

Lizard Epidermal Gland Secretions. II. Chemical Characterization of the Generation Gland Secretion of the Sungazer, *Cordylus giganteus*

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Abstract

In lizards, the epidermal glands of the femoral and precloacal regions are involved in the production of semiochemicals. In addition to its femoral glands, the giant girdled lizard, or sungazer, *Cordylus giganteus*, which is endemic to South Africa, has generation glands as an additional potential source of semiochemicals. These epidermal glands are described as glandular scales that overlay the femoral glands and are included in the normal epidermal profile located in the femoral (thigh) and anterior antebrachial (fore-leg) regions of the male sungazer.

GC-MS analysis of the generation gland secretions and the trimethylsilyl derivatives of some of the steroidal constituents was employed to identify 59 constituents, including alkenes, carboxylic acids, alcohols, ketones, aldehydes, esters, amides, nitriles, and steroids. The quantitative differences of the volatile constituents of the fore- and hind-leg generation glands were compared between individuals. This is the first report on the chemical composition of generation glandular material of lizards.

Several potential sources of pheromones in lizards have been described, including epidermal and cloacal glands and the blood–skin barrier.(1) A comprehensive review of the natural products originating from the integument of reptiles, including a summary of the chemicals secreted from the femoral (thigh) and precloacal (preanal) glands of a number of lizard species, was recently published.(2) The chemical characterization of the secretion of the femoral glands of the giant girdled lizard, or sungazer, *Cordylus giganteus* (Squamata, Cordylidae), located in the ventral femoral region, has also been reported.(2, 3) López and co-workers(4–7) recently summarized the potential functions of femoral gland semiochemicals, which could, for example, include mate choice, male–male competition, and interspecific recognition.

In addition to femoral glands, cordylid lizards have holocrine secretory cells located in the beta-layer of the epidermis, termed generation glands,(8, 9) which may occur in different body locations, for example, in the femoral, precloacal, antebrachial (fore-arm), and dorsal epidermal regions.(10) In most cases, the glandular material is stacked as a series of beta-layers.(10) In the case of *C. giganteus*, only males have generation glands, as patches of “glandular scales” anterior to the single row of femoral gland pores and on the ventral aspect of the fore-arm region (Figure 1).(10) In other cordylid species, females may also possess generation glands, the number which may vary according to geographical region.(11, 12)

It has been suggested that the generation glands could have a semiochemical function in combination with other sources such as the femoral glands.(12, 13) The purpose of this study was to identify the volatile constituents secreted from the generation glands by GC-MS analysis of the secretion and the trimethylsilyl derivatives of some of its constituents. Additionally, the quantitative differences of the volatile constituents of the fore- and hind-leg generation glands were compared between individuals. The histology of the generation glands of *C. giganteus* is also briefly described.

Results and Discussion

Basic Histology

In Figure 1 the generation glands in the ventral femoral region as well as the fore-leg antebrachial region can be clearly discerned from the normal epidermal scales. The histological profiles of both the femoral and generation gland (Figure 2) correspond to the description given by Van Wyk and Mouton.(10) The stacked beta-layer generations are observed in the generation glands located in the femoral region (Figure 2).

Typical of the beta-type generation glands is the absence of the outer oberhautchen layer, indicating that the outer layer of the generation series is exposed on a continuous basis. Although the generation glands on the fore-legs appear more swollen than those in the ventral femoral region, histological appearance is similar and the bulging is created by the underlying enlarged epidermal plates.(10)

Chemical Characterization

GC-MS analyses of the dichloromethane extracts of the glandular material showed that the samples collected from the fore- and hind-legs were qualitatively identical. The compounds identified in the material collected from the generation glands of *C. giganteus* are listed in Table 1. Of the 168 chromatographic peaks observed in the GC flame ionization detector (FID) analyses, 59 components, constituting more than 92% of the total amount of the detected compounds, could be analyzed with sufficient sensitivity utilizing GC-MS to yield mass spectra amenable to MS interpretation and comparison with Wiley, National Bureau of Standards (NBS), National Institute of Standards and Technology (NIST), and the in-house mass spectra libraries.

A typical GC-FID chromatogram is depicted in Figure 3. The structures of the majority of these compounds were confirmed by GC coelution with authentic reference compounds. Quantitative GC-FID analyses using solventless sample introduction revealed quantitative differences between fore- and hind-leg samples, as well as between individuals. The results from the quantitative analyses are summarized in Table 1. The EI-MS of constituents **5** and **7** displayed prominent ions corresponding to the general formulas $[C_nH_{2n-1}]^+$ and $[C_nH_{2n}]^+$ and decreasing in abundance with increasing mass, characteristic of monounsaturated alkenes and saturated aliphatic alcohols.(14)

Comparison of the mass spectra of these constituents with the available mass spectral libraries indicated that they could be either 1-alkenes or 1-alcohols. Although no molecular ions were present in their mass spectra, they were identified as 1-tetradecene (**5**) and 1-pentadecene (**7**) by co-injection of the generation gland extract with authentic synthetic samples of these compounds. Alkenes have been found in the skin of some lizards and snakes, but not in the secretions of lizards.(2) The occurrence of alkenes in the generation gland secretions could, however, be due to the fact that the generation glands are in fact glandular scales, forming part of the lizard's skin.

A number of additional compound classes, commonly found in lizard secretions,(2) including the femoral gland secretion of *C. giganteus*,(3) were also identified in its generation gland secretions, i.e., the C₉, C₁₀, and C_{12–22} unbranched saturated and unsaturated fatty acids **3**, **4**, **8**, **11**, **15**, **18**, **24**, **26**, **27**, **32**, **34**, **37**, **40**, **42**, and **44**, the C₁₂, C₁₄, C₁₅, and C₁₆ primary alcohols **6**, **12**, **16**, and **20**, the C₁₄, C₁₅, and C₁₇ methyl ketones **9**, **13**, and **22**, the C₉, C₁₀, and C_{14–17} saturated and unsaturated aldehydes **1**, **2**, **10**, **14**, **17**, and **19**, the methyl-C₁₆ and dodecyl-C₁₆ saturated and unsaturated esters **23**, **47**, and **48**, 11 steroids (**49–59**), squalene (**46**), and a γ -lactone, hexadecan-4-olide (**29**). It is interesting to note that cholesterol, one of the major components identified in the femoral gland secretions of *C. giganteus*,(3) does not occur in the generation gland material.

Hexadecanoic acid (**26**) is the major component of all the generation gland secretions analyzed. The identification of steroids **49–58** could not be confirmed by retention time comparison, due to the unavailability of standards. However, in some cases additional diagnostic information was obtained by the interpretation of the EI-mass spectra of these steroids employing published MS data. An interesting application of this technique is illustrated by the identification of the three cholestan-3-ol isomers **49**, **50**, and **51**, the mass spectra of which did not allow unambiguous differentiation between the three isomers.

However, some differences exist between the isomers with respect to the relative abundance of the ions formed by the elimination of water from the molecular ion due to the influence of the compounds' configuration on the mechanism of this elimination.(15, 16) In general, the mass spectra of C₁₇-substituted steroids with *cis*-A/B ring junctions (5 β -H) have prominent $[M - H_2O]^+$ ions, whereas the corresponding mass spectra of steroids with *trans*-

A/B ring junctions (5 α -H) have [M – H₂O]⁺⁺ ions with low relative abundances.(15) From this information it was deduced that constituent **49** is a 5 β -H isomer of cholestan-3-ol and that constituents **50** and **51** are the 5 α -H isomers. Comparison of the mass spectra of the TMS derivatives of these constituents with those in mass spectra libraries revealed that constituent **49** is probably 5 β -cholestan-3 α -ol, while confirming that constituents **50** and **51** are the two 5 α -H isomers of cholestan-3-ol.

The C-3 configuration of the latter two constituents could not be determined. The sterols **54**, **55**, **56**, and **58**, the identities of which were not confirmed by retention time comparison with standards, were positively identified on the basis of the good correlation of their mass spectra, as well as the mass spectra of their TMS derivatives, with those of the mass spectra libraries. Constituents **52**, **53**, and **57** were tentatively identified as 3-ethoxycholestane, cholestan-2/3-one, and cholest-4-en-3-one, respectively, by MS interpretation and comparison of their EI-mass spectra with mass spectra libraries, but their identities could not be confirmed.

The aliphatic nitriles hexadecanitrile (**21**) and octadecanitrile (**30**) and the aliphatic amides pentadecanamide (**28**), (Z)-9-hexadecenamide (**33**), hexadecanamide (**35**), heptadecanamide (**38**), octadecanamide (**41**), nonadecanamide (**43**), and icosanamide (**45**) have not been found in the secretions of any lizard species, although long-chain aliphatic amides have been identified in secretions from the scent glands of two snake species, Dumeril's ground boa, *Acrantophis dumerili*,(17) and the western diamondback rattlesnake, *Crotalus atrox*.(18) The scent glands produce secretions that contain chemicals that are, for example, used as allomones to repel predators.(2)

A few short-chain nitriles have been identified in the sternal gland secretions used by the male koala, *Phascolarctos cinereus*, for territorial marking.(19) Even though GC-MS analysis yielded good mass spectra for constituents **25**, **31**, **36**, and **39**, these constituents could not be identified by MS library comparison or the interpretation of their mass spectra, due to the paucity of relevant mass spectrometric information. The present investigation showed that the generation glands produce more diverse chemicals than the femoral glands, including groups of compounds (e.g., nitriles and amides) that are absent from the femoral gland secretions.(3)*C. giganteus* is the only cordylid that is recognized as a terrestrial grassland species, and the unique presence of generation glands on the fore-legs(10) may relate to marking of the grass substrate.

The fact that the chemical composition of the hind- and fore-leg generation glands is qualitatively identical supports the assumption that the glands on the fore-legs not only constitute a functional increase in the secretion volume but might also facilitate effective depositing of glandular material in the grassland environment. Although the quantities of chemicals produced by the generation glands differ between individual lizards, no pattern could be found in these differences (Table 1). The information made available in the present study should facilitate investigations into the semiochemical communication of *C. giganteus* and other species.

Experimental Section

General Experimental Procedures

All Pyrex glassware used for the chemical analysis was thoroughly cleaned with H₂O and acetone and heated to 500 °C to remove any residual organic material. Dichloromethane (Fluka, residue analysis grade) was used for extraction of organic material from collected samples. Syringes were cleaned with the same solvent. GC analyses were performed on a Carlo Erba 5300 gas chromatograph, using Borwin Intuitive Chromatography software, version 1.22, for data acquisition. The determination of the absolute abundance of constituents was done with an HP 5890 Series II GC, equipped with an autosampler.

Agilent Chemstation software, version A.09.01, was used for instrument control and data acquisition. These instruments were equipped with FIDs and split/splitless inlet systems, and their injectors and detectors were operated at 220 and 280 °C, respectively. Glass capillary columns (40 m × 0.3 mm i.d.) were coated with PS-089-OH, a silanol-terminated (95%)-methyl-(5%)-phenylpolysiloxane copolymer, at a film thickness of 0.25 μ m. Helium was used as carrier gas at a linear flow velocity of 28.6 cm/s. Samples were injected in the split mode using a split ratio of 1:10. The sample constituents were thermally focused on the column at a temperature of 30 °C, after which the column was heated ballistically to 40 °C and then programmed at 2 °C/min from 40 to 280 °C (hold 60 min).

EI-MS data were acquired at 70 eV on a Carlo Erba QMD 1000 GC-MS instrument, using the above-mentioned column and conditions, with He as carrier gas at an average linear velocity of 28.6 cm/s. Steroid derivatives were analyzed using a mass range of m/z 25 to 550. Ion source and interface temperatures of 200 and 250 °C, respectively, were used for all analyses. Some GC-MS analyses were done on a Fisons MD-800 instrument using the parameters specified above.

NMR spectra of synthetic reference compounds were recorded in CDCl₃ on Varian Unity INOVA 600 NMR, Varian Unity INOVA 400 NMR, and Varian 300 VNMRs instruments (Varian, Palo Alto, USA). Chemical shifts are given in parts per million (δ) relative to CHCl₃ (7.26 and 77.04 ppm for ¹H and ¹³C NMR, respectively).

Histology of the Generation Glands

Skin biopsies were obtained from specimens collected for studies regarding the reproductive cycle of these lizards.(20) Biopsies were subjected to routine histological procedures using paraffin wax (Merck, Histosec, 56–58 °C) as embedding medium. Sections (6–10 μ m) were cut with a rotary microtome and stained with hematoxylin (Merck) and eosin yellowish (Saarchem).(21)

Collection and Sample Preparation

Generation gland secretions were collected individually from the fore- and hind-legs of 10 adult male lizards in a study population on a farm in the Lindley district (Free State Province, South Africa; 28°01' S, 28°05' E) during April 1998. The secretions were collected by removing the external layers (reminiscent of shed skin) of the glandular scales on the fore- and hind-legs of the lizards, using a pair of sharp-tipped stainless steel forceps. The collected material was transported to Stellenbosch at –10 °C and stored at –70 °C until processed for analysis. The organic constituents were extracted from individually collected secretions with 100 μ L of DCM.

The extracts were subsequently centrifuged and concentrated to ca. 10 μ L by slow evaporation of the solvent at room temperature in a N₂ atmosphere.(22) Samples of 0.2 to 1.0 μ L of these concentrated extracts were used per analysis for qualitative GC-MS analyses. Quantitative GC determination of the relative abundance of the constituents of the secretions of the group of males was performed using solventless sample introduction(23) of the raw secretion. For the determination of the absolute abundance of each constituent in the generation gland secretion, extracts of the secretions from the fore- and hind-legs of a male lizard were prepared by extracting 11.3 and 10.3 mg of the respective secretions with four portions of 200 μ L of DCM each.

Each extract was slowly evaporated to dryness in an inert atmosphere at 22 °C, and the residual material was dissolved in 20 μ L of an internal standard solution containing 35.9 μ g of octadecane/mL in DCM. Quantities of 2 μ L of each of these solutions were used for quantitative GC determinations. Absolute constituent amounts were calculated proportional to that of the internal standard, using their chromatographic peak area ratios.

Trimethylsilyl (TMS) Derivatives

An extract of the generation gland material was concentrated in a Reacti-Vial left uncapped in a purified N₂ atmosphere to yield approximately 1 mg of solvent-free material. TMS derivatizations(24) were performed as previously described.(3)

Reference Compounds

Most of the reference compounds used to confirm the identity of the constituents of the generation gland secretions by retention time comparison were obtained commercially. Hexadecanitrile (**21**), octadecanitrile (**30**), pentadecanamide (**28**), (Z)-9-hexadecenamide (**33**), hexadecanamide (**35**), octadecanamide (**41**), icosanamide (**45**), dodecyl (Z)-9-hexadecenoate (**47**), and dodecyl hexadecanoate (**48**) were synthesized from authentic starting materials according to procedures exemplified by the syntheses of **21**, **28**, and **47**. (For other syntheses and the spectroscopic data of other compounds, see the Supporting Information.)

Hexadecanitrile (21)

was synthesized from 1-bromopentadecane by substitution of the bromine with cyanide.(25, 26) A mixture of 1-bromopentadecane (0.48 g, 1.65 mmol), NaCN (0.101 g, 2.06 mmol), and triethyleneglycol (ca. 1 mL) was stirred and slowly heated to 140 °C. After stirring the reaction mixture at this temperature for another 3 h, the mixture was added to H₂O (10 mL), and the organic product was extracted with CHCl₃.

The extract was washed with H₂O to neutral pH and dried on anhydrous MgSO₄, and the organic product was isolated in the usual manner to yield pure (GC-MS) hexadecanitrile (0.25 g, 64%): ¹³C NMR (100 MHz, CDCl₃) δ 14.06 (C-16), 17.06 (C-2), 22.65 (C-15), 25.35 (C-3), 28.63 (C-4), 28.73 (C-5), 29.27 (C-13), 29.33 (CH₂), 29.47 (CH₂), 29.56 (CH₂), 29.61 (CH₂), 29.62 (CH₂), 29.64 (CH₂), 29.65 (CH₂), 31.89 (C-14), 119.75 (C-1); EIMS *m/z* 222 (2), 208 (12), 194 (17), 180 (13), 166 (11), 152 (13), 138 (21), 124 (41), 110 (70), 97 (82), 82 (42), 69 (44), 57 (86), 43 (100), 29 (45).

Pentadecanamide (28)

was prepared from pentadecanoic acid via the corresponding acid chloride. (25, 26) Pentadecanoic acid (0.20 g, 0.825 mmol) was refluxed with thionyl chloride (1 mL, 13.8 mmol) for 30 min, after which the remaining thionyl chloride was distilled off. The acid chloride was cooled in an ice bath and treated with concentrated ammonia solution (3 mL, 25%). The resulting solid material was filtered and washed with H₂O containing a small percentage of EtOH.

The product was dried at room temperature on filtration paper, yielding the pure (GC-MS) pentadecanamide (0.18 g, 90%): ¹H NMR (400 MHz, CDCl₃) δ 0.87 (3H, t, *J* = 6.8, CH₃), 1.24 (22H, m), 1.62 (2H, m, H-3), 2.21 (2H, t, *J* = 7.8, H-2), 5.45 (1H, s, NH), 5.82 (1H, s, NH); ¹³C NMR (CDCl₃, 100 MHz) δ 14.14 (C-15), 22.71 (C-14), 25.55 (C-3), 29.25 (C-4), 29.35 (C-5), 29.38 (C-12), 29.49 (CH₂), 29.63 (CH₂), 29.67 (CH₂), 29.67 (CH₂), 29.69 (CH₂), 29.71 (CH₂), 31.95 (C-13), 36.00 (C-2), 176.17 (C-1); EIMS *m/z* 241 (1, M⁺), 212 (2), 198 (3), 184 (2), 170 (2), 156 (3), 142 (3), 128 (5), 114 (5), 97 (7), 86 (8), 72 (36), 59 (100), 43 (35), 29 (8).

Dodecyl (Z)-9-hexadecenoate (47)

was synthesized by refluxing a mixture of (Z)-9-hexadecenoic acid (25.4 mg, 0.01 mmol), dodecanol (18.4 mg, 0.01 mmol), and pTSA monohydrate (1 mg) in benzene (20 mL) for 30 h. (25)

The product was washed twice with H₂O and dried, and the solvent was removed on a rotary evaporator to yield practically pure (GC-MS) dodecyl (Z)-9-hexadecenoate in quantitative yield: ¹³C NMR (100 MHz, CDCl₃) δ 14.13 (C-12'), 14.15 (C-16), 22.69 (C-15), 22.72 (C-11'), 25.06 (C-3), 25.97 (C-3'), 27.20 (C-11), 27.26 (C-8), 28.69 (C-2'), 29.02 (C-4), 29.14 (CH₂), 29.17 (CH₂), 29.21 (CH₂), 29.29 (CH₂), 29.38 (CH₂), 29.56 (CH₂), 29.61 (CH₂), 29.67 (CH₂), 29.68 (CH₂), 29.72 (CH₂), 29.77 (CH₂), 31.82 (C-10'), 31.95 (C-14), 34.44 (C-2), 64.45 (C-1'), 129.78 (C-9), 130.01 (C-10), 174.03 (C-1); EIMS *m/z* 422 (1, M⁺), 255 (8), 236 (40), 194 (14), 152 (31), 125 (19), 111 (43), 98 (57), 97 (77), 83 (85), 71 (60), 69 (95), 57 (90), 43 (79), 41 (57), 29 (24).

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Supporting Information

Mass spectrometric data of identified constituents and spectroscopic data of synthesized compounds. This material is available free of charge via the Internet at <http://pubs.acs.org>.

Figures

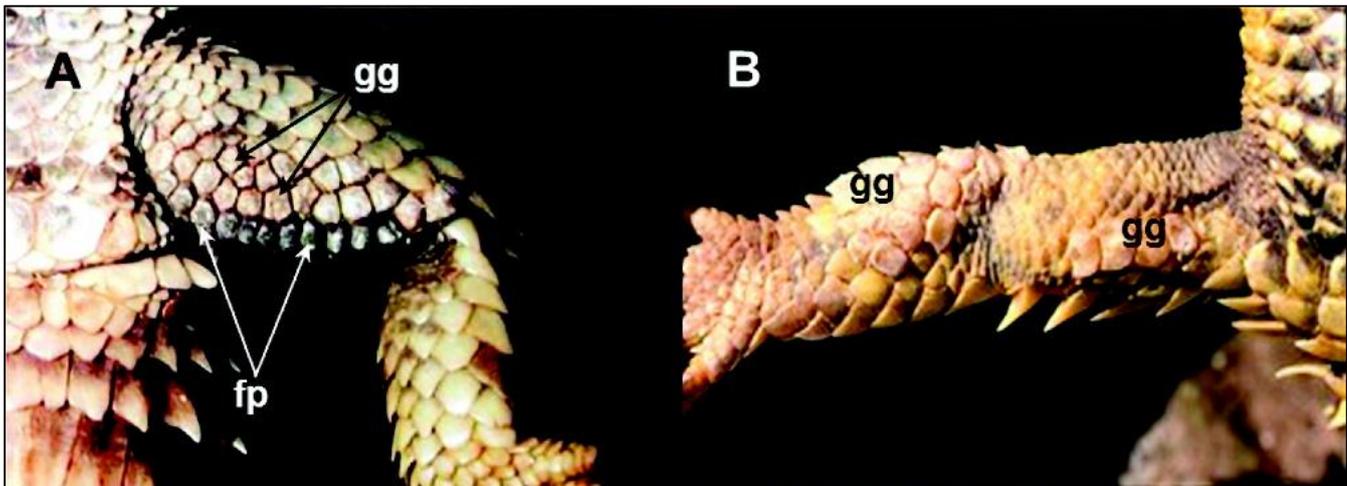


Figure 1. Generation glands in *C. giganteus* lizards. (A) Ventral view of the hind-leg of a male lizard showing a single row of femoral pores (fp) and an anterior patch of generation glands (gg). (B) Ventral view of the fore-leg of a male lizard showing groups of generation glands (gg).

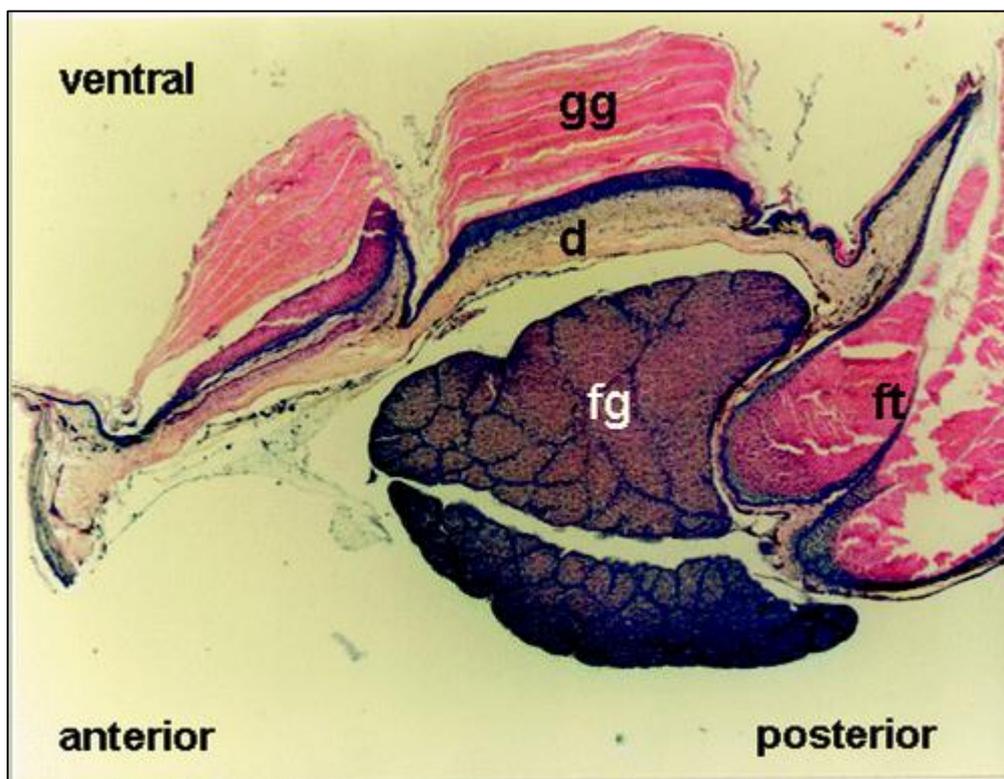


Figure 2. Histological cross-section of the epidermis in the ventral femoral region of an adult male *C. giganteus*. The femoral gland (fg) is located in the dermis (d), and the femoral gland tube (ft) opens on the ventral surface of the femoral region of the hind-leg. The stacked generation layers of the generation gland (gg) overlays the femoral gland as part of the epidermal profile.

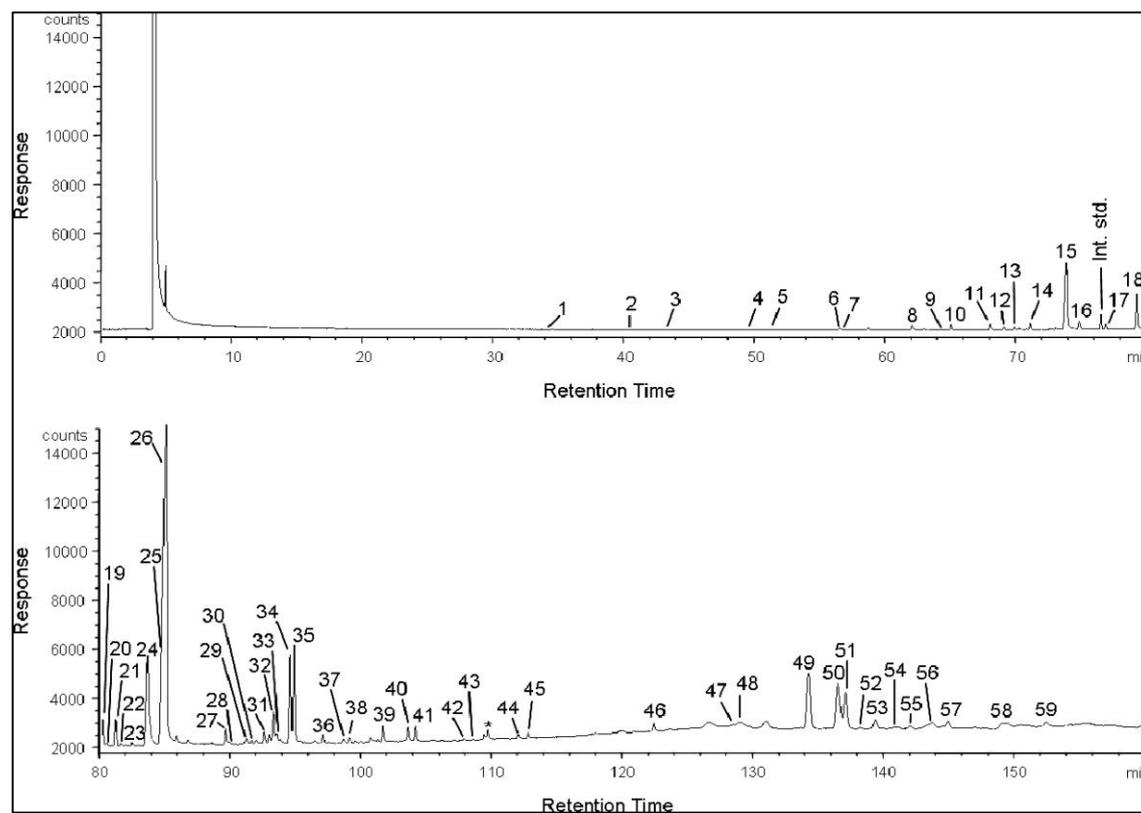


Figure 3. GC-FID chromatogram of an extract of fore-leg generation gland material of a male sungazer, *C. giganteus*. The analytical parameters are given in the text, and the identified compounds are numbered in order of elution from the GC column. *Impurity.

Tables

Table 1. Constituents Identified in the Generation Gland Secretion of the Male Sungazer, *C. giganteus*

number in Figure 3 ^a	constituent	ident. ^b	quantity (ng/mg) ^c		relative amounts ^d	
			fore-leg	hind-leg	fore-leg (n = 10)	hind-leg (n = 10)
Alkenes						
5	1-tetradecene	A, B	5	2	0.09 ± 0.06	0.02 ± 0.01
7	1-pentadecene	A, B	4	4	0.09 ± 0.03	0.04 ± 0.02
Acids						
3	nonanoic acid	A, B	2	3	0.01 ± 0.00	0.01 ± 0.01
4	decanoic acid	A, B	1	2	0.01 ± 0.00	<0.005
8	dodecanoic acid	A, B	33	31	0.19 ± 0.08	0.23 ± 0.12
11	tridecanoic acid	A, B	40	16	0.06 ± 0.03	0.06 ± 0.07
15	tetradecanoic acid	A, B	483	440	2.70 ± 0.54	2.27 ± 0.56
18	pentadecanoic acid	A, B	220	164	1.36 ± 0.71	0.84 ± 0.33
24	(Z)-9-hexadecenoic acid	A, B	838	1488	12.16 ± 6.92	13.75 ± 4.27
26	hexadecanoic acid	A, B	3479	3971	39.32 ± 3.54	38.30 ± 3.57
27	heptadecanoic acid	A, B	88	77	0.59 ± 0.26	0.42 ± 0.18
32	(Z)-9-octadecenoic acid	A, B	160	378	2.31 ± 1.11	2.80 ± 0.69
34	octadecanoic acid	A, B	501	715	6.72 ± 2.04	5.92 ± 1.36
37	nonadecanoic acid	A, C	34	29	0.01 ± 0.01	<0.005
40	icosanoic acid	A, B	88	132	0.45 ± 0.18	0.53 ± 0.19
42	heneicosanoic acid	A, B	15	21	0.05 ± 0.02	0.06 ± 0.01
44	docosanoic acid	A, B	28	39	0.10 ± 0.07	0.12 ± 0.05
Alcohols						
6	1-dodecanol	A, B	13	63	0.57 ± 0.51	1.06 ± 0.55
12	1-tetradecanol	A, B	16	6	0.13 ± 0.03	0.05 ± 0.02
16	1-pentadecanol	A, B	45	23	0.03 ± 0.01	0.03 ± 0.01
20	1-hexadecanol	A, B	21	18	0.03 ± 0.02	0.01 ± 0.01
Ketones						
9	2-tetradecanone	A, C	4	3	0.01 ± 0.01	0.01 ± 0.00
13	2-pentadecanone	A, B	12	28	0.09 ± 0.04	0.02 ± 0.01
22	2-heptadecanone	A, B	15	15	0.08 ± 0.04	0.05 ± 0.01
Aldehydes						
1	nonanal	A, B	1	1	0.02 ± 0.01	0.01 ± 0.01
2	decanal	A, B	2	1	0.01 ± 0.01	0.02 ± 0.04
10	tetradecanal	A, B	31	12	0.02 ± 0.01	0.01 ± 0.00
14	pentadecanal	A, C	36	12	0.03 ± 0.02	0.03 ± 0.03
17	hexadecanal	A, B	28	8	0.03 ± 0.07	0.01 ± 0.00
19	2-heptadecenal	A, C, E	145	192	0.57 ± 0.86	0.12 ± 0.16
Esters						
23	methyl hexadecanoate	A, B	19	2	0.02 ± 0.01	0.02 ± 0.02
47	dodecyl (Z)-9-hexadecenoate	A, B	29	17	0.03 ± 0.02	0.10 ± 0.06
48	dodecyl hexadecanoate	A, B	102	17	0.07 ± 0.12	0.08 ± 0.07
Nitriles						
21	hexadecanitrile	A, B	133	62	1.84 ± 0.92	0.41 ± 0.24
30	octadecanitrile	A, B	22	24	0.18 ± 0.06	0.06 ± 0.04

number in Figure 3 ^a	constituent	ident. ^b	quantity (ng/mg) ^c		relative amounts ^d	
			fore-leg	hind-leg	fore-leg (n = 10)	hind-leg (n = 10)
Amides						
28	pentadecanamide	A, B	16	15	0.12 ± 0.17	0.05 ± 0.02
33	(Z)-9-hexadecenamide	A, B	46	80	0.79 ± 0.16	0.73 ± 0.09
35	hexadecanamide	A, B	575	288	0.25 ± 0.14	0.19 ± 0.12
38	heptadecanamide	A, C	34	24	0.12 ± 0.10	0.05 ± 0.04
41	octadecanamide	A, B	83	78	0.45 ± 0.16	0.32 ± 0.11
43	nonadecanamide	A, C	11	8	0.02 ± 0.01	0.02 ± 0.00
45	icosanamide	A, B	32	26	0.24 ± 0.09	0.33 ± 0.12
Steroids						
49	5β-cholestan-3α-ol	A, D	609	1099	3.81 ± 1.18	5.16 ± 1.69
50	5α-cholestan-3α/β-ol	A, D	607	770	1.86 ± 0.73	2.02 ± 0.73
51	5α-cholestan-3α/β-ol	A, D	445	712	1.13 ± 0.35	1.90 ± 0.71
52	3-ethoxycholestane	A	25	18	0.10 ± 0.08	0.08 ± 0.05
53	cholestan-2/3-one	A	107	17	0.07 ± 0.03	0.17 ± 0.13
54	5α-cholest-7-en-3β-ol	A, D	57	54	0.06 ± 0.05	0.07 ± 0.03
55	5β-ergostanol	A, D	45	27	0.12 ± 0.05	0.21 ± 0.07
56	ergost-5-en-3β-ol	A, D	94	90	0.16 ± 0.11	0.32 ± 0.20
57	cholest-4-en-3-one	A	66	169	0.11 ± 0.10	0.17 ± 0.14
58	5β-stigmastanol	A, D	109	38	0.03 ± 0.01	0.05 ± 0.02
59	lanosterol	A, B, D	60	184	0.14 ± 0.07	0.31 ± 0.16
Miscellaneous						
29	hexadecane-4-olide	A, B	43	20	0.14 ± 0.07	0.10 ± 0.09
46	squalene	A, B	51	123	0.28 ± 0.17	0.69 ± 0.45
Unknown constituents						
25	m/z 39, 42, 57, 69, 71, 83, 85, 239	A	1088	1389	12.63 ± 1.14	12.30 ± 1.15
31	m/z 41, 43, 55, 57, 69, 71, 239	A	80	130	0.12 ± 0.04	0.05 ± 0.04
36	m/z 41, 43, 55, 57, 69, 71, 239	A	48	40	0.06 ± 0.04	0.03 ± 0.04
39	m/z 41, 43, 55, 60, 101, 114	A	99	58	0.35 ± 0.20	0.19 ± 0.16

a Compound classes given in order of elution from GC column.

b Identification method(s). A, GC-MS analysis; B, retention time comparison with authentic synthetic compounds; C, retention time increment comparison; D, EI-mass spectra of TMS derivatives; E, E/Z-isomerism unknown.

c Amount in ng of individual constituents per mg of secretion, calculated proportional to the amount of internal standard (octadecane) using their chromatographic peak area ratios.

d Relative amounts calculated as a percentage of the 168 compounds detected by GC-FID. The mean ± standard deviation is reported.

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