause of persistent cervical lymphadenopathy in this TB endemic area.11 The challenge of diagnosing TB in children highlights the urgent need for rapid and definitive mycobacterial diagnosis in children with persistent cervical lymphadenopathy, which is the focus of our research.

Fine-needle aspiration biopsy (FNAB) has many qualities that make it an excellent diagnostic procedure in children with persistent superficial lymphadenopathy, especially in TB-endemic countries with limited resources. FNAB is a simple technique to perform. It is minimally invasive, safe and has very limited side effects when performed with a small gauge needle (e.g., no sinus formation). It can be performed as an outpatient procedure by trained medical or paramedical staff.13,15 Collecting a representative sample of material from a lymph node allows cytomorphological analysis and identification of the organism by direct microscopy and/or culture.

The diagnostic value of various microscopy techniques used to evaluate an FNAB sample has not been fully evaluated in children. A pilot study of 65 aspirates in children using cytomorphology, ZN staining, and autofluorescence as diagnostic modalities had showed 92.3% concordance with culture when all three modalities were combined.16 The current study aimed to compare the diagnostic performance of cytomorphology, ZN staining, autofluorescence, and culture in a large cohort of children.

Materials and Methods
A prospective study was performed in Tygerberg Hospital during the period of January 2003 to June 2005. All children (a) less than 13 years of age (eligible for access to pediatric services), (b) with persistent superficial lymphadenopathy not responsive to first line antibiotic therapy, and (c) with a clinical suspicion of mycobacterial infection routinely referred for diagnostic FNAB were included in the study.

The referring clinician was requested to give oral sedation and analgesia 30 minutes prior to the procedure, which was performed in the ward or outpatient clinic with the assistance of the referring doctor and/or nursing staff. Consent was obtained from the legal guardian accompanying the child. The FNAB was performed using a 23-g or 25-g needle attached to a 10-cc syringe. The skin was cleaned using an alcohol swab, the child firmly immobilized and the node stabilized by the aspirator while performing the aspirate using no more than 2-cc suction. Standard precautions were taken to minimize any complications. Two smears were prepared from each aspirate and the needle and syringe rinsed in mycobacterial growth tube indicator (MGIT, Beckton Dickinson, USA) medium for mycobacterial culture. One smear was spray fixed with commercial cytology fixative for Papanicolaou staining and the other air-dried for Giemsa staining. Smears were sent to cytology for cytomorphology, autofluorescence, and ZN staining.

The stained smears were screened and evaluated for adequacy and diagnosis using each of the modalities described. Cytological results were reported as nonspecific reactive lymphadenopathy or cytologically consistent with mycobacterial infection, based on specific morphological criteria which have been previously described.17–19 Immune competent patients with TB present with the more classical morphological picture of epithelioid granulomata and epithelioid histiocytes in a background of reactive lymphocytes and plasma cells (Fig. C-1). A small amount of amorphous necrosis and occasional giant cells may be present. Patients with tuberculosis who are immune compromised have smears showing abundant “dirty” necrosis in which neutrophils and cellular debris are prominent (Fig. C-2). There is a spectrum of morphology between these extremes, and aspirates from patients with mycobacterial lymphadenitis may occasionally show morphological features of suppurative acute lymphadenitis.
In patients with lymphadenitis due to *M. bovis* BCG, histiocytes with abundant foamy cytoplasm are present. Because of the considerable morphological variation, the fact that patients with mycobacterial infection may present with aspirates showing nonspecific lymphadenitis and that other organisms such as fungal infections may present a similar clinical and morphological picture, staining for mycobacteria was performed on all aspirates, irrespective of the cytological diagnosis. ZN stains were performed on one of the Giemsa stained slides according to a slightly modified technique. The smears were differentiated in 3% acid alcohol for 1 minute and counterstained with 1% methylene blue for 10 seconds only (Fig. C-3).

One of the Papanicolaou stained slides was screened using a Zeiss Axiophot microscope with a fluorescent attachment and a wide-band blue excitation filter (450–480 nm). Mycobacteria auto fluoresce as brilliant yellow bacilli, thin and slightly curved with polar enhancement and a uniform length of 2.0–2.7 μm (Fig. C-4). Despite identification of the mycobacteria on ZN staining and/or autofluorescence, *Mycobacterium tuberculosis*, *Mycobacterium bovis* BCG, and *Mycobacterium avium–intracellulare* are morphologically indistinguishable.

Culture samples were inoculated into 7H9 liquid growth medium at the bedside and sent directly to microbiology, according to standard protocol. Cultures were incubated at 37°C for 8 weeks or until indicated as positive, according to standard protocol using the MGIT system (Becton Dickinson, Cockeysville, MD). All mycobacterial isolates were identified as *Mycobacterium tuberculosis*, *Mycobacterium bovis* BCG or nontuberculous mycobacteria (NTM) by polymerase chain reaction (PCR) testing.
Parents gave written informed consent for study participation and ethics approval was obtained from the Institutional Review Board of Stellenbosch University.

Results

FNABs were collected from 200 children, and 25 (12.5%) aspirates were inadequate. Samples from 175 children were included in the analysis. Patient demographics and sample data are reported in Table I.

The results for the three diagnostic techniques were as follows. Cultures were positive in 79 cases (45%): 61 *Mycobacterium tuberculosis*, 18 *Mycobacterium bovis* BCG and no NTM positive samples. The median age of children with positive cultures was similar to those with negative cultures (2 years). However, all children with *Mycobacterium bovis* BCG were less than 2 years of age, the majority 16/18 (89%) being infants.

Table II compares the diagnostic performance of the three microscopy techniques cytomorphology, autofluorescence, and ZN staining alone and in combination to culture as the gold standard. However, culture is probably not the optimal gold standard, as a false negative culture is not uncommon. The previous pilot study investigated the culture negative cases in which any of the other modalities were positive and identified eight false culture negative cases that were clinically assessed as having TB and responded to TB therapy. Patients may commence TB treatment prior to referral for FNAB. Therefore, using culture alone as the gold standard may be flawed, and we identified an alternative gold standard as culture positive or positive cytomorphology plus positive autofluorescence or ZN staining.

Using this alternative gold standard improved the diagnostic performance of all three modalities. The concordance rate with the new standard is 83% for cytomorphology, 79% for autofluorescence, 76% for ZN staining, and 84% for culture (Table III). The sensitivity, specificity, and overall efficacy of these diagnostic modalities are shown in Table IV.

Discussion

In many low- and middle-income countries with limited resources, the diagnosis of TB is still based on poorly validated symptom-based algorithms, often not resulting in a definitive diagnosis. FNAB provides a rapid and definitive tissue diagnosis in the majority of children with superficial lymphadenopathy. This study demonstrates that it also permits confirmation of the presence of mycobacteria, with fluorescence microscopy being more sensitive than ZN staining. Recent studies have demonstrated that light emitting diode (LED) technology provides a cheap and reliable light source with minimal energy requirements that performs as well as the traditional mercury vapour lamp used in this study. To date, the high cost and short half-life of the mercury vapour lamp has limited the availability of fluorescence microscopy in healthcare systems with constrained financial resources.

The 2007 WHO report noted that over half of the populations in the African, South-East Asia, and Western Pacific regions had limited coverage of culture services. Most countries had neither national policies to expand culture and sensitivity testing services nor the technical capacity to implement and support such services. In South Africa, there were 143 laboratories performing smear microscopy and 18 laboratories performing culture. This is mainly due to centralization of the more expensive and

| Table I. Demographics and Sample Characteristics |
|---------------------------------|-----------------|-----------------|-----------------|
| Number of aspirations           | 200             | Percentage      |
| Inadequate samples             | 25              | 12.5            |
|Specimens included in analysis  | 175             | 87.5            |
|Age                             |                 |                 |
|≤ 1 year                        | 75              | 42.9            |
|2–4 years                       | 48              | 27.4            |
|5–9 years                       | 32              | 18.3            |
|≥ 10 years                      | 20              | 11.4            |
|Gender                          |                 |                 |
|Male                            | 82              | 46.9            |
|Female                          | 93              | 53.1            |
|Diagnosis                       |                 |                 |
|Malignancy                       | 0               | –               |
|Nonspecific node                | 86              | 49.1            |
|Consistent with mycobacterial infection | 89           | 50.1            |
|Fluorescence positive           | 73              | 41.7            |
|ZN positive                     | 68              | 38.9            |
|Culture positive mycobacterial infection | 79           | 45.0            |
|Organism isolated               |                 |                 |
|M. Tuberculosis                 | 61              | 77.0            |
|M. bovis BCG                    | 18              | 23.0            |
|Nontuberculous mycobacteria     | 0               | –               |

| Table II. Concordance of Diagnostic Modalities with Culture as Gold Standard |
|---------------------------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|
|                                | Fluorescence    | ZN              | Any diagnostic modality | Any two diagnostic modalities |
|                                | 27%            | 29%            | 39%             | 30%             |
|Positive agreement              | 27%            | 45%            | 34%             | 39%             |
|Negative agreement              | 14%            | 73%            | 73%             | 69%             |
|Concordant                      | 18%            | 10%            | 21%             | 15%             |
|False positive                  | 18%            | 17%            | 6%              | 15%             |
|False negative                  | 12%            | 17%            | 6%              | 15%             |
|Pearson χ²                      | 36.19          | 43.21          | 24.86           |
|Phi                             | 0.50           | 0.38           |                 |

n = 175; all Pearson χ² statistics are significant at P ≤ 0.0001.
sophisticated culture techniques. Centralization is also a feasible option in resource constrained healthcare systems because FNAB can be done as an outpatient procedure by trained medical or paramedical personnel and correctly fixed slides can be sent to specialized diagnostic facilities for cytological evaluation and fluorescent microscopy.

The diagnosis of TB lymphadenitis is increasingly complicated by the HIV pandemic. HIV infection, in children and adults, is a major contributor to the escalating TB epidemic worldwide. At the end of 2000, it was estimated that there were 11.5 million HIV-infected people coinfected with TB in sub-Saharan Africa. In 2005, South Africa with 0.7% of the world’s population had 19% of all cases of TB in adult HIV-positive people. Infection with the HIV virus is estimated to increase the lifetime risk of a patient infected with \( M. tuberculosis \) of developing TB by 10-fold to 50%. Children are not exempt from this burden.

A hospital-based study from Zambia demonstrated a HIV seroprevalence rate in children with TB to be >70%, while the minority of childhood TB cases were HIV negative. In HIV-infected children the diagnosis of intrathoracic TB is even more difficult due to concomitant chronic diseases such as lymphocytic interstitial pneumonitis (LIP) and other opportunistic infections. FNAB is widely utilized in the diagnosis of palpable masses including peripheral lymphadenopathy, and its value in the diagnosis of mycobacterial lymphadenitis in adults is well documented. It offers a simple, effective, and safe modality for obtaining a representative sample of material from a lymph node, permitting cytological evaluation, identification of the organism by morphology, culture and molecular techniques such as the polymerase chain reaction (PCR). Cytomorphology is simple, but not specific, as other opportunistic infections may present with similar cytological changes and the quality of the smear is operator dependent. With good clinical triage of patients, its sensitivity and specificity can be increased. In our study, cytomorphology achieved a sensitivity of 78% and a specificity of 91%.

ZN staining, even in good laboratories, is generally regarded as insensitive, identifying organisms in only about 20% of culture positive samples. This may be dependent on experience, the staining, and the bacterial load. In our study, ZN staining achieved sensitivity of 62% but with a specificity of 97%. Although autofluorescence is simple, sensitive, and inexpensive, it is not widely used. It requires a fluorescent microscope, which may not be readily available, but has the advantage of not requiring additional stains and is therefore inexpensive and rapid. In our study, it performed similar to ZN stain-

### Table III. Concordance of Diagnostic Modalities with New Gold Standard Culture Positive or Cytomorphology Positive Plus Positive Autofluorescence or ZN

<table>
<thead>
<tr>
<th></th>
<th>Cytology (%)</th>
<th>Fluorescence (%)</th>
<th>ZN (%)</th>
<th>Culture (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Positive agreement</td>
<td>47</td>
<td>41</td>
<td>38</td>
<td>45</td>
</tr>
<tr>
<td>Negative agreement</td>
<td>36</td>
<td>38</td>
<td>38</td>
<td>39</td>
</tr>
<tr>
<td>Concordant</td>
<td>83</td>
<td>79</td>
<td>76</td>
<td>84</td>
</tr>
<tr>
<td>False positive</td>
<td>3</td>
<td>1</td>
<td>1</td>
<td>0</td>
</tr>
<tr>
<td>False negative</td>
<td>13</td>
<td>20</td>
<td>23</td>
<td>15</td>
</tr>
<tr>
<td>Pearson ( \chi^2 )</td>
<td>81.02</td>
<td>70.59</td>
<td>61.99</td>
<td>93.74</td>
</tr>
<tr>
<td>Phi</td>
<td>0.68</td>
<td>0.64</td>
<td>0.60</td>
<td>0.73</td>
</tr>
</tbody>
</table>

\( n = 175; \) all Pearson \( \chi^2 \) statistics are significant at \( P \leq 0.0001 \).

### Table IV. Summary of All Diagnostic Modalities Using an Alternative Gold Standard: Culture Positive or Cytomorphology Positive Plus Positive Autofluorescence or ZN

<table>
<thead>
<tr>
<th></th>
<th>Cytology (%)</th>
<th>Fluorescence (%)</th>
<th>ZN (%)</th>
<th>Culture (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sensitivity</td>
<td>78</td>
<td>67</td>
<td>62</td>
<td>75</td>
</tr>
<tr>
<td>Specificity</td>
<td>91</td>
<td>97</td>
<td>97</td>
<td>100</td>
</tr>
<tr>
<td>PPV</td>
<td>93</td>
<td>97</td>
<td>97</td>
<td>100</td>
</tr>
<tr>
<td>NPV</td>
<td>73</td>
<td>66</td>
<td>63</td>
<td>72</td>
</tr>
<tr>
<td>Efficacy</td>
<td>83</td>
<td>79</td>
<td>76</td>
<td>84</td>
</tr>
</tbody>
</table>

Disseminated BCG disease is a rare complication of BCG vaccination, classically only described in children with severe congenital immune deficiencies. However, HIV-infected children also appear to be at high risk. BCG vaccination may also be associated with local complications such as abscess formation at the vaccination site and/or regional lymphadenitis, with reported rates of lymphadenitis in BCG recipients ranging from 0.5% to 17.6%. In South Africa, the recent change in vaccine policy in July 2000 from percutaneous Tokyo strain BCG to intradermal Danish strain BCG vaccine has seen a number of adverse events reported in both HIV-infected and immune competent children. In children with axillary or other regional lymph node enlargement, FNAB offers a simple first line diagnostic procedure to confirm a diagnosis of BCG disease.
ing in identifying mycobacterial infection with a sensitivity of 67% and a specificity of 97%. Its specificity is dependant on the experience of the operator and, as ZN staining, it cannot differentiate between the various mycobacteria. These results may be partly due to the high incidence of mycobacterial disease in South Africa and good clinical selection of patients.

Culture, where feasible and affordable, is an important part of the investigation of suspected mycobacterial lymphadenitis. Direct inoculation into MGIT or Bactec medium at the bedside gives a high yield, but it takes 1–6 weeks for a result, and fails to differentiate the various mycobacteria unless additional PCR-based testing is available for accurate speciation of \textit{M. tuberculosis} complex. Culture has inherent problems and may be influenced by many factors such as inefficient sampling, contamination, variable bacterial load, and commencement of therapy prior to the diagnostic procedure. If the cytology is consistent with mycobacterial infection, and the organism is identified by ZN staining or autofluorescence, the probability of a false positive diagnosis is small, and patients may safely commence therapy. FNAB of superficial lymph nodes is an outpatient procedure and requires little infrastructure and equipment and is therefore ideal for resource-limited countries, which are the very same countries that bear the burden of HIV and TB. The basic diagnostic modalities of cytology and subsequent morphological identification of the organism are readily available and relatively inexpensive. Although culture adds to the cost of the diagnosis, its value lies in providing an opportunity for speciation and sensitivity testing, as well as surveillance of prevalent strains within a community. Although radiometric methods such a MGIT have reduced the time to culture mycobacterial organisms, this may still be unacceptably long for patients who have difficulty in returning for results. Other innovative methods for diagnosing TB are being developed but remain mainly experimental and are still relatively expensive.

Most children referred with suspected mycobacterial lymphadenitis have a tuberculin skin test performed and must return to the clinic for this to be read with in 48–72 hours. If FNAB is performed at the time of the first visit, the results of the aspirate can be available at the return visit 48 hours later. This enables children to be put on appropriate therapy with a high degree of certainty, pending the results of culture and speciation. In countries where culture is not available, FNAB provides the clinician with the ability to make a definitive diagnosis based on cytomorphology and identification of the organism, particularly in immune compromised children, who require urgent and appropriate treatment. If cytology services are not available, children with suspected mycobacterial lymphadenopathy may have an FNAB performed and smears made for direct TB microscopy. Although this is not ideal, it provides reasonable diagnostic accuracy, with the additional option to courier-fixed slides to a center with adequate cytology services. This will help health care workers in TB-endemic countries with limited resources to navigate the diagnostic minefield of persistent lymphadenopathy in children.

In conclusion, FNAB proved highly feasible and provided a rapid and definitive diagnosis in the majority of cases, irrespective of the diagnostic modality used.

References


CHAPTER 6

OPTIMISING CULTURE-BASED AND MOLECULAR DIAGNOSIS

Mycobacterial Transport medium for routine culture of Fine Needle Aspiration Biopsies
CA Wright, C Bamford, Y Prince, A Vermaak, KGP Hoek, BJ Marais, RM Warren,
Archives of Disease in Childhood: in press

Combining Fine Needle Aspiration Biopsy (FNAB) and High Resolution Melt Analysis
to reduce diagnostic delay in Mycobacterial Lymphadenitis
CA Wright, KGP Hoek, BJ Marais, P van Helden, RM Warren,
Diagnostic Cytopathology: in press
Mycobacterial Transport medium for routine culture of Fine Needle Aspiration Biopsies

CA Wright,∗ C Bamford,† Y Prince,† A Vermaak,∗ KGP Hoek,‡ BJ Marais,§ RM Warren,‡
∗Divisions of Anatomical Pathology and †Medical Microbiology, Department of Pathology,
Stellenbosch University and NHLS Tygerberg Hospital, Tygerberg, South Africa
‡NRF Centre of Excellence in Biomedical Tuberculosis Research / MRC Centre for
Molecular and Cellular Biology, Division of Molecular Biology and Human Genetics,
Stellenbosch University, South Africa
§Department of Pediatrics and Child Health and Desmond Tutu TB Centre, Stellenbosch
University, South Africa

SUMMARY

Fine needle aspiration biopsy (FNAB) offers a simple outpatient technique for specimen
collection in paediatric tuberculosis (TB) suspects with peripheral lymphadenopathy. Culture
facilities are usually centralized; to perform FNAB with mycobacterial culture on an
outpatient basis requires use of a sterile transport medium to facilitate bedside inoculation,
maintain organism viability and reduce contamination risk en route to the laboratory.
We compared the mycobacterial yield and time to positive culture following bedside
inoculation into standard MGIT tubes versus initial inoculation into an inexpensive “in house”
liquid growth medium used for transport to the laboratory followed by immediate and delayed
MGIT inoculation (laboratory inoculation).
Over the period of one year 150 FNAB’s were performed; 57 (38%) cultured M. tuberculosis
complex. There was one case each of NTM and M bovis BCG, with the remaining 55 being M
tuberculosis. Results were concordant in 142 (94.7%) bedside and laboratory inoculation
pairs. There was no significant difference in time to positive culture between the bedside and
laboratory inoculation tubes (16.2 days S.D. 0.87 versus 17.1 days S.D. 0.85). There were 31
pairs with matched immediate and delayed laboratory inoculation culture results, of which 29 were concordant (93.6 %). The 2 discordant pairs showed no growth in the delayed culture; both had required repeated decontamination due to bacterial contamination introduced in the laboratory.

The use of inexpensive “in house” liquid growth medium transport bottles, combined with practical tuition in FNAB, will improve cost effective diagnosis of TB at primary health care level.

INTRODUCTION

Tuberculosis (TB) may be extrapulmonary in 10-30% children,¹ and TB lymphadenitis is the most common extra-pulmonary manifestation of TB in endemic areas;² 5-10% of children may have TB adenitis in association with pulmonary involvement.³ This provides an excellent opportunity to obtain bacteriologic confirmation using fine needle aspiration biopsy (FNAB).⁴ FNAB is a simple and safe outpatient procedure that can be performed by nurses in resource-limited settings⁵ and provides material for direct microscopy as well as culture and susceptibility testing. Cultures from FNAB specimens in children give a greater yield in a shorter time, compared to conventional respiratory specimens such as gastric washings.⁶

Direct bedside inoculation of FNAB specimens is ideal and excellent results have been obtained with commercial liquid media systems such as Mycobacterial Growth Indicator Tubes (MGIT, Beckton Dickinson, USA).⁴ This practice however, is limited by cost and availability.⁴ We aimed to develop a simple and cheap mycobacterial transport medium and then to compare the bacteriologic yield and time to positive culture achieved with direct bedside inoculation into MGIT versus initial use of the transport medium.
MATERIALS AND METHODS

All adults and children referred to the FNAB Clinic at Tygerberg Hospital, Cape Town, South Africa (June 2007-May 2008) with clinical mycobacterial lymphadenitis and in whom written informed consent to participate in the study was obtained (following ethics approval) were included.

TB transport bottles, containing 1 ml aliquots of Middlebrook 7H9 broth (with 0.2% glycerol and 0.05% Tween 80 added) in 10ml headspace glass vials, were prepared “in-house” (Biomedical TB Research Unit by a study scientist, KGPH) in a laminar flow cabinet, sealed with 20mmTFE/Sil Septa and 20mm Aluminium open top seals and autoclaved at 120°C / 20min (Figure 1). Unlike the MGIT tube, the transport bottle does not require removal of the lid, minimizing the risk for contamination.

FNAB was done following standard protocol. Two needle passes were performed: from the first, cytology slides were prepared before the syringe and needle were rinsed by withdrawing an aliquot of liquid growth media into the syringe and then expelling the contents back into the MGIT tube (bedside inoculation or control). After transport to the laboratory within the same hospital, within 2-24 hours, the MGIT tube was incubated in a BACTEC MGIT 960 machine for 42 days.

From the second FNAB pass, slides were prepared for cytology and the syringe and needle were rinsed into the TB transport bottle. In the laboratory, 0.5ml was aspirated from the transport bottle, inoculated into a separate MGIT tube (immediate laboratory inoculation) and incubated as above. In a subset of consecutive samples, the transport bottle was held at room temperature (12-30 deg Celsius) for 7 days after which subculture was performed as described.
above (delayed laboratory inoculation). The delayed inoculation group was designed to mimic the situation of rural clinics where specimens may take some days to reach the laboratory.

PANTA antibiotic mixture (containing polymixin B, amphotericin B, nalidixic acid, trimethoprim and azlocillin) (Becton Dickinson, New York, NY, USA) was added to all MGIT tubes before incubation. All positive MGIT tubes were confirmed to contain acid fast bacilli in the absence of bacterial contamination, followed by accurate polymerase chain reaction (PCR) speciation. The mycobacterial yield and time to positive culture in the respective MGIT tubes were compared using Statistica Version 8 software and the Chi² test.

RESULTS

Over the period of one year 150 FNAB’s were performed, and 24.5% of these were in patients less than 16 years, the youngest being 5 months old. *M. tuberculosis* complex was cultured in 57 patients (38%). There was one case each of NTM and *M. bovis BCG*, with the remaining 55 being *M. tuberculosis*; consistent with the profile of disease in our population. In the TB culture negative patients there were an additional 12 cases (11.4%) positive on cytology (morphological evidence of mycobacterial infection and identification of the organism). In 17 patients there was a specific diagnosis other than mycobacterial infection, made on cytology, 13 of these malignant neoplasms.

Results of culture were concordant in 142 (94.7%) bedside and laboratory inoculation pairs, with 8 discordant pairs: 3 were positive on bedside inoculation only and 5 were positive on laboratory inoculation only. In one of the discordant pairs, positive on bedside inoculation and negative on laboratory inoculation there was inadequate material on the cytology slides. There was no significant difference in time to positive culture between the bedside and laboratory inoculation tubes (16.2 days S.D. 0.87 versus 17.1 days S.D. 0.85).
There were 31 pairs with matched immediate and delayed laboratory inoculation culture results, of which 29 were concordant (93.6 %). The 2 discordant pairs showed no growth in the delayed culture, compared to growth in bedside and immediate laboratory cultures; both had required repeated decontamination due to bacterial contamination introduced in the laboratory. There was no significant difference in time to positive culture between the immediate and delayed laboratory inoculation groups (Table 1).

DISCUSSION

Although FNAB has been shown to be an excellent method for obtaining specimens for culture as well as cytology, the method for transporting specimens to the laboratory is rarely specified.9 Mycobacteria need to remain viable during transportation if culture is to be successful and traditional methods for the preservation of specimens, e.g. refrigeration10 are not applicable to FNAB specimens in resource-poor settings. In most patients FNAB yields insufficient material to express into a sterile tube, and commercial liquid culture media, e.g. BACTEC™ MYCO/F LYTIC are recommended only for blood and body fluids.11 Submission of the syringe (with or without the needle attached) directly to the laboratory compromises the quality of the specimen and puts health care workers at risk. This practice is unacceptable, especially in settings with a high prevalence of MDR TB and/or HIV.

Rinsing the needle and syringe in commercial liquid culture medium, such as MGIT, is very successful in obtaining a positive culture.4,5 However, the widespread distribution of MGIT tubes is limited by issues of cost and storage requirements. These tubes need to be stored in the dark and removal of the cap for inoculation outside the laboratory increases the risk for contamination. This study suggests that subculture from a TB transport bottle, even after a 7 day delay, gives similar yields in a similar time period, to direct bedside inoculation. The
few discrepancies (8/150) noted between the bedside and laboratory cultures may be due to random sampling differences as the tubes were inoculated from separate needle passes.

The transport medium described in this study is inexpensive (MGIT tube $4 vs. $1 TB transport bottle, 2008 prices in South Africa), easy to prepare “in house”, does not require refrigeration as it is stable through a range of temperatures, and does not require removal of the lid for inoculation. Transport of these specimens from rural clinics may follow the same route as other laboratory specimens such as sputum and peripheral blood. The positive culture yield and the time to positive culture from the transport medium, even after 7 days at room temperature, are statistically comparable to direct bedside inoculation into MGIT tubes. The production and distribution of these bottles to clinics and hospitals, combined with practical tuition in the optimal performance of FNAB, will have a positive impact on the cost effective diagnosis of TB at primary health care level.

ACKNOWLEDGEMENTS

Professor Martin Kidd (Stellenbosch University) for statistical analysis. This study is in partial fulfillment of a PhD thesis.

CA Wright was funded by grant from the National Health Laboratory Services, South Africa.

Competing interests: none declared

Copyright licence statement

The Corresponding Author has the right to grant on behalf of all authors and does grant on behalf of all authors, an exclusive licence (or non-exclusive for government employees) on a worldwide basis to the BMJ Publishing Group Ltd and its Licensees to permit this article (if accepted) to be published in Archives of Disease in Childhood and any other BMJPGL
products to exploit all subsidiary rights, as set out in our licence (http://adc.bmj.com/iforalicence.pdf).

**What is already known about this topic.**

FNAB is used as to diagnose suspected mycobacterial lymphadenopathy.

Mycobacteria may be seen on staining FNAB material, but this does not allow speciation (TB or non-tuberculous mycobacteria) or antimicrobial sensitivity testing.

**What this study adds**

A simple and inexpensive means to obtain mycobacterial culture from FNAB specimens was developed using an “in house” liquid growth medium for transport to the laboratory.

Transporting FNAB material to the laboratory in the “in house” medium was similar to a commercial medium with respect to culturing mycobacteria and in time to positive culture.

Similar results were obtained when the ‘in house” transport bottle was kept at room temperature for 7 days before culturing for mycobacteria.

**REFERENCES**


Table 1

The mycobacterial yield and time to positive culture achieved with fine needle aspiration biopsy and variable timing of MGIT* inoculation

<table>
<thead>
<tr>
<th>Timing of MGIT* inoculation</th>
<th>Total no specimens</th>
<th>Mycobacterial culture</th>
<th>Time to positive culture</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Positive</td>
<td>Negative</td>
</tr>
<tr>
<td>Bedside vs. immediate laboratory inoculation from transport bottle</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Bedside</td>
<td>150</td>
<td>53 (35.3%)</td>
<td>97 (64.7%)</td>
</tr>
<tr>
<td>Immediate laboratory</td>
<td>150</td>
<td>55 (36.7%)</td>
<td>95 (63.3%)</td>
</tr>
<tr>
<td>Subset - matched pairs: delayed vs. immediate laboratory inoculation from transport bottle</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Delayed laboratory (day 7)</td>
<td>31</td>
<td>11 (35.5%)</td>
<td>20 (64.5%)</td>
</tr>
<tr>
<td>Immediate laboratory (day 0)</td>
<td>31</td>
<td>13 (41.9%)</td>
<td>18 (58.1%)</td>
</tr>
</tbody>
</table>

*MGIT- Mycobacterial Growth Indicator Tube
# CI- Confidence Interval
Figure 1. TB Transport Bottle
Combining Fine Needle Aspiration Biopsy (FNAB) and High Resolution Melt Analysis to reduce diagnostic delay in Mycobacterial Lymphadenitis

CA Wright,1* KGP Hoek,2 BJ Marais,3 P van Helden,2 RM Warren,2
Division of Anatomical Pathology ,Department of Pathology, Stellenbosch University and NHLS Tygerberg, South Africa; NRF Centre of Excellence in Biomedical Tuberculosis Research / MRC Centre for Molecular and Cellular Biology, Division of Molecular Biology and Human Genetics, Stellenbosch University, South Africa; 2 and Department of Pediatrics and Child Health, Stellenbosch University, South Africa.

ABSTRACT

Tuberculous lymphadenitis is the most common cause of extra-pulmonary tuberculosis (TB) in developing countries. Lymphadenitis caused by non-tuberculous mycobacteria (NTM) requires consideration, particularly in immunocompromised patients and children in developed countries. Fine Needle Aspiration Biopsy (FNAB) offers a valuable specimen collection technique, but culture confirmation, mycobacterial speciation and drug resistance testing (if indicated) is often unavailable in TB endemic areas and result in unacceptable diagnostic delay.

We evaluated the diagnostic value of high-resolution DNA melting (HRM) analysis in the diagnosis of mycobacterial lymphadenopathy using FNAB and an inexpensive transport medium.

Specimens were collected from patients referred to the FNAB Clinic at Tygerberg Hospital (June 2007-May 2008) with clinical mycobacterial lymphadenitis. Cytology, culture and HRM were performed on all specimens. The reference standard for disease was defined as positive cytology (morphological evidence plus mycobacterial visualization) and/or a positive culture.
Specimens were collected from 104 patients and mycobacterial disease was confirmed in 54 (51.9%); 46 *M. tuberculosis*, 1 *M. Bovis BCG*, 1 NTM, and 8 not speciated. Cytology was positive in 83.3% (45/54) and culture in 72.2% (39/54) of patients. HRM identified 57.4% (31/54) of cases. Using the defined reference standard we recorded 51.9% sensitivity and 94.0% specificity (positive predictive value 90.3%) with HRM analysis.

HRM analysis allowed rapid and species specific diagnosis of mycobacterial lymphadenitis in the majority of patients, permitting early institution of appropriate therapy. Optimization of this technique requires further study.

**INTRODUCTION**

Resource limited countries carry the brunt of the global tuberculosis (TB) epidemic, particularly those affected by the parallel human immunodeficiency virus (HIV) pandemic.1 According to the most recent World Health Organisation (WHO) estimates, in 2007 there were 9.3 million incident (newly diagnosed) TB cases of whom 1.4 million (14.8%) were HIV-infected. The African region accounted for 79% of HIV-infected TB cases.1 Although TB incidence rates seem to plateau and/or decline in most regions, absolute numbers continue to rise due to increases in population size. The projected scale of the epidemic and ongoing transmission of drug resistant TB remains alarming.1 WHO estimated that between 2000 and 2010 1 billion people will be newly infected with *M. tuberculosis*, resulting in 200 million TB cases and 35 million deaths.2

Peripheral lymphadenitis is the most common extra-pulmonary manifestation of TB.3-4 TB lymphadenitis is also the most common cause of persistent cervical lymphadenopathy in children from TB endemic areas.5-7 In developed countries where the incidence of TB is low, non-tuberculous mycobacteria (NTM) are frequently responsible for mycobacterial lymphadenitis, particularly in children and HIV-infected immune compromised adults.3,8-9 In patients with a peripheral lymph node mass fine needle aspiration biopsy (FNAB) is a
valuable and underutilized specimen collection technique. This simple and safe procedure allows rapid confirmation of mycobacterial disease using cytomorphology and direct mycobacterial visualization with either Ziehl-Neelsen (ZN) staining or fluorescence microscopy.

Mycobacterial culture is required for accurate species determination (speciation) and drug susceptibility testing, since organisms in the *M. tuberculosis* complex are morphologically indistinguishable. The amount of material harvested during FNAB is minimal, and the needle needs to be rinsed at the bedside in liquid medium to facilitate culture. Although FNAB can be safely performed as an outpatient procedure by well trained nurses, the need for direct inoculation and unavailability of liquid culture tubes limited decentralization. Use of an inexpensive transport medium for direct inoculation has been described, which should facilitate mycobacterial culture from FNAB’s performed in rural clinics and hospitals. Direct bedside inoculation at the time of FNAB collection provides an excellent diagnostic yield but culture results may take up to 6 weeks and requires additional speciation.

Performing nucleic acid amplification tests (NAATs) on FNAB specimens may provide a rapid species specific diagnosis and expedite access to appropriate therapy. A recent systematic review demonstrated highly variable results with NAATs to diagnose TB lymphadenitis, reported sensitivities ranged between 2-100% (specificities 28-100%). Most NAATs analyze the polymerase chain reaction (PCR) products by gel electrophoresis or other open tube formats, which afford the opportunity for laboratory cross contamination. These technically challenging techniques will pose problems in countries with limited laboratory resources. High resolution DNA melt analysis (HRM) is a simple “closed-tube” technique that reduces the risk of cross contamination. Specific PCR amplicons are identified according to their characteristic DNA melting profile. The amplicons are combined with a saturating dye that fluoresces in the presence of double stranded DNA. It is heated through a range of
temperatures while fluorescence is monitored. As the double stranded DNA dissociates (melts) into single strands the fluorescence decreases. The melting peak of the specific infectious agent is identified. The amplicon specific for \textit{M.tuberculosis} melt at 90.5°C and other members of the \textit{M.tuberculosis} complex at 86°C. This simple technique can be used in routine diagnostic laboratories.\textsuperscript{18-19}

The aim of our study was to evaluate the value of PCR-based HRM analysis,\textsuperscript{18} to provide a rapid and accurate diagnosis of mycobacterial disease using routinely collected FNAB specimens directly inoculated into liquid transport medium.

**MATERIALS AND METHODS**

All adults and children referred to the FNAB Clinic at Tygerberg Hospital (June 2007-May 2008) with a superficial lymph node mass suggestive of possible mycobacterial lymphadenitis and in whom written informed consent to participate in the study was obtained, were included.

**Specimen Collection**

FNAB was done following standard protocol as an outpatient procedure by a pathologist.\textsuperscript{10} The lymph node was stabilized by the aspirator and two needle passes were performed using a 23g or 25g needle attached to a 10 ml syringe while applying a constant suction of no more than 2 ml. From each aspirate two smears were prepared: one fixed with commercial cytology fixative for Papanicolaou staining and the other air dried for the Giemsa and subsequent ZN staining. The residual material in the syringe and needle was collected by withdrawing an aliquot of liquid growth media from the TB transport bottle into the syringe and then expelling the contents back into the bottle. No additional needle passes were performed to collect material for microbiology or PCR. The TB transport bottles were prepared “in-house” in a laminar flow cabinet: 10ml headspace glass vials containing 1 ml of Middlebrook 7H9
broth (with 0.2% glycerol and 0.05% Tween 80 added), sealed with 20mm TFE/Sil Septa and 20mm Aluminium open top seals and autoclaved at 120º C for 20min.20

Cytology

Stained smears were evaluated for adequacy and to make a morphological diagnosis. If any specific lesion such as a lymphoepithelial cyst or neoplasia was present, this was reported. Alternatively we reported “non-specific reactive lymphadenopathy” or “cytologically consistent with mycobacterial infection”, based on specific morphological criteria.21-23 In immunocompetent patients with TB, epithelioid granulomata and epithelioid histiocytes may be identified in a background of reactive lymphocytes and plasma cells (Fig 1). A small amount of amorphous necrosis may also be identified. In immunocompromised patients, the cytological picture is that of abundant necrosis in which neutrophils and cellular debris are prominent (Fig 2). In patients with lymphadenitis due to M. bovis BCG or NTM infection, histiocytes with abundant foamy cytoplasm may be present.8, 24-25

Mycobacteria were visualized using a ZN stain performed on a Giemsa stained slide according to a slightly modified technique (Fig 3). Smears were differentiated in 3% acid alcohol for one minute and counterstained with 1% methylene blue for 10 seconds only. In addition, one of the Papanicolaou stained slides was screened using a Zeiss Axiophot microscope with a fluorescent attachment and a wide-band blue excitation filter (450–480 nm); mycobacteria fluoresce as brilliant yellow bacilli, thin and slightly curved with polar enhancement and a uniform length of 2.0 –2.7 microns (Fig 4).11

Microbiology

In the laboratory, 0.5ml of the media was aspirated from the TB transport bottle, inoculated into a separate MGIT tube containing the PANTA antibiotic mixture and incubated in a BACTEC MGIT 960 machine for 42 days. All positive MGIT tubes were confirmed to contain acid fast bacilli in the absence of bacterial contamination by ZN staining. Mycobacteria were identified as M. tuberculosis or other by standard PCR.26
Extraction of mycobacterial DNA

The TB transport bottles containing the remaining 0.5ml media were stored at -20°C until further analysis. Mycobacteria within a 250µl aliquot were pelleted by centrifugation at full speed (14 000rpm) for 10 minutes. The supernatant was discarded and the bacterial pellet was resuspended in 1ml phosphate buffered saline and re-centrifuged at full speed (14 000rpm) to remove residual blood. Thereafter the bacterial pellet was resuspended in 30µl ddH2O and heat inactivated at 100°C for 20 minutes. The lysed bacterial extract was stored on filter paper (FTA® Classic Card Collection, Storage and Purification system, Whatman, UK). Genomic DNA was eluted from the filter paper cards following manufacturers’ instructions (high pH, low pH protocol) and served as the template for subsequent PCR amplification.

Speciation and HRM Analysis

Primers were designed to amplify the Region of Deletion 9 (RD9), present in *M. tuberculosis* and *M. canetti*, but absent from all other members of the *M. tuberculosis* complex. PCR was performed as previously described 19 using an annealing temperature of 62°C and the following primers RD9Fs1 5’-CAA GTT GCC GTT TCG AGC C-3’, RD9FR 5’-GCT ACC CTC GAC CAA GTG TT-3’ and RD9INT 5’-CAA TGT TTG TTG CGC TGC-3’. 27 Resulting amplification products underwent HRM analysis in a Rotorgene™6000 real-time rotary analyzer (Corbett Life Science, Australia). The thermal denaturation profiles were measured as previously described.19 The infectious agent was identified by the Rotorgene™6000 software according to the presence of derivative melt peaks located within defined temperature bins. *M. tuberculosis* PCR products melt at 90°C (*M. tuberculosis* could not be differentiated from *M. canettii*, but *M. canettii* is very rarely observed and has not been recorded in the study setting) while PCR products of other members of the *M. tuberculosis* complex melted at 86°C. (Fig.4)
Reference standard for mycobacterial disease

The reference standard for the presence of mycobacterial disease\textsuperscript{17} was defined as positive cytology (morphology consistent with mycobacterial infection plus direct visualization of the organism) and/or positive culture with speciation.

Statistical Analysis

We assessed the diagnostic accuracy of PCR-based HRM analysis compared to cytology, mycobacterial culture and the reference standard as defined using Pearson’s chi-square ($X^2$) and Fisher’s exact tests.\textsuperscript{28} All analyses were conducted using Statistica Version 8.\textsuperscript{29} Ethics approval was obtained from the Institutional Review Board of Stellenbosch University (N05/03/043).

RESULTS

FNAB specimens were collected from 104 patients with possible mycobacterial lymphadenitis in whom complete cytopathology and mycobacterial culture results were available together with PCR-based HRM analysis. The median age was 30 years with a range from 4 months to 62 years. Children less than 13 years of age compromised 21.2 % of the study population. There was no significant difference in gender distribution. HIV status was known in 46.1\% of patients (48 /104) and 75\% of these were HIV positive (36/48). There were 23 HIV-positive patients and 6 HIV negative patients with mycobacterial disease as defined by the reference standard above. Table 1 summarizes the demographics and diagnostic outcome of the study population.

Applying the defined reference standard 54 of the 104 patients (51.8 \%) were diagnosed with mycobacterial lymphadenitis. Mycobacterial culture was positive in 39/54 (72.2 \%) patients; 37 \textit{M. tuberculosis}, one \textit{M. bovis} BCG and one NTM. The child with \textit{M. bovis} BCG was less than two years old; four others in this age group were positive for \textit{M. tuberculosis}. Cytology was positive in 83.3.9 \% (45/54) of the cases. Cytology and culture
were both positive in 30/54 (55.6%) cases, while HRM analysis was positive in 57.4% (31/54) of cases identified using the reference standard. The calculated sensitivity of HRM analysis was 51.9% and the specificity 94.0% (Fig 2) with a negative predictive value of 64.4% and a positive predictive value of 90.3% (Fig 5). Concordance between the different diagnostic modalities and HRM analysis was poor, with Kappa values of 0.39 vs. cytology, 0.27 vs. mycobacterial culture, 0.19 vs. both cytology and culture, and 0.45 vs. cytology and/or culture (the defined reference standard).

To investigate a possible association between mycobacterial load and the result of PCR-based HRM analysis we tested for any correlation between the time to positivity (TTP, often used as a surrogate of mycobacterial load) and PCR outcome. Among those with positive mycobacterial cultures no correlation between the time to positivity and PCR-based HRM analysis could be demonstrated; mean TTP in PCR positive cases was 18 days compared to 22 days in PCR negative cases (p=0.22).( Fig 5)

**DISCUSSION**

With global coordination of control efforts TB incidence rates seem to be stabilizing and showing signs of decline in all six world regions. However, the TB disease burden in areas plagued by the concurrent HIV pandemic remains at unprecedented levels. Although there is some assistance for the development of strong laboratory networks in developing countries, the existing infrastructure remains poor in most TB endemic areas. South Africa, despite its relatively good infrastructure and health care services, in 2007 reported a national TB incidence of 739.6/100,000 population and an adult TB incidence in the Western Cape Province, a high burden region, of 1005.7/100,000.30

In TB endemic areas with ongoing transmission, children constitute a significant percentage of the total caseload, estimated at 15-20%. Immune immaturity and/or compromise, most often due to HIV infection, influence the risk to develop extra
pulmonary and/or disseminated TB as well as the likelihood of rapid disease progression and TB-related mortality.\textsuperscript{34-35} Autopsy studies in Africa \textsuperscript{33, 36-39} have shown that up to 54\% of deaths in HIV-infected adults and 20\% in HIV-infected children are due to TB. Reducing this mortality and morbidity necessitates early detection, efficient diagnosis and timely institution of appropriate therapy.

FNAB is a simple, non-invasive specimen collection technique in patients with peripheral lymph node masses\textsuperscript{10}. It assists with rapid diagnosis of mycobacterial disease, but also helps to rule out alternative diagnoses that may require urgent treatment such as neoplasia. Diagnostic sensitivity is dependent on the experience of the pathologist and for mycobacterial disease varies from 32-78\%\textsuperscript{.10, 40-41} Identification of the organism is essential particularly in immune compromised patients, as other opportunistic infections such as fungal organisms may elicit a similar morphological reaction pattern. Mycobacteria may be visualized using ZN staining, which is a very simple stain but has suboptimal sensitivity (20-62\%).\textsuperscript{10, 17, 42-43} Autofluorescence, using the ability of mycobacteria on Papanicolaou stains smears to fluoresce using a blue excitation filter requires no additional staining and improves sensitivity (65-67\%).\textsuperscript{10-11} Culture is often regarded as the “gold standard”, but is limited by poor sensitivity (reported to be as low as 2-34\%) \textsuperscript{41, 44} and is highly dependent on the culture medium and inoculation technique used. A positive result is delayed by 1-6 weeks and requires additional PCR-based testing for speciation.

Direct NAAT application provides results in 3 to 6 hours and has been evaluated with respiratory and non-respiratory specimens such as FNAB’s.\textsuperscript{17, 43-47} Test sensitivity with respiratory specimens is highly dependent on mycobacterial load and use current PCR-based tests are only advised in patients with sputum smear-positive TB. Results with extra-pulmonary disease have been variable but recent studies have shown excellent results. Use of a nested PCR in cervical TB lymphadenitis in Mexican children showed a sensitivity of 96\% and a specificity of 93\%\textsuperscript{, 47} whilst a study utilizing DNA from dried and fixed cytology smears
showed a sensitivity of 85% and specificity of 95% using nested PCR, although reference standards are not necessarily based on bacteriological confirmation of disease. The majority of these studies use agarose gel electrophoresis to visualize the products, which is not practical in a busy microbiology laboratory. The open tube nature of the procedure also allows the release of amplicons, which pose a real risk of cross-contamination. A recent systematic review evaluating the diagnostic utility of NAATs in TB lymphadenitis identified 36 peer reviewed publications. The authors found marked variation in populations, test techniques, reference standards, volume of material utilized and quality indicators. Few studies controlled for cross contamination and inhibitors in clinical samples, thereby increasing the possibility of both false positive and false negative results. The reference standard used most frequently was culture, although the authors regard the sensitivity of culture from FNAB to be about 62%. Using an imperfect reference standard may lead to an underestimation of test performance and this is a major limitation that is rarely acknowledged.

The current study attempted to address some of the shortcomings noted above. The reference standard used is a combination of positive cytology, defined as cytomorphological features consistent with mycobacterial infection combined with identification of the organism using ZN staining or autofluorescence and/or positive culture. The residual material from the fine needle aspirates was collected in TB transport medium in a sealed bottle, minimizing the possibility of contamination. After washing and concentration of the specimen the pellets were placed on filter paper for storage and to remove inhibitors present in the specimen. After extraction of DNA, the products were amplified using primers designed to amplify the Region of Deletion 9 (RD9), present in *M. tuberculosis* and *M. canetti*, but absent from all other members of the complex. The amplified products were then identified using high resolution melt analysis, which is a closed tube format that minimizes the possibility of cross-contamination.
This technique is rapid and simple, and the equipment required for the HRM such as the Rotorgene 6000™ real-time rotary analyzer (Corbett Life Science, Australia) is relatively inexpensive. Up to 72 samples may be processed in a cycle which takes approximately 3 hours.

The results of PCR using this technique did not differ significantly from that of cytology or culture when these were assessed independently against the reference standard. The sensitivity is relatively low, but the specificity is high at 94% as is the PPV of 90%. This enables appropriate therapy to be implemented early with a high degree of confidence in the majority of patients with disease, while continuing with culture in the PCR negative cases.

Limited sample volume may have accounted for the low sensitivity observed. No additional needle passes or aspirates were performed and the material collected was limited to the residual amount left in the needle. Sensitivity may be improved if an additional needle pass is performed to obtain material for PCR. No single diagnostic modality was adequately sensitive to enable it to be used alone. FNAB provides material for cytology, culture and PCR using the transport medium described. This is of particular value in resource limited countries where laboratories tend to be centralized. Cytology slides once prepared are stable at room temperature, as is the transport medium where the mycobacterial organism has been shown to remain viable for up to 7 days at room temperature, enabling both culture and PCR.

Ideally, the PCR technique could be refined, increasing the sensitivity and thereby eliminating the need for culture, which is costly and lengthy. Use of the transport bottle facilitates both culture and PCR analysis, but material may be deposited directly onto filter paper for subsequent PCR analysis. Refined techniques need to remain simple inexpensive and appropriate for use in routine laboratories in countries with limited resources and skills that bear the burden of this devastating and persistent disease.
ACKNOWLEDGEMENTS

Financial support was received from the NHLS Trust Fund, National Health Laboratory Services, South Africa and The Division of Molecular Biology and Human Genetics, Stellenbosch University.

We thank Prof Martin Kidd (Stellenbosch University) for statistical assistance

No industry funding was received to perform this study, and none of the authors have any commercial interest to declare.

This study was done in partial fulfillment of a PhD thesis (CA Wright, KGP Hoek)

REFERENCES


### Table 1

**Demographics and diagnostic outcome**

<table>
<thead>
<tr>
<th></th>
<th>Numbers</th>
<th>Percentage</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Number of cases</strong></td>
<td>104</td>
<td></td>
</tr>
<tr>
<td><strong>Age</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>≤ 2 years</td>
<td>6</td>
<td>5.8</td>
</tr>
<tr>
<td>&lt;13 years</td>
<td>22</td>
<td>21.2</td>
</tr>
<tr>
<td>&gt; 13 years</td>
<td>77</td>
<td>74</td>
</tr>
<tr>
<td>Unknown</td>
<td>5</td>
<td>5</td>
</tr>
<tr>
<td><strong>Gender</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Male</td>
<td>48</td>
<td>46.2</td>
</tr>
<tr>
<td>Female</td>
<td>51</td>
<td>49.0</td>
</tr>
<tr>
<td>Unknown</td>
<td>5</td>
<td>4.8</td>
</tr>
<tr>
<td><strong>HIV status</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Positive</td>
<td>36</td>
<td>34.6</td>
</tr>
<tr>
<td>Negative</td>
<td>12</td>
<td>11.5</td>
</tr>
<tr>
<td>Unknown</td>
<td>56</td>
<td>53.9</td>
</tr>
<tr>
<td><strong>Cytodiagnoses</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Malignancy</td>
<td>12</td>
<td>11.5</td>
</tr>
<tr>
<td>Reactive node</td>
<td>43</td>
<td>41.4</td>
</tr>
<tr>
<td>Other (e.g. cyst)</td>
<td>4</td>
<td>3.9</td>
</tr>
<tr>
<td>Cytology positive mycobacterial infection</td>
<td>45</td>
<td>43.3</td>
</tr>
<tr>
<td><strong>Cases with mycobacterial disease</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Culture and/or cytology positive (Reference Std)</td>
<td>54</td>
<td>100</td>
</tr>
<tr>
<td>Cytology positive</td>
<td>45</td>
<td>83.3</td>
</tr>
<tr>
<td>Culture positive</td>
<td>39</td>
<td>72.2</td>
</tr>
<tr>
<td>Culture positive and cytology positive</td>
<td>30</td>
<td>55.6</td>
</tr>
<tr>
<td>PCR positive mycobacterial infection</td>
<td>31</td>
<td>57.4</td>
</tr>
</tbody>
</table>
Figure 1. Epithelioid granuloma in an immune competent patient. (Papanicolaou, x400)

Figure 2. Abundant necrosis in which neutrophils and cellular debris are prominent consistent with tuberculous lymphadenitis in an immune compromised patient. (Papanicolaou, x400)
Figure 3. Modified ZN stain in a lymph node aspirate. (Ziehl-Neelson, x400)

Figure 4. Autofluorescence of mycobacteria in a lymph node aspirate.
(Papanicolaou, x1000 with a wide-band blue excitation filter)
Fig 5 Rotorgene software depicting the derivative melt peaks located within defined temperature bins. *M. tuberculosis* products melted at 90°C and *M. tuberculosis* complex at 86°C.

<table>
<thead>
<tr>
<th>No.</th>
<th>Colour</th>
<th>Name</th>
<th>Genotype</th>
<th>Peak 1</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Red</td>
<td>M. tuberculosis</td>
<td>MTB</td>
<td>89.98 (MTB)</td>
</tr>
<tr>
<td>2</td>
<td>Green</td>
<td>M. bovis</td>
<td>MTB complex</td>
<td>85.95 (MTBC)</td>
</tr>
</tbody>
</table>
CHAPTER 7

CONCLUSION

The aim of this dissertation was to demonstrate the value of Fine Needle Aspiration Biopsy in the diagnosis of mycobacterial lymphadenitis with particular reference to children.

Mycobacterial disease in children poses a particular diagnostic and management problem. It is a major cause of childhood morbidity and mortality in developing countries, where it is under reported due to problems in obtaining an accurate diagnosis. The traditional methods used to diagnose tuberculosis (TB) in adults are not feasible in children under the age of 7-8 years, young children experience the greater disease burden and a significant percentage (~10%) of children have extra pulmonary disease manifestations only. Clinical criteria and chest radiography have diagnostic limitations which are compounded in children who are HIV infected. In young children and in those with HIV/AIDS, early diagnosis is essential, as dissemination may occur rapidly due to their immature or compromised immune system.

We first established that TB lymphadenitis is a common clinical problem in children in TB endemic areas and that FNAB is an efficient, simple and effective diagnostic modality in determining the cause of lymphadenopathy in children.

We showed that FNAB provides superior diagnostic yields and a significant reduction in time to diagnosis when compared with conventional laboratory specimens in children.

Autofluorescence was demonstrated to be an accurate, rapid and inexpensive method for demonstrating mycobacteria on routinely stained cytology smears and this was confirmed in a prospective study in children where FNAB was shown to provide a rapid and definitive diagnosis in the majority of cases with TB lymphadenitis.
We developed a new reference standard for mycobacterial disease using FNAB-positive cytology (cytomorphological evidence of disease and identification of the organism) and/or positive culture.

Lymphadenopathy induced by BCG vaccination needs to be distinguished from TB lymphadenitis. Although not central to this thesis, a small study during this time period demonstrated cytomorphological features on FNAB which may assist in distinguishing between these two organisms, together with clinical features. (See Appendix A)

Although cytomorphology may confirm mycobacterial disease, organisms in the *M tuberculosis* complex are morphologically indistinguishable. This requires submission of material for culture and subsequent speciation or submission of material for direct PCR. As FNAB yields a miniscule amount of material, immediate bedside inoculation is required for culture. We developed an inexpensive liquid transport medium which facilitated culture from FNAB specimens, in which organisms remain viable for up to 7 days at room temperature. This is particularly useful in resource limited countries where laboratories are centralised and transport of specimens may be problematical.

Culture for speciation of mycobacterial organisms takes up to 6 weeks and this may result in inappropriate treatment or delayed commencement of therapy. Using the transport medium developed we established a rapid simple and inexpensive technique using PCR and High Resolution Melt Analysis, which is feasible for implementation in routine diagnostic laboratories.

In summary, the outcome of these studies has demonstrated the need for, and value of, FNAB in the diagnosis of mycobacterial lymphadenitis particularly in children in TB endemic areas.

We have improved the ability of cytology to confirm mycobacterial disease using autofluorescence. We have developed a means to obtain a culture from FNAB using a novel
transport medium and have developed a simple nucleic acid amplification technique which
will enable rapid confirmation and speciation of mycobacterial disease.

There are many challenges that remain, but we are working to develop an even simpler
technique for collecting material from FNAB for PCR/HRM analysis which would provide
higher sensitivity by removing inhibitors in the sample.

Conventional fluorescent microscopy using mercury vapour lamps is expensive and
therefore not feasible in resource-limited countries, and we intend to demonstrate the utility of
inexpensive rewindable LED light sources in autofluorescence and auramine stained FNAB
smears. A retrospective comparative study has already been completed. (see addendum A)

The quest to find and establish the ideal modality for diagnosing mycobacterial
disease has been a long one for many researchers. The many clinical manifestations of this
elusive disease and the reality that it is a disease that affects the poor and less influential
segments of the global population compounds the problem.

Currently a lot of attention is focused on identifying improved methods for TB
diagnosis. The field is ever-changing and no sooner has a technique been optimised when it is
evident that a better methodology has emerged. We hope that this research has contributed a
small step towards success in the fight against this devastating disease.
REFLECTIVE ASSESSMENT OF CONTRIBUTION.

In summary this thesis makes the following theoretical, empirical and methodological contributions to the literature.

**Theoretical contribution:**
FNAB is utilised in adult patients in neoplastic and infectious diseases, and to a lesser extent in paediatric oncology, but its potential contribution in paediatrics is not sufficiently appreciated. These results demonstrate the utility of FNAB in the diagnosis of TB, especially in children. The studies show that FNAB is equal to or superior to conventional diagnostic specimen types in children and that it is accurate, easy to perform, and yields rapid diagnostic information.

**Empirical contribution:**
Countries with low economic resources and high healthcare resource constraints bear the brunt of TB and HIV infections. This body of research demonstrates that FNAB can be implemented in these settings in the diagnosis of mycobacterial disease in children. It shows how FNAB and a novel transport medium solve problems in the collection of material for culture and NAAT, and autofluorescence assists in the cytological diagnosis of mycobacterial disease.

**Methodological contribution:**
The methodologies and concepts used in this thesis have been developed specifically for utilisation in resource limited countries. A new method of NAAT on FNAB material is developed and its benefits assessed. This new method is appropriate for use in resource limited countries. Although NAAT’s have
been used in the diagnosis of mycobacterial disease on FNAB, the results are extremely variable and impractical for implementation in routine diagnostic laboratories. The new method utilises high resolution DNA melt analysis was developed which is rapid inexpensive, minimises contamination and is applicable for implementation in routine diagnostic laboratories. This was performed on specimens collected in the mycobacterial transport medium.

A novel transport medium was developed to facilitate mycobacterial culture from FNAB specimens, particularly in rural hospitals and clinics. Autofluorescence is used in the research as an additional inexpensive and rapid means of bacteriological confirmation of mycobacterial disease on cytology smears, which was not previously published. This was then applied to fine needle aspirates in children with mycobacterial disease.
“I am not discouraged, because every wrong attempt discarded is another step forward”

Thomas A. Edison

I hope that this body of research will help others remember the inestimable honour we all have in serving humankind by undertaking research into the diseases plaguing our nation. All of us have an invaluable and unique role to play from the clinicians in their day-to-day contact with patients experiencing the frustration of practical problems associated with diagnosis and management of disease, to the biomedical scientists with out whose expertise solutions would not be possible. It is through collaboration and commitment to a common goal that we will achieve the Millennium Developmental Goal 6 of reducing the prevalence and death rate from tuberculosis by 2015 through early diagnoses of new cases.

The primary, the most urgent requirement is the promotion of education. It is inconceivable that any nation should achieve prosperity and success unless this paramount, this fundamental concern is carried forward. The principal reason for the decline and fall of peoples is ignorance. Today the mass of the people are uninformed even as to ordinary affairs, how much less do they grasp the core of the important problems and complex needs of the time.

It is therefore urgent that beneficial articles and books be written, clearly and definitely establishing what the present-day requirements of the people are, and what will conduce to the happiness and advancement of society. These should be published and spread throughout the nation, so that at least the leaders among the people should become, to some degree, awakened, and arise to exert themselves along those lines which will lead to their abiding honor. The publication of high thoughts is the dynamic power in the arteries of life; it is the very soul of the world. Thoughts are a boundless sea, and the effects and varying conditions
of existence are as the separate forms and individual limits of the waves; not until the sea boils up will the waves rise and scatter their pearls of knowledge on the shore of life.