



# Microsatellite cross-species amplification and utility in southern African elasmobranchs: A valuable resource for fisheries management and conservation

Maduna *et al.*

RESEARCH ARTICLE

Open Access

# Microsatellite cross-species amplification and utility in southern African elasmobranchs: A valuable resource for fisheries management and conservation

Simo N Maduna, Charné Rossouw, Rouvay Roodt-Wilding and Aletta E Bester-van der Merwe\*

## Abstract

**Background:** Similarly to the rest of the world, southern Africa's diverse chondrichthyan fauna is currently experiencing high fishing pressures from direct and non-direct fisheries to satisfy market demands for shark products such as fins and meat. In this study, the development of microsatellite markers through cross-species amplification of primer sets previously developed for closely related species is reported as an alternative approach to *de novo* marker development. This included the design of four microsatellite multiplex assays and their cross-species utility in genetic diversity analysis of southern African elasmobranchs. As this study forms part of a larger project on the development of genetic resources for commercially important and endemic southern African species, *Mustelus mustelus* was used as a candidate species for testing these multiplex assays in down-stream applications.

**Results:** Thirty five microsatellite primer sets previously developed for five elasmobranch species were selected from literature for testing cross-species amplification in 16 elasmobranch species occurring in southern Africa. Cross-species amplification success rates ranged from 28.6%-71.4%. From the successfully amplified microsatellites, 22 loci were selected and evaluated for levels of polymorphism, and four multiplex assays comprising of the 22 microsatellites were successfully constructed, optimised and characterised in a panel of 87 *Mustelus mustelus* individuals. A total of 125 alleles were observed across all loci, with the number of alleles ranging from 3–12 alleles. Cross-species amplification of the four optimised multiplex assays was further tested on 11 commercially important and endemic southern African elasmobranch species. Percentage of polymorphism ranged from 31.8%-95.5% in these species with polymorphic information content decreasing exponentially with evolutionary distance from the source species.

**Conclusions:** Cross-species amplification of the 35 microsatellites proved to be a time- and cost-effective approach to marker development in elasmobranchs and enabled the construction of four novel multiplex assays for characterising genetic diversity in a number of southern African elasmobranch species. This study successfully demonstrated the usefulness of these markers in down-stream applications such as genetic diversity assessment and species identification which could potentially aid in a more integrative, multidisciplinary approach to management and conservation of commercially important cosmopolitan and endemic elasmobranch species occurring in southern Africa.

**Keywords:** Cross-species amplification, Microsatellites, Multiplex assays, Genetic diversity, Species identification, Conservation management

\* Correspondence: aeb@sun.ac.za  
Department of Genetics, Stellenbosch University, Private Bag X1, Matieland  
7602, South Africa

## Background

The subclass Elasmobranchii (sharks, skates and rays) comprises a diverse group of over 1000 species, and is representative of one of the most ancient extant vertebrate lineages [1]. Recently, pressures from direct and non-direct fisheries have resulted in the depletion of elasmobranch populations globally [2]. Decline in wild populations of elasmobranchs is further compounded by their life history traits that are more similar to those of mammals (e.g. low fecundity, late maturity and long gestation periods) than those of teleost fishes [3,4]. In comparison, elasmobranchs may not respond well to the high fishing pressures. This trend has been particularly pronounced for sharks due to unregulated harvesting to support an increase in demand for shark products (e.g. fins, meat, liver oil, skin and cartilage). A drastic reduction in population size (population bottleneck) can result in small populations experiencing the accumulating effects of inbreeding leading to severe loss of genetic diversity [5,6]. These trends have previously been reported for species such as the basking shark (*Cetorhinus maximus*) [7] and the narrownose smoothhound shark (*Mustelus schmitti*) [8]. Assessing genetic diversity and population structure of wild populations is therefore important for sustainable long-term management of the global shark fishery industry.

Misidentification of shark species in fisheries operations is also a widespread concern [9-12], and molecular individual identification methods have been developed to alleviate this problem [4,13-17]. To integrate genetic knowledge with fisheries management, it is imperative for shark fisheries to report shark landings by species instead of lumping them into species- or family groups (e.g. houndsharks, carcharinids, hammerheads etc.). This stems from the difficulties involved with unambiguously identifying species within and across families [13,18] e.g., carcharinids (*Carcharhinus brachyurus*, *C. obscurus* and *C. plumbeus*) and houndsharks (*Mustelus mustelus*, *M. palumbes* and *Galeorhinus galeus*) due to a high degree of conserved interspecific morphology [14]. Neglecting to report shark landings by species overlooks important differences in species susceptibility and population vulnerability to exploitation [15], and that in turn has important implications for species-specific conservation, management and trade monitoring programmes [19].

The general lack of molecular genetic markers (e.g. microsatellites) for many elasmobranch species impedes population and conservation genetic studies in that these markers can provide valuable information relating to population dynamics (spatial and temporal genetic variation) of individual species. Microsatellites are highly polymorphic due to their high mutation rate (between  $10^{-3}$  and  $10^{-4}$  mutations per gamete per generation) resulting in extensive length polymorphism [20,21]. This makes

microsatellite markers one of the most powerful molecular genetic tools with a remarkable array of applications ranging from genetic diversity [22,23] and population structure inference [24,25] to discerning genetic mating systems [26,27] and the identification of species [28-30].

Because the *de novo* development of microsatellites is challenging due to notoriously low rates of polymorphism in elasmobranchs [31], the development of microsatellite markers through cross-species amplification is the most effective alternative approach to *de novo* development of microsatellites and has recently also been reported in sharks [32]. Microsatellite cross-species amplification relies on the presence of conserved microsatellite flanking sequences [33], which in some organisms markedly demonstrate a high degree of conservation following millions of years of divergent evolution (e.g., 250 million years in sharks [28] and 470 million years in fish [34]). The success rate of microsatellite cross-species amplification has directly been correlated to the evolutionary distance between the source species and the target species [33,35].

The underrepresentation of endemic taxa in many cross-species amplification studies is unfortunate as endemics should be of great interest for conservation of biodiversity on a regional scale. Southern Africa has one of the most diverse chondrichthyan faunas in the world, consisting of some 181 species in 44 families of which 34 species are endemic to southern Africa [36,37]. Growing concerns regarding the sustainability of the southern African shark fishery, stemming from the local declines of cosmopolitan and endemic species, lead to stricter regulations being imposed so as to avert the collapse of natural populations [38-40]. Accordingly, we report here the development of microsatellite markers through cross-species amplification of species-specific primers from closely related species. This included the design and optimisation of four microsatellite multiplex assays and their cross-species utility in genetic diversity analysis of 11 southern African elasmobranch species.

## Results and discussion

### Cross-species amplification

Development of microsatellite loci through cross-species amplification proved useful in establishing genetic markers for shark species that are commercially important and those (typically endemics) that are indirectly affected by fisheries' operations. Amplification of the 35 microsatellites in 50 individuals from 16 different elasmobranch species (1-4 individuals per species) proved to be effective (Table 1). Cross-species amplification success rates or the percentage of microsatellites that amplified successfully ranged from 60.00%-71.40% in the Triakidae and Carcharhinidae families and from 28.57%-48.57% in the Scyliorhinidae, Sphyrnidae and Rajidae families (Figure 1). The higher success rates in the Triakidae and Carcharhinidae is

**Table 1 Cross-species amplification of the 35 microsatellites among 16 elasmobranch species of southern Africa**

Species	References	MM	MP	GG	SQ	CB	CL	CO	CP	HP	HE	PP	PA	SL	SZ	RS	RA
Loci		(n = 4)	(n = 3)	(n = 4)	(n = 1)	(n = 4)	(n = 3)	(n = 4)	(n = 4)	(n = 4)	(n = 4)	(n = 4)	(n = 1)	(n = 3)	(n = 3)	(n = 1)	(n = 3)
Mh1	[56]	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	++
Mh2	[57]	+	+	+	+	+	+	+	+	-	-	-	-	+	+	-	-
Mh6	[56]	-	-	+	-	+	-	-	-	-	-	-	-	-	-	-	-
Mh9	[57]	+	+	+	+	+	-	+	+	-	-	-	-	-	-	-	-
Mh25	[56]	+	+	+	+	+	+	+	+	+	+	+	+	-	+	+	++
Mca25	[49]	+	-	-	-	+	+	++	++	+	-	+	+	-	-	+	+
Mca31	[49]	-	-	-	+	+	+	+	-	-	-	-	-	-	+	-	-
Mca33	[49]	+	+	+	+	+	+	+	-	+	+	+	+	-	-	+	+
Mca44	[49]	++	-	+	-	-	-	-	-	+	+	+	+	-	-	+	++
McaB5	[49]	+	+	+	+	+	+	+	+	+	+	+	+	+	-	+	+
McaB6	[49]	+	-	+	+	+	+	+	+	+	+	-	-	+	-	-	-
McaB22	[49]	+	+	+	+	-	+	+	+	+	+	+	+	-	+	+	-
McaB27	[49]	+	+	+	+	+	-	-	+	-	+	-	-	-	+	-	-
McaB33	[49]	-	-	-	-	+	+	+	+	-	-	-	-	+	-	-	-
McaB35	[49]	+	+	-	+	-	-	-	+	-	+	+	+	-	-	-	++
McaB37	[49]	+	+	+	+	-	+	-	-	-	-	-	-	+	-	-	-
McaB39	[49]	+	-	+	-	+	+	+	+	+	+	+	+	-	-	+	-
Gg2	[50]	+	+	+	+	+	-	-	-	+	-	-	-	-	+	++	+
Gg3	[50]	+	+	+	+	+	+	+	+	+	+	-	-	+	-	++	+
Gg7	[50]	+	+	+	-	+	-	+	+	-	+	-	-	-	+	-	+
Gg11	[50]	+	+	+	++	++	+	++	++	+	+	-	-	-	+	-	-
Gg12	[50]	+	+	+	-	+	-	+	+	+	-	-	-	-	-	-	-
Gg15	[50]	+	+	+	+	+	+	++	++	+	+	+	+	+	+	+	-
Gg17	[50]	+	-	+	+	+	+	+	+	-	+	-	-	+	-	-	+
Gg18	[50]	+	+	+	+	+	+	++	+	+	-	-	-	-	++	+	+
Gg22	[50]	+	+	+	+	+	+	+	+	+	-	-	-	-	++	+	-
Gg23	[50]	+	+	+	+	+	+	+	+	-	-	-	-	+	++	+	+
Rp16-nfrdi	[44]	+	++	-	+	+	++	+	+	-	-	-	+	++	+	+	++
Rp35-nfrdi	[44]	-	+	-	++	-	+	-	-	-	-	-	+	+	++	+	++
Scan02	[45]	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
Scan06	[45]	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
Scan12	[45]	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
Scan14	[45]	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
Scan15	[45]	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
Scan16	[45]	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-

-, no visible band or faint bands with insufficient band intensity for scoring alleles were observed; +, solid bands with sufficient intensity for scoring alleles were detected; ++, solid bands with artefacts were produced but with at least one band of expected allele size. *Mustelus mustelus* (MM), *Mustelus palumbes* (MP), *Galeorhinus galeus* (GG), *Scylliogaleus queckettii* (SQ), *Carcharhinus brachyurus* (CB), *Carcharhinus limbatus* (CL), *Carcharhinus obscurus* (CO), *Carcharhinus plumbeus* (CP), *Haploblepharus pictus* (HP), *Haploblepharus edwardsii* (HE), *Poroderma africanum* (PA), *Poroderma pantherinum* (PP) *Sphyrna lewini* (SL), *Sphyrna zygaena* (SZ), *Raja straeleni* (RS) and *Raja alba* (RA).

expected as most microsatellites tested in this study were originally developed for species within the Triakidae family. Overall, the microsatellites showed less successful cross-species amplification to the taxa more divergent from the source species. Notably none of the individuals

showed PCR amplification at any of the six *Scyliorhinus canacula* microsatellites. This may in part be attributed to *S. canacula* being more distantly related to the study species. The mean genetic distance between the taxa was  $21.4 \pm 1.7\%$  (mean  $\pm$  SD) (*G. galeus* as source species;

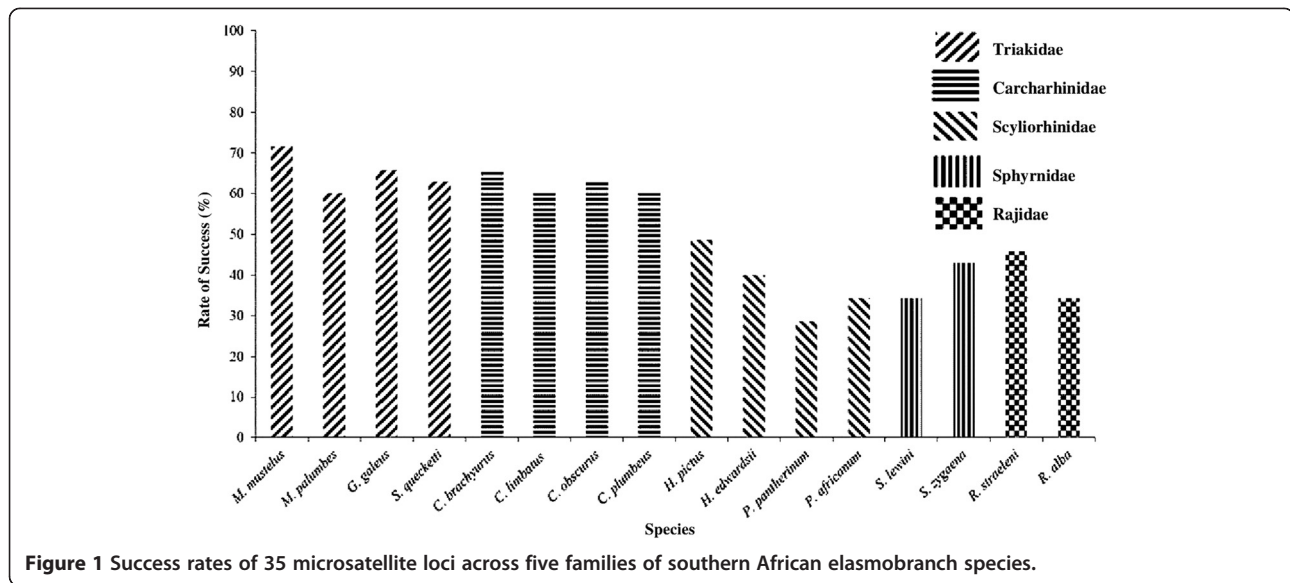
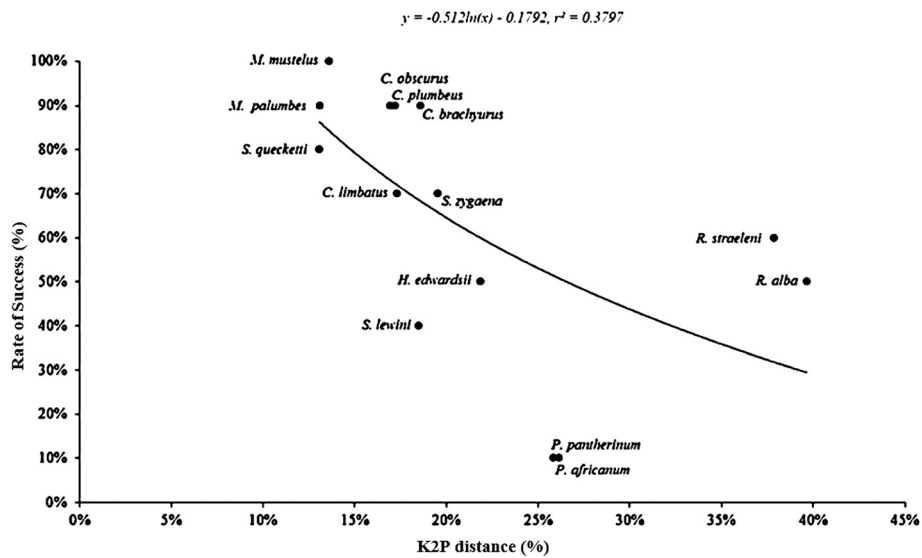


Figure 2) and  $18.7 \pm 1.5\%$  (*M. canis* as source species; Figure 3). *Haploblepharus pictus* could not be represented in the distance plot due to the lack of genetic information available in GenBank and Global Cestode Database: Elasmobranchs Specimens.

Results of cross-species amplification performance of *G. galeus* microsatellites exhibited a logarithmic regression function (Figure 2), albeit non-significant (slope within the 95% CI for no difference from zero), that may explain the general trend of negative correlation between cross-species amplification performance and genetic divergence seen across taxa [30,33,35,41]. Cross-species amplification of microsatellite markers from source to target species is generally negatively correlated with evolutionary divergence [30,42-45]. A similar trend was not observed for the *M. canis* microsatellites (Figure 3). This can be due to different life history traits (*i.e.* mating system and generation time) and genome size (*C* value) between the source and target species, which have been previously found to have significant negative effects on cross-species amplification success [33]. However, apart from the source-target species evolutionary distance other factors, such as mutations in microsatellite flanking sequences, may affect the success rate of cross-species amplification. Since microsatellites are usually found in non-coding regions where the substitution rate is higher than in coding regions [46], these microsatellite flanking sequences which serve as regions for PCR primer design and binding sites are prone to mutations [35]. Mutations (indels) in these regions may therefore result in null alleles and in turn affect the patterns of cross-species amplification as demonstrated in birds [47] and salmonids [48].

Additionally, *M. canis* microsatellite loci were isolated from an enriched genomic library [49] whereas for *G. galeus* the microsatellites were developed by a high-throughput sequencing approach (Roche 454 pyrosequencing) [50]. Based on the observed data it is hypothesised that these different approaches may have influenced the cross-species performance possibly due to the different resolving power of each approach for capturing microsatellites distributed across different parts of the genome. Castoe *et al.* [51] argues that enrichment-based approaches commonly use a few specific repeated motifs, which are largely selected without prior knowledge of their abundance in the genome and therefore could introduce potential bias in genome representativeness. In contrast, microsatellite identification from randomly sequenced genomic regions (*e.g.* Roche 454 NGS) allows for an unbiased assessment of all types of microsatellite loci present in a genome [51].

Cross-species amplification of orthologous microsatellites, due to the presence of conserved microsatellite flanking sequences [33], can persist over millions of years following divergent evolution as previously shown in sharks (250 million years [28]) and in fish (470 million years [34]). This indicates lower mutation rates within microsatellite flanking regions in aquatic organisms [30,34]. The reported cross-amplified microsatellite markers will thus contribute to establishing a molecular genetic marker repository for each of the Southern Africa elasmobranchs species included in this study. Future research efforts may be dedicated to generating microsatellite primers that have a high cross-species utility (*e.g.* [52]) as well as *in silico* mining of polymorphic microsatellite markers from expressed sequence tag data [53].

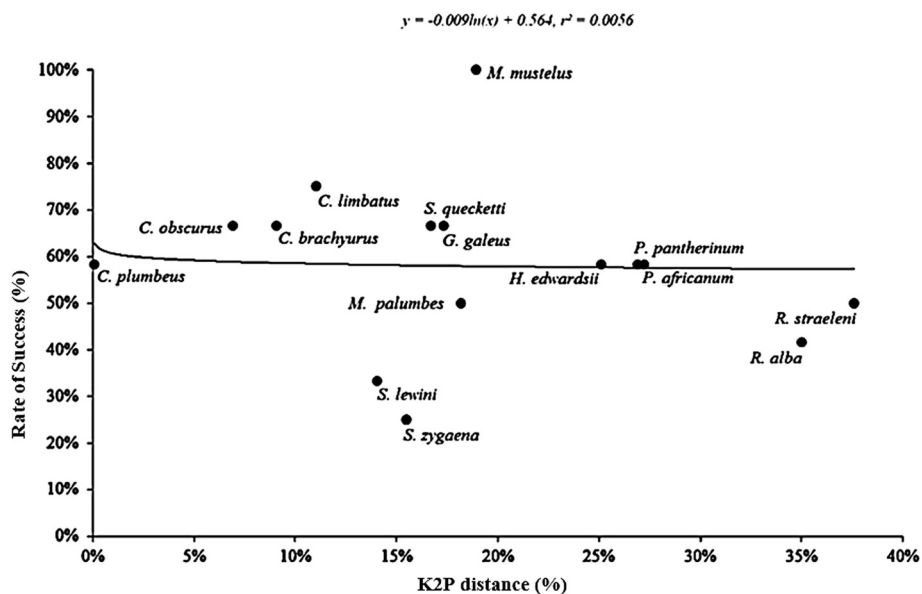


**Figure 2** Cross-species amplification performance of *Galeorhinus galeus* microsatellites in 15 of the 16 elasmobranch species, and genetic divergence between *G. galeus* and the target species based on *ND2* sequences.

### Multiplex assay characterisation

Twenty-two microsatellite loci that successfully cross-amplified across the study species and showed polymorphism in initial screening tests were used to develop four multiplex assays (MPS) comprised of at least five microsatellites each. These multiplex assays were characterised in a panel of 87 common smoothhound sharks (*Mustelus mustelus*) (Table 2). All 22 microsatellite loci were polymorphic across the multiplexes. With the

exception of one locus, *McaB22*, all the microsatellite loci were found to deviate significantly from Hardy-Weinberg equilibrium most likely due to Wahlund effect as samples were pooled from diverse geographical locations for analysis. MicroChecker detected no significant genotyping errors but indicated that null alleles were present at two loci (*Mh9* and *Gg7*). Slatkin's exact test for neutrality indicated that two loci (*McaB22* and *Gg3*) were candidates for being subjected to selection.



**Figure 3** Cross-species amplification performance of *Mustelus canis* microsatellites in 15 of the 16 elasmobranch species, and genetic divergence between *M. canis* and the target species based on *ND2* sequences.



**Table 2 Characterisation of four multiplex assays for *Mustelus mustelus* based on 87 individuals from southern Africa**

Locus	Microsatellite repeat motif	[P]	Dye	Size range (bp)	$A_N$	$A_E$	$H_O$	$H_E$	PIC	$F_{IS}$	$F_{NULL}$	$P_{E-W}$
Mh1	(AG) <sub>n</sub>	0.2	VIC	191-211	7	2.2	0.885	0.544	0.443	-0.633**	-0.223	0.931
Mh2	(GA) <sub>9</sub>	0.3	VIC	587-597	4	1.7	0.367	0.402	0.342	0.089**	0.023	0.688
Mh9	(GA) <sub>9</sub>	0.4	FAM	312-326	5	1.7	0.337	0.429	0.373	0.214**	0.062 <sup>b</sup>	0.723
Mh25	(CT) <sub>n</sub>	0.2	FAM	122-148	8	1.6	0.356	0.404	0.385	0.118**	0.032	0.802
Mca25	(CA) <sub>n</sub> (CT) <sub>n</sub>	0.2	PET	232-240	3	1.9	0.563	0.463	0.382	-0.217**	-0.070	0.226
McaB39	(CA) <sub>10</sub> GAT(AC) <sub>8</sub>	0.2	NED	177-212	3	2.0	1.000	0.509	0.384	-0.977**	-0.328	0.501
<b>MPS1 (mean)</b>	-	-	-	-	<b>5</b>	<b>1.9</b>	<b>0.585</b>	<b>0.459</b>	<b>0.385</b>	<b>-0.234</b>	<b>-0.084</b>	<b>0.645</b>
McaB5	(GT) <sub>11</sub>	0.2	VIC	189-210	10	3.5	0.826	0.716	0.674	-0.155*	-0.067	0.330
McaB6	(CA) <sub>10</sub>	0.2	FAM	226-266	9	3.3	0.756	0.702	0.655	-0.077*	-0.034	0.498
McaB22	(AC) <sub>18</sub>	0.2	NED	137-179	12	8.2	0.874	0.882	0.865	0.010	0.002	0.002
McaB27	(GT) <sub>6</sub>	0.2	PET	138-199	4	2.1	0.965	0.536	0.424	-0.808**	-0.282	0.589
Mca33	(ATC) <sub>5</sub>	0.2	FAM	189-199	6	3.0	0.872	0.674	0.609	-0.295**	-0.121	0.347
McaB37	(GT) <sub>5</sub>	0.2	NED	219-251	11	1.9	0.483	0.486	0.431	0.007**	-0.016	0.997
<b>MPS2 (mean)</b>	-	-	-	-	<b>9</b>	<b>3.7</b>	<b>0.796</b>	<b>0.666</b>	<b>0.610</b>	<b>-0.220</b>	<b>-0.086</b>	<b>0.461</b>
Gg2	(TG) <sub>n</sub>	0.2	NED	249-259	7	3.2	1.000	0.688	0.632	-0.458**	-0.188	0.324
Gg3	(GATT) <sub>n</sub>	0.2	PET	257-265	2	2.0	1.000	0.503	0.375	-1.000**	-0.333	0.001
Gg7	(AG) <sub>n</sub>	0.2	NED	296-312	4	1.6	0.310	0.393	0.343	0.212**	0.058 <sup>b</sup>	0.584
Gg11	(TCCC) <sub>n</sub>	0.2	NED	329-363	4	1.2	0.061	0.182	0.173	0.666**	0.000	0.792
Gg12	(TA) <sub>n</sub>	0.2	FAM	276-296	4	1.8	0.610	0.454	0.361	-0.347**	-0.110	0.807
<b>MPS3 (mean)</b>	-	-	-	-	<b>4.2</b>	<b>2.0</b>	<b>0.596</b>	<b>0.444</b>	<b>0.377</b>	<b>-0.185</b>	<b>-0.115</b>	<b>0.495</b>
Gg15	(GA) <sub>n</sub>	0.2	FAM	147-169	3	2.05	0.977	0.514	0.392	-0.910**	-0.308	0.370
Gg17	(AC) <sub>n</sub>	0.2	PET	159-181	3	1.02	0.023	0.023	0.023	-0.003**	0.000	1.000
Gg18	(GA) <sub>n</sub>	0.2	VIC	179-187	6	2.24	0.976	0.558	0.456	-0.759**	-0.272	0.776
Gg22	(GT) <sub>n</sub>	0.2	FAM	237-247	4	2.25	0.964	0.559	0.455	-0.733**	-0.263	0.488
Gg23	(AC) <sub>n</sub>	0.2	VIC	258-278	6	2.84	1.000	0.651	0.582	-0.540**	-0.214	0.562
<b>MPS4 (mean)</b>	-	-	-	-	<b>4.4</b>	<b>2.08</b>	<b>0.788</b>	<b>0.461</b>	<b>0.3816</b>	<b>-0.589</b>	<b>-0.211</b>	<b>0.615</b>
<b>Overall (mean)</b>	-	-	-	-	<b>5.7</b>	<b>2.4</b>	<b>0.691</b>	<b>0.512</b>	<b>0.444</b>	<b>0.010</b>	<b>-0.139</b>	<b>0.561</b>

Primer concentration in the final reaction as  $\mu\text{M}/\text{primer}$  ([P]); Number of alleles per locus ( $A_N$ ); effective number of alleles ( $A_E$ ); observed heterozygosity ( $H_O$ ); expected heterozygosity ( $H_E$ ); polymorphic information content (PIC); inbreeding coefficient ( $F_{IS}$ ) with statistically significant deviations from Hardy-Weinberg expectations indicated by \* ( $P < 0.01$ ) and \*\* ( $P < 0.001$ ); null allele frequency ( $F_{NULL}$ ) with <sup>b</sup> indicating the presence of null alleles at statistical significance at the 5% nominal level and Ewans-Watterson probability ( $P_{E-W}$ ). Mean values for each multiplex assay and overall are indicated in bold.

### Multiplex assay cross-species amplification and efficiency in species identification

Cross-species amplification of the four multiplex assays was tested for 11 other southern African shark species (Table 3). The number of alleles observed was highest in *G. galeus* and *M. palumbes*, varying from 1 to 7, while the percentage of polymorphism (PP) for each marker ranged from 31.8%-95.5%. The polymorphic information content (PIC) decreased exponentially with evolutionary distance from the source species (Table 3) and the four multiplex assays showed the highest PIC in *M. mustelus*, *M. palumbes* and *G. galeus*.

The mean genetic diversity estimates for each species in terms of number of alleles ( $A_N$ ), effective number of alleles ( $A_E$ ), observed heterozygosity ( $H_O$ ), expected heterozygosity ( $H_E$ ) and PIC are shown in Figure 4. In

group 1, the mean  $H_E$ ,  $A_E$  and PIC varied from relatively low in *C. brachyurus* (mean  $H_E = 0.230$ ; mean  $A_E = 1.4$ ; mean PIC = 0.181) to relatively high in *M. palumbes* (mean  $H_E = 0.653$ ; mean  $A_E = 3.3$ ; mean PIC = 0.606). Group 2 exhibited similar patterns of genetic diversity that varied from moderate in *S. zygaena* (mean  $H_E = 0.593$ ; mean  $A_E = 3.2$ ; mean PIC = 0.554) to relatively high in *P. pantherinum* (mean  $H_E = 0.662$ ; mean  $A_E = 3.4$ ; mean PIC = 0.603). For group 3 with  $n = 4$ , the mean  $H_E$ ,  $A_E$  and PIC ranged from relatively low in *C. plumbeus* (mean  $H_E = 0.249$ ; mean  $A_E = 1.5$ ; mean PIC = 0.193) to relatively high in *C. obscurus* (mean  $H_E = 0.429$ ; mean  $A_E = 2.1$ ; mean PIC = 0.367).

The genotypic distribution of the study species is depicted in Figure 5. Most of the study species could be differentiated on PC1 and PC2 of the PCoA plot as can

**Table 3 Multiplex transferability results of a total of 22 microsatellite loci showing the number of alleles per locus for the 11 elasmobranch species tested**

Locus	MP (n = 8)	GG (n = 8)	CB (n = 8)	CL (n = 4)	CO (n = 4)	CP (n = 4)	HP (n = 8)	HE (n = 4)	SL (n = 5)	SZ (n = 5)	PP (n = 5)
<b>MPS1</b>											
Mh1	4	3	1	2	1	1	1	1	4	2	3
Mh2	-	5	*	1	1	*	-	-	2	1	3
Mh9	4	4	*	-	*	*	-	-	<b>4</b>	5	<b>3</b>
Mh25	5	5	2	3	2	3	1	3	<b>4</b>	6	3
Mca25	3	1	1	3	2	1	2	1	<b>3</b>	<b>4</b>	3
McaB39	<b>3</b>	3	2	2	1	2	2	*	2	<b>3</b>	3
<b>MPS2</b>											
McaB5	3	1	2	4	2	3	1	1	<b>5</b>	<b>5</b>	7
McaB6	4	4	*	4	2	1	-	1	2	<b>3</b>	<b>5</b>
McaB22	2	1	1	4	2	4	2	4	<b>4</b>	7	7
McaB27	2	2	2	-	<b>1</b>	*	-	1	-	2	-
Mca33	4	2	2	4	2	2	2	3	<b>4</b>	<b>7</b>	6
McaB37	3	5	<b>1</b>	3	<b>1</b>	<b>1</b>	-	-	<b>4</b>	<b>6</b>	<b>7</b>
<b>MPS3</b>											
Gg2	5	4	1	-	<b>1</b>	<b>1</b>	2	-	-	2	-
Gg3	3	2	1	1	2	*	2	1	-	2	<b>2</b>
Gg7	4	1	1	-	1	1	<b>2</b>	-	-	1	1
Gg11	6	4	1	-	2	2	*	*	-	1	3
Gg12	4	5	-	-	1	*	1	<b>1</b>	2	2	<b>1</b>
<b>MPS4</b>											
Gg15	7	5	1	4	1	4	4	2	5	3	5
Gg17	4	4	1	4	1	2	<b>2</b>	2	2	3	2
Gg18	6	3	2	3	1	3	3	<b>3</b>	7	6	4
Gg22	6	5	2	3	2	2	2	1	3	2	3
Gg23	4	2	2	1	2	1	<b>3</b>	<b>2</b>	6	4	3
<b>Total P loci</b>	21	18	8	13	10	10	12	7	16	19	18
<b>PP</b>	95.5	81.8	36.4	59.1	45.5	45.5	54.5	31.8	72.7	86.3	81.8

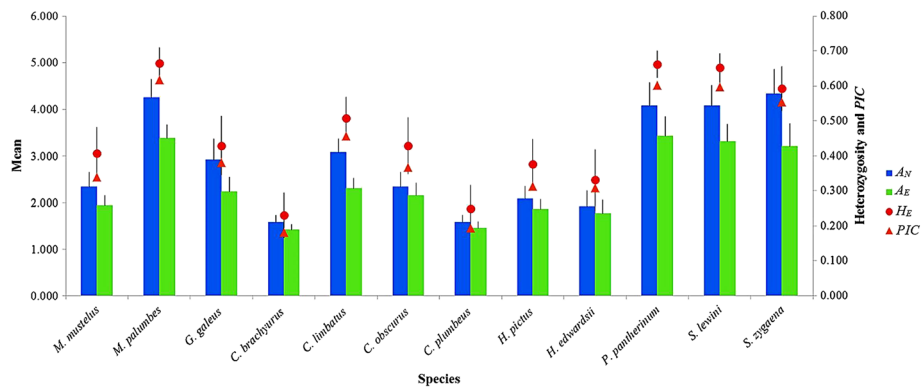
n - number of individuals tested; \*Failed to amplify but showed successful transferability initially (see Table 1); - No amplification; allele numbers in bold indicate loci that failed to cross-amplify according to Table 1; P - polymorphic and PP - percentage of polymorphism. For species abbreviations refer to Table 1.

be seen from individuals of each respective species clustering together. Individuals of the catshark species (*H. edwardsii*, *P. pantherinum* and *H. pictus*) however, were dispersed across quadrant 3 and 4. The PCoA also revealed that one of the *M. mustelus* individuals was misidentified as *G. galeus*. The identity of this particular individual was subsequently confirmed using the genetic identification method developed specifically for smooth-hound sharks [16]. Briefly, this method involves using four primers (1 universal forward primer and 3 species-specific reverse primers) for the mitochondrial gene, *nicotinamide adenine dehydrogenase subunit 2 (ND2)*, in a multiplex PCR reaction. The reverse primers amplify a fragment of different length for each species (*M.*

*asterias*, 564 bp; *M. mustelus*, 392 bp; *G. galeus*, 671 bp) and can therefore be utilised for distinguishing species based on fragment size.

In this study, the potential use of microsatellite loci in species identification was successfully demonstrated using shared microsatellite loci between species. The polymorphic information of these microsatellite loci was characterised by low genetic variation as previously proposed for elasmobranchs [31]. The genotypic distribution of the study species could also be differentiated based on PCoA analysis. Markedly, the lack of differentiation between the catshark species (*H. edwardsii* and *H. pictus*) on the PCoA plot may be explained by the misidentification of the *Haploblepharus* species that is a





**Figure 4** Mean genetic diversity estimates using 12 microsatellite loci shared between species: number of alleles ( $A_N$ ), effective number of alleles ( $A_E$ ), heterozygosity ( $H_E$ ) and polymorphic information content ( $PIC$ ). Error bars represent standard error.

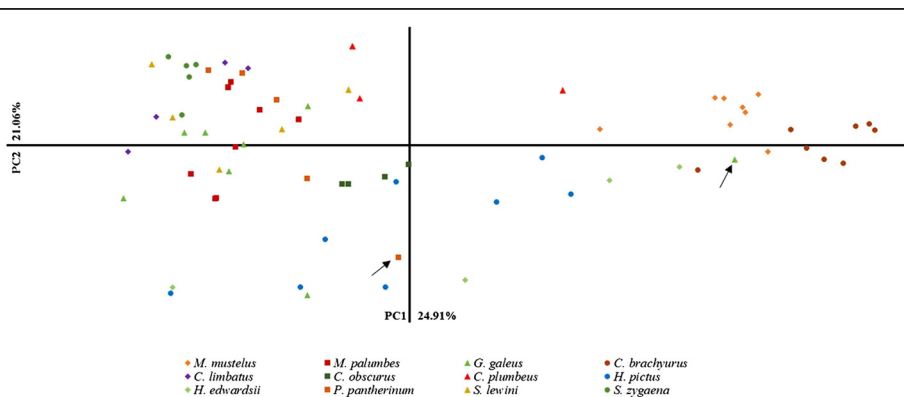
common occurrence in the catshark family [54]. To further investigate whether the lack of differentiation detected with microsatellites was indeed due to misidentification, the *cytochrome b* (*Cyt b*) and *ND2* genes were applied for species comparisons [54]. For both genes, sequence analyses revealed individuals with cryptic identification, suggesting that in the case of the catsharks, the microsatellites optimised in the current study were not successful in discriminating between the *Haploblepharus* species but could in the future aid in the identification of cryptic speciation within the catshark family.

In South Africa, the aforementioned misidentification issue is prominent in fishing operations (particularly in longline and trawl fisheries) where there is a high rate of incidental capture of non-target shark species [9-12]. This hinders the collection of reliable data on shark catch and trade on a species-specific basis making robust stock assessments and identification of overfished and potentially threatened species nearly impossible in most situations [4]. This was apparent in a study by Attwood *et al.* [12], which assessed bycatch in South Africa's inshore trawl fishery based on observer records.

In the aforementioned study, certain taxa were difficult to identify, and therefore taxonomic groups were lumped in species groups (e.g. *Raja* spp., *Mustelus* spp. and *Scyliorhinidae*), even though every attempt was made to analyse data at species level. Therefore, the molecular genetic markers developed by the current study may facilitate in obtaining species-specific catch data for stock assessment, characterising genetic diversity and delineating population genetic structure. This in turn will contribute to the implementation of future conservation and management plans on a species-specific level in southern Africa.

### Conclusions

Cross-species amplification of available microsatellite loci to target species has proven to be more time- and cost-effective in comparison to the *de novo* development approach and permitted the cross-amplification of 22 markers across 12 elasmobranch species. Cross-species amplification of the four multiplex assays developed in the current study highlighted the usefulness of microsatellites for characterising genetic diversity and potentially also species identification of a number of commercially



**Figure 5** Principle coordinates analysis (PCoA) of study species based on 12 shared amplified microsatellite loci between species. Arrows depict misidentified/mislabelled individuals.

important and endemic elasmobranch species. The molecular genetic markers developed in this study and their usefulness in down-stream applications could therefore aid in a more integrative, multidisciplinary approach to conservation management of elasmobranchs in southern Africa.

## Methods

### Ethics statement

The collection of specimens from various shark species used in this study complied with the Convention on Biological Diversity (<http://www.cbd.int/convention/>) and the Convention on the Trade in Endangered Species of Wild Fauna and Flora (<http://www.cites.org/>). All permits to collect finclip or muscle tissue for research purposes were granted by the Department of Agriculture, Forestry and Fisheries (Republic of South Africa).

### Study species and DNA extraction

Sixteen elasmobranch species occurring in southern African waters belonging to five families within two orders were selected for cross-species amplification (Additional file 1). Where possible, specimens were collected from at least two sampling locations to better capture allelic diversity present within populations of each respective species. However, due to opportunistic sampling for a majority of the study species, samples were obtained from only a single location (Additional file 1). Muscle tissue or finclips were preserved in 99% ethanol and stored at room temperature until further use. Total genomic DNA was isolated using the standard cetyltrimethylammonium bromide (CTAB) method of Saghai-Marooof *et al.* [55]. The extracted DNA was quantified using a NanoDrop ND-1000 spectrophotometer v.3.0.1 (*NanoDrop*<sup>®</sup>). For testing cross-species amplification, each DNA sample was adjusted to a working concentration of 50 ng/μl and stored at -20°C prior to polymerase chain reaction (PCR) analysis.

### Microsatellite primer transfer

A total of 35 microsatellite markers previously developed in five elasmobranch species (*Raja pulchra* [44], *Scyliorhinus canacula* [45], *M. canis* [49], *G. galeus* [50] and *M. henlei* [56,57]) were selected for testing cross-species amplification. Primer sequences and annealing temperature ( $T_A$ ) of each primer set optimised for each respective source species are shown in Table S2 (Additional file 2). Polymerase chain reaction conditions optimised for the majority of the source species [*G. galeus*, *M. henlei*, *R. pulchra* and *S. canacula*] were applied for cross-species microsatellite examinations except for the *M. canis* ([56] PCR protocol) and some *R. pulchra* (modified [57] PCR protocol) primer sets. Polymerase chain reactions for all individuals were executed in a GeneAmp<sup>®</sup> PCR System 2700.

The PCR amplicons were visualised on a 2% agarose gel stained with ethidium bromide together with negative controls and Promega 100 bp molecular size ladder for preliminary size determination. Success or failure of PCR amplification in cross-species trials was determined simply on the basis of whether band intensity was sufficient to score alleles. In most instances, less stringent PCR conditions were not employed in the cross-species assays so as to minimise the risk of amplification of non-orthologous loci in the target species. The number of markers that showed amplification success at all or a percentage of individuals in the target species (“+/++” in Table 1) were counted as an index to measure the cross-species microsatellite amplification performance.

### Multiplex design and optimisation

As this study forms part of a larger project on the development of genetic resources for commercially important and endemic species of southern Africa, *Mustelus mustelus* was used as a candidate species for testing of the four novel multiplex assays. Levels of polymorphism were initially assessed at all the successfully cross-amplified microsatellite loci in a panel of eight individuals of *M. mustelus*. Amplicons were subjected to electrophoresis for two hours at 150 volts on a 12% polyacrylamide gel to detect size variants. Microsatellites were considered to be polymorphic when two bands were distinguishable in a single individual (*i.e.* heterozygote) and/or clear size differences were detected between different individuals.

Twenty-two polymorphic microsatellite loci were selected, and primers fluorescently labelled and optimised in four multiplex assays (5–6 loci per MPS) using a strategy outlined by Guichoux *et al.* [58] with one of the following dyes: FAM, VIC, PET, or NED. The use of different dyes was to facilitate co-amplification of multiple microsatellite markers in a single reaction for cost- and time-efficient genotyping (Multiplex PCR).

After optimisation of the newly designed MPS (MPS1, MPS2, MPS3 and MPS4), a panel of 87 *M. mustelus* individuals from across the distribution range in southern Africa was genotyped for marker characterisation purposes. The multiplex assays were then tested on 11 additional species to show their overall application in genetic diversity and population structure analysis. Finally, a total of 12 microsatellite loci that were successfully genotyped across the study species (*Mh1*, *Mh25*, *Mca25*, *McaB39*, *McaB5*, *McaB22*, *Mca33*, *Gg15*, *Gg17*, *Gg18*, *Gg22* and *Gg23*) were selected to demonstrate the potential use of microsatellite loci in species identification.

The percentage of polymorphism (*PP*) was calculated using the formula:

$$PP = \frac{N_P}{N_T} \times 100$$

where  $N_p$  is the total number of polymorphic loci and  $N_T$  is the total number of loci multiplied by 100.

For the multiplex reaction, the Qiagen Multiplex PCR kit was used and PCR conducted according to the manufacturer's instructions except for varying  $T_A$ , 59°C for MPS1, MPS3 and MPS4; and 56°C for MPS2. For subsequent analysis on an ABI 3730XL DNA Analyzer, PCR products were diluted in distilled water and fragment analysis performed together with the LIZ600 internal size standard. Individual genotypes were scored based on fragment size via Peak Scanner® software v.1 (Life Technologies). Auto Bin v.0.9 macro for Excel (<http://www.bordeaux-aquitaine.inra.fr/biogeoco/Ressources/Logiciels/Autobin>; see [58]) was used to detect discreet size variants where allele binning of genotype data obtained from Peak Scanner® software v.1 was based on raw size.

### Genetic diversity analysis

MicroChecker v.2.2.3 [59] was used to evaluate the presence of genotypic errors caused by allele dropout, stuttering and null alleles. Null allele frequencies ( $F_{rNULL}$ ) were calculated using the Brookfield 1 estimator implemented in this program. Locus-specific fixation index ( $F_{IS}$ ) and over all loci was estimated to measure departure from Hardy-Weinberg equilibrium using the exact probability test (20 batches, Dememorization; 10000 and 5000 iterations) using Genepop v.4.0 [60]. Linkage disequilibrium between all pairs of loci was calculated using an exact test implemented also in Genepop. Slatkin's exact test (1000 permutations) for neutrality, based on Ewens-Watterson sampling theory [61] was used to detect loci under selection as implemented in Arlequin v.3.5.1.2 [62]. The number ( $A_N$ ) of alleles at each microsatellite locus, as well as the effective number of alleles  $A_E$ :

$$A_E = 1 / \sum_{i=1}^n p_i^2$$

where  $p_i$  is the frequency of the  $i^{\text{th}}$  allele and  $n$  is the number of alleles was calculated using the GenAlEx v.6.5 program [63]. The proportion of individual samples that were heterozygous [direct count heterozygosity ( $H_O$ ) and expected under Hardy-Weinberg equilibrium ( $H_E$ )] was calculated using MsatTools [64]. MsatTools was also used to calculate the polymorphic information content ( $PIC$ ) of each marker according to the following equation in [65]:

$$PIC = 1 - \sum_{i=1}^n p_i^2 - 2 \left[ \sum_{i=1}^{n-1} \sum_{j=i+1}^n p_i^2 p_j^2 \right] \text{ where } p_i \text{ and } p_j \text{ are}$$

the frequency of the  $i^{\text{th}}$  and  $j^{\text{th}}$  allele respectively and  $n$  is the number of alleles.

Direct comparison of genetic diversity estimates ( $H_E$ ,  $A_E$  and  $PIC$ ) across the 11 species was not plausible due to

the different sample sizes that were used. Species were therefore grouped into three groups according to sample size: (1) *M. mustelus*, *M. palumbes*, *G. galeus*, *C. brachyurus* and *H. pictus* ( $n = 8$ ); (2) *P. pantherinum*, *S. lewini* and *S. zygaena* ( $n = 5$ ) and (3) *C. obscurus*, *C. limbatus*, *C. plumbeus* and *H. edwardsii* ( $n = 4$ ). The potential use of microsatellite data for species-assignment was assessed through principle coordinate analysis (PCoA) in GenAlEx v.6.5 using genetic distances between individuals.

To evaluate cross-species amplification performance, DNA sequences derived from the mitochondrial *ND2* gene (1044 bp) of each species were downloaded from GenBank and Global Cestode Database: Elasmobranchs Specimens (<http://elasmobranchs.tapewormdb.uconn.edu>) (Additional file 3). The genetic distance of the study taxa was estimated using the Kimura 2-parameter model with the rate variation among sites modelled with a gamma distribution (shape parameter = 5) implemented in MEGA v.5 [66].

### Additional files

**Additional file 1:** The 16 elasmobranch species of southern Africa selected for cross-species amplification, including family, species, distribution and sampling locations.

**Additional file 2:** The 35 putative microsatellite markers developed from five closely related species for cross-species amplification in the study taxa, including the primers sequence, microsatellite repeat motif, annealing temperature ( $T_A$ ) and GenBank accession numbers.

**Additional file 3:** The *ND2* sequence information of the study taxa used to estimate the genetic distance to evaluate cross-species performance, including ID Verified, availability of images (yes or no) which are available in the on-line host specimen database (<http://elasmobranchs.tapewormdb.uconn.edu>) and GenBank accession numbers.

### Abbreviations

CI: Confidence interval; PP: Percentage of polymorphism; P: Polymorphic; PIC: Polymorphic information content;  $A_N$ : Number of alleles;  $A_E$ : Effective number of alleles;  $H_O$ : Observed heterozygosity;  $H_E$ : Expected heterozygosity;  $F_{IS}$ : Inbreeding coefficient;  $F_{rNULL}$ : Null allele frequency;  $P_{E-W}$ : Ewens-Watterson probability; K2P: Kimura 2-parameter model; *ND2*: Nicotinamide adenine dehydrogenase subunit 2; MM: *Mustelus mustelus*; MP: *Mustelus palumbes*; GG: *Galeorhinus galeus*; SQ: *Scylliogaleus quecketti*; CB: *Carcharhinus brachyurus*; CL: *Carcharhinus limbatus*; CO: *Carcharhinus obscurus*; CP: *Carcharhinus plumbeus*; HP: *Haploblepharus pictus*; HE: *Haploblepharus edwardsii*; PA: *Poroderma africanum*; PP: *Poroderma pantherinum*; SL: *Sphyrna lewini*; SZ: *Sphyrna zygaena*; RS: *Raja straeleni*; RA: *Raja alba*.

### Competing interests

The authors have no competing interests to declare.

### Authors' contributions

SNM performed cross-species amplification, microsatellite genotyping, genetic data analyses, organised the samples and drafted the manuscript. CR participated in cross-species amplification and microsatellite genotyping. RR-W participated in experimental design and coordination and contributed to manuscript preparation. AEB-vdM conceived the study, provided funds, participated in its design and coordination and contributed to manuscript preparation. All authors read and approved the final manuscript.

## Acknowledgements

The authors gratefully acknowledge and thank the KwaZulu-Natal Sharks Board, Oceans Research, South African Department of Agriculture, Forestry and Fisheries (DAFF), South African Shark Conservancy and White Shark Africa for providing valuable samples. The authors also thank Michelle-Angelique Hallendorff for editorial revision and Clint Rhode and Luca Mirimin for valuable discussions. This work was funded by the National Research Foundation, South Africa.

Received: 7 February 2014 Accepted: 2 June 2014

Published: 10 June 2014

## References

- Corrigan S, Beheregaray LB: **A recent shark radiation: molecular phylogeny, biogeography and speciation of wobbegong sharks (family: Orectolobidae).** *Mol Phyl Evol* 2009, **52**:205–216.
- Worm B, Davis B, Kettner L, Ward-Paige CA, Chapman D, Heithaus MR, Kessel ST, Gruber SH: **Global catches, exploitation rates, and rebuilding options for sharks.** *Mar Policy* 2013, **40**:194–204.
- Stevens JD, Bonfil R, Dulvy NK, Walker PA: **The effects of fishing on sharks, rays, and chimaerans (Chondrichthyes), and the implications for marine ecosystems.** *ICES J Mar Sci* 2000, **57**:476–494.
- Shivji M, Clarke S, Pank M, Natanson L, Kohler N, Stanhope M: **Genetic identification of pelagic shark body parts for conservation and trade monitoring.** *Conserv Biol* 2002, **16**:1036–1047.
- Nei M, Maruyama T, Chakraborty R: **Bottleneck effect and genetic variability in populations.** *Evolution* 1975, **29**:1–10.
- Glenn TC, Stephan W, Braun MJ: **Effects of a population bottleneck on whooping crane mitochondrial DNA variation.** *Conserv Biol* 1999, **13**:1097–1107.
- Hoelzel AR, Shiviji MS, Magnussen J, Francis MP: **Low worldwide genetic diversity in the basking shark.** *Biol Lett* 2006, **2**:630–642.
- Pereyra S, García G, Miller P, Oviedo S, Domingo A: **Low genetic diversity and population structure of the narrownose shark (*Mustelus schmitti*).** *Fish Res* 2010, **106**:468–473.
- Myers RA, Worm B: **Rapid worldwide depletion of predatory fish communities.** *Nature* 2003, **423**:280–283.
- Barker MJ, Schluessel V: **Managing global shark fisheries: suggestions for prioritising management strategies.** *Aquat Conserv Mar and Freshwat Ecosyst* 2005, **15**:325–347.
- Petersen SL, Honig MB, Ryan PG, Underhill LG, Compagno LJV: **Pelagic shark bycatch in the pelagic longline fishery off Southern Africa.** In *Understanding and Mitigating Vulnerable Bycatch in Southern African Trawl and Longline Fisheries*. Edited by Petersen SL, Nel DC, Ryan PG, Underhill LG; 2008. WWF South Africa Report Series - 2008/Marine/002.
- Attwood CG, Peterson SL, Kerwath SE: **Bycatch in South Africa's inshore trawl fishery as determined from observer records.** *ICES J Mar Sci* 2011, **68**:2163–2174.
- Pank M, Shivji MS, Stanhope M, Natanson L, Kohler N: **Rapid and simultaneous identification of body parts from the morphologically similar sharks *Carcharhinus obscurus* and *Carcharhinus plumbeus* (Carcharhinidae) using multiplex PCR.** *Mar Biotechnol* 2001, **3**:231–240.
- Domingues RR, de Amorim AF, Hilsdorf AWS: **Genetic identification of Carcharhinus sharks from the southwest Atlantic Ocean (Chondrichthyes: Carcharhiniformes).** *J Appl Ichthyol* 2013, **29**:738–742.
- Abercrombie DL, Clarke SC, Shivji MS: **Global-scale genetic identification of hammerhead sharks: Application to assessment of the international fin trade and law enforcement.** *Conserv Genet* 2005, **6**:775–788.
- Farrell ED, Clarke MW, Mariani S: **A simple genetic identification method for northeast Atlantic smoothhound sharks (*Mustelus* spp.).** *ICES J Mar Sci* 2009, **66**:561–565.
- Naylor GJP, Cairn JN, Jensen K, Rosana KAM, White WT, Last PR: **A DNA sequence based approach to the identification of shark and ray species and its implications for global elasmobranch diversity and parasitology.** *Bull Am Mus of Nat Hist* 2012, **367**:1–262.
- Da Silva C, Bürgener M: **South Africa's demersal shark meat harvest.** *Traffic Bulletin* 2007, **21**:55–65.
- Clarke SC: **Understanding pressures on fishery resources through trade statistics: A pilot study of four products in the Chinese dried seafood market.** *Fish Fish* 2004, **5**:53–74.
- Tautz D: **Hypervariability of simple sequences as a general source for polymorphic DNA markers.** *Nucleic Acids Res* 1989, **17**:6463–6471.
- Weber JL, Wong C: **Mutation of human short tandem repeats.** *Hum Mol Genet* 1993, **2**:1123–1128.
- Edwards CJ, Soulsbury CD, Statham MJ, Ho SYW, Wall D, Dolf D, Iossa G, Baker PJ, Harris S, Sacks BN, Bradley DG: **Temporal genetic variation of the red fox, *Vulpes vulpes*, across western Europe and the British Isles.** *Quat Sci Rev* 2012, **57**:95–104.
- Ozerov MY, Veselov AE, Lumme J, Primmer CR: **Temporal variation of genetic composition in Atlantic salmon populations from the Western White Sea Basin: influence of anthropogenic factors?** *BMC Genet* 2013, **14**:88.
- Urian KW, Hofmann S, Wells RS, Read AJ: **Fine-scale population structure of bottlenose dolphins (*Tursiops truncatus*) in Tampa Bay, Florida.** *Mar Mamm Sci* 2009, **25**:619–638.
- Mourier J, Mills SC, Planes S: **Population structure, spatial distribution and life-history traits of blacktip reef sharks *Carcharhinus melanopterus*.** *J Fish Biol* 2013, **82**:979–993.
- Mobley KB, Amundsen T, Forsgren E, Svensson PA, Jones AG: **Multiple mating and low incidence of cuckoldry for nest-holding males in the two-spotted goby, *Gobiusculus flavescens*.** *BMC Evol Biol* 2009, **9**:6.
- Boomer JJ, Harcourt RG, Francis MP, Walker TI, Braccini JM, Stow AJ: **Frequency of multiple paternity in gummy shark, *Mustelus antarcticus*, and rig, *Mustelus lentiginatus*, and the implications of mate encounter rate, postcopulatory influences, and reproductive mode.** *J Hered* 2013, **104**:371–379.
- Martin AP, Pardini AT, Noble LR, Jones CS: **Conservation of a dinucleotide simple sequence repeat locus in sharks.** *Mol Phyl Evol* 2002, **23**:205–213.
- Costa M, Fernandes C, Rodrigues M, Santos-Reis M, Bruford MW: **A panel of microsatellite markers for genetic studies of European polecats (*Mustela putorius*) and ferrets (*Mustela furo*).** *Eur J Wildl Res* 2012, **58**:629–633.
- Sekino M, Hara M: **Individual assignment tests proved genetic boundaries in a species complex of Pacific abalone (genus *Haliotis*).** *Conserv Genet* 2007, **8**:823–841.
- Dudgeon CL, Blower DC, Broderick D, Giles JL, Holmes BJ, Kashiwagi T, Krück NC, Morgan JAT, Tillett BJ, Ovenden JR: **A review of the application of molecular genetics for fisheries management and conservation of sharks and rays.** *J Fish Biol* 2012, **80**:1789–1843.
- Boomer JJ, Stow AJ: **Rapid isolation of the first set of polymorphic microsatellite loci from the Australian gummy shark, *Mustelus antarcticus* and their utility across divergent shark taxa.** *Conserv Genet Resour* 2010, **2**:393–395.
- Barbara T, Palma-Silva C, Paggi GM, Bered F, Fay MF, Lexer C: **Cross-species transfer of nuclear microsatellite markers: potential and limitations.** *Mol Ecol* 2007, **16**:3759–3767.
- Rico C, Rico I, Hewitt G: **470 million years of conservation of microsatellite loci among fish species.** *Proc R Soc Lond B* 1996, **263**:549–557.
- Primmer CR, Painter JN, Koskinen MT, Palo JU, Merilä J: **Factors affecting avian cross-species microsatellite amplification.** *J Avian Biol* 2005, **36**:348–360.
- Compagno LJV: **An overview of chondrichthyan systematics and biodiversity in Southern Africa.** *Trans R Soc S Afr* 2000, **54**:75–120.
- Department of Agriculture, Forestry and Fisheries (DAFF): **South Africa's National Plan of Action for the Conservation and Management of Sharks.** [[http://www.daff.gov.za/doiDev/topMenu/DRAFT\\_NPOA\\_SHARKS.pdf](http://www.daff.gov.za/doiDev/topMenu/DRAFT_NPOA_SHARKS.pdf)]
- Navigating Global Shark Conservation: **Current Measures and Gaps.** [[http://www.pewenvironment.org/uploadedFiles/PEG/Publications/Report/Navigating%20Global%20Shark%20Conservation\\_Current%20Measures%20and%20Gaps%207%206%2012.pdf](http://www.pewenvironment.org/uploadedFiles/PEG/Publications/Report/Navigating%20Global%20Shark%20Conservation_Current%20Measures%20and%20Gaps%207%206%2012.pdf)]
- Namibia's National Plan of Action to prevent, deter and eliminate illegal, unreported and unregulated fishing. [[http://209.88.21.36/opencms/export/sites/default/grnnet/MFMR/downloads/docs/Namibia\\_NPOA\\_IUU\\_Final.pdf](http://209.88.21.36/opencms/export/sites/default/grnnet/MFMR/downloads/docs/Namibia_NPOA_IUU_Final.pdf)]
- Pierce SJ, Trerup M, Williams C, Tilley A, Marshall AD, Raba N: **Shark fishing in Mozambique: A preliminary assessment of artisanal fisheries.** Maputo: Eyes on the Horizon; 2008:1–28.
- Karaiskou N, Buggiotti L, Leder E, Primmer CR: **High degree of transferability of 86 newly developed zebra finch EST-linked microsatellite markers in 8 bird species.** *J Hered* 2008, **99**:688–693.
- FitzSimmons NN, Moritz C, Moore S: **Conservation and dynamics of microsatellite loci over 300 million years of marine turtle evolution.** *Mol Biol Evol* 1995, **12**:432–440.
- Primmer CR, Møller AP, Ellegren H: **A wide-range survey of cross-species microsatellite amplification in birds.** *Mol Ecol* 1996, **5**:365–378.
- Kang J, Park J, Jo H: **Rapid development of microsatellite markers with 454 pyrosequencing in a vulnerable fish, the mottled skate, *Raja pulchra*.** *Int J Mol Sci* 2012, **13**:7199–7211.



45. Griffiths AM, Casane D, McHugh M, Wearmouth VJ, Sims DW, Genner MJ: **Characterisation of polymorphic microsatellite loci in the small-spotted catshark (*Scyliorhinus canicula* L.).** *Conserv Genet Resour* 2011, **3**:705–709.
46. Zane L, Bargelloni L, Patarnello T: **Strategies for microsatellite isolation: a review.** *Mol Ecol* 2002, **11**:1–16.
47. Primmer CR, Ellegren H: **Patterns of molecular evolution in avian microsatellites.** *Mol Biol Evol* 1998, **15**:997–1008.
48. Angers B, Bernatchez L: **Complex evolution of a salmonid microsatellite locus and its consequences in inferring allelic divergence from size information.** *Mol Biol Evol* 1997, **14**:230–238.
49. Giresi M, Renshaw MA, Portnoy DS, Gold JR: **Isolation and characterization of microsatellite markers for the dusky smoothhound shark, *Mustelus canis*.** *Conserv Genet Resour* 2012, **4**:101–104.
50. Chabot C, Nigenda S: **Characterization of 13 microsatellite loci for the tope shark, *Galeorhinus galeus*, discovered with next generation sequencing and their utility for eastern Pacific smooth-hound sharks (*Mustelus*).** *Conserv Genet Resour* 2011, **3**:553–555.
51. Castoe TA, Poole AW, Gu W, Jason De Koning AP, Daza JM, Smith EN, Pollock DD: **Rapid identification of thousands of copperhead snake (*Agkistrodon contortrix*) microsatellite loci from modest amounts of 454 shotgun genome sequence.** *Mol Ecol Res* 2010, **10**:341–347.
52. Dawson DA, Horsburgh GJ, Küpper C, Stewart IR, Ball AD, Durrant KL, Burke T: **New methods to identify conserved microsatellite loci and develop primer sets of high cross-species utility—as demonstrated for birds.** *Mol Ecol Res* 2010, **10**:475–494.
53. Hoffman JI, Nichols HJ: **A novel approach for mining polymorphic microsatellite markers *in silico*.** *PLoS One* 2011, **6**:e23283.
54. Human BA: **A taxonomic revision of the catshark genus *Haploblepharus* Garman 1913 (Chondrichthyes: Carcharhiniformes: Scyliorhinidae).** *Zootaxa* 2007, **1451**:1–40.
55. Saghai-Marouf MA, Solima KM, Jorgenson RA, Allard RW: **Ribosomal DNA spacerlength polymorphisms in barley: Mendelian inheritance, chromosomal location, and population dynamics.** *Proc Natl Acad Sci U S A* 1984, **81**:8014–8018.
56. Chabot CL: **Characterization of 11 microsatellite loci for the brown smooth-hound shark, *Mustelus henlei*, discovered with next-generation sequencing.** *Conserv Genet Resour* 2012, **4**:23–25.
57. Byrne RJ, Avis JC: **Genetic mating system of the brown smoothhound shark (*Mustelus henlei*), including a literature review of multiple paternity in other elasmobranch species.** *Mar Biol* 2012, **159**:749–756.
58. Guichoux E, Lagache L, Wagner S, Chaumeil P, Léger P, Lepais O, Lepoittevin C, Malaus T, Revardel E, Salin F, Petit RJ: **Current trends in microsatellite genotyping.** *Mol Ecol Resour* 2011, **11**:591–611.
59. Van Oosterhout C, Hutchinson WF, Wills DPM, Shipley P: **MICROCHECKER: software for identifying and correcting genotyping errors in microsatellite data.** *Mol Ecol Notes* 2004, **4**:535–538.
60. Rousset F: **GENEPOP'007: a complete re-implementation of the GENEPOP software for Windows and Linux.** *Mol Ecol Resour* 2008, **8**:103–106.
61. Slatkin M: **An exact test for neutrality based on the Ewens sampling distribution.** *Genet Res* 1994, **64**:71–74.
62. Excoffier L, Laval G, Schneider S: **Arlequin ver. 3.0: an integrated software package for population genetics data analysis.** *Evol Bioinform Online* 2005, **1**:47–50.
63. Peakall R, Smouse PE: **GenAEx 6.5: genetic analysis in Excel: population genetic software for teaching and research – an update.** *Bioinformatics* 2012, **28**:2537–2539.
64. Park S: **The Excel microsatellite toolkit.** [<http://animalgenomics.ucd.ie/sdepar/mstoolkit>]
65. Botstein D, White RL, Skolnick M, Davis RW: **Construction of a genetic linkage map in man using restriction fragment length polymorphisms.** *Am J Hum Genet* 1980, **32**:314–331.
66. Tamura K, Peterson D, Peterson N, Stecher G, Nei M, Kumar S: **MEGA5: molecular evolutionary genetics analysis using maximum likelihood, evolutionary distance, and maximum parsimony methods.** *Mol Biol Evol* 2011, **28**:2731–2739.

doi:10.1186/1756-0500-7-352

**Cite this article as:** Maduna et al.: Microsatellite cross-species amplification and utility in southern African elasmobranchs: A valuable resource for fisheries management and conservation. *BMC Research Notes* 2014 **7**:352.

**Submit your next manuscript to BioMed Central and take full advantage of:**

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at  
[www.biomedcentral.com/submit](http://www.biomedcentral.com/submit)

